Primary fatty amides in plasma associated with brain amyloid burden, hippocampal volume, and memory in the European Medical Information Framework for Alzheimer’s Disease biomarker discovery cohort

Min Kim a,b,1,2, Stuart Snowden a,c,d,1, Tommi Suvitaival b, Ashfaq Ali b, David J. Merkler e, Tahmina Ahmad a, Sarah Westwood f, Alison Baird i, Petroula Proitsi g, Alejo Nevado-Holgado f, Abdul Hye b, Isabelle Bos h, Stephanie Vos h, Rik Vandenberghe i, Charlotte Teunissen l, Mara ten Kate i, Philip Scheltens k, Silvy Gabel i, Karen Meersmans i,1, Olivier Blin m, Jill Richardson n, Ellen De Roeck o, Kristel Sleegers o, p, Régis Bordet q, Lorena Rami r, Petronella Kettunen s, t, u, Magda Tsalaki v, Frans Verhey h, Isabel Sala w, Alberto Lleó w, Gwendoline Peyratout x, Mikel Tainta y, Peter Johannsen z, Yvonne Freund-Levi aa, bb, Lutz Frölich cc, Valerija Dobricic dd, Sebastiaan Engelborghs oo, dd, ee, Giovanni B. Frisoni ff, José L. Molinuevo gg, Anders Wallin i, Julius Popp x, hh, Pablo Martinez-Lage y, Lars Bertram ii, Frederik Barkho f, Nicholas Ashton b, s, Kaj Blennow s, jj, Henrik Zetterberg s, jj, kk, ll, Johannes Streffer mm, Pieter J. Visser hj, Simon Lovestone f, Cristina Legido-Quigley a,b, g

aInstitute of Pharmaceutical Science, King’s College London, London, UK
bSteno Diabetes Center Copenhagen, Gentofte, Denmark
cInstitute of Metabolic Science, University of Cambridge, Cambridge, UK
dDepartment of Biochemistry, University of Cambridge, Cambridge, UK
eDepartment of Chemistry, University of South Florida, Tampa, FL, USA
fDepartment of Psychiatry, University of Oxford, Oxford, UK

1Institute of Psychiatry, Psychology and Neuroscience, Maurice Wohl Clinical Neuroscience Institute, King’s College London, London, UK
2Alzheimer Centrum Limburg, Maastricht University, Maastricht, The Netherlands
3University Hospital Leuven, Leuven, Belgium

4Neurochemistry Laboratory and Biobank, Department of Clinical Chemistry, Amsterdam University Medical Centers, Amsterdam Neuroscience, Amsterdam, The Netherlands
5Department of Neurology, Alzheimer Center, Amsterdam University Medical Centers, Amsterdam, The Netherlands
6Laboratory for Cognitive Neurology, Department of Neurosciences, KU Leuven, Leuven, Belgium
7Service de Pharmacologie Clinique et Pharmacovigilance, Institut de Neurosciences des Systèmes, Aix Marseille University, APHM, INSERM, Marseille, France

8Neurosciences Therapeutic Area, GlaxoSmithKline R&D, Stevenage, UK

Conflict of interest: S.E. reports research funding from Janssen Pharmaceutica N.V. and ADx Neurosciences (paid to institution). A.N.-H. holds a funding award from Ono Pharmaceutical Co Ltd. L.F. has participated in advisory boards for Allergan, Eli Lilly, Avraham Pharmaceuticals, Axon Neuroscience, Axovant, Biogen, Boehringer Ingelheim, Eisai, Functional Neuromodulation, Lundbeck, MerckSharpe & Dohme, Novartis, Pfizer, Pharmnext, Roche, Schwan and has received research grants from Novartis. H.Z. has served as advisory boards for Eli Lilly, Roche Diagnostics, Wave, Samumed and CogRx, has received travel support from Teva, and is a cofounder of Brain Biomarker Solutions in Gothenburg AB, a GU Ventures-based platform company at the University of Gothenburg (all un-related to this study). F.B. has served at advisory boards for Bayer-Schering Pharma, Biogen, Teva, Merck-Serono, Novartis, Roche, Jansen Research, Genzyme-Sanoﬁ, IXICO, GeNeuro, Apitope. All other authors report no biomedical financial interests or potential conflicts of interest.

1Both authors contributed equally to the study.
2Current address: Steno Diabetes Center Copenhagen, Gentofte, Denmark.
3Corresponding author. Tel.: +45-30913083.
4E-mail address: cristina.legido.quigley@regionh.dk

https://doi.org/10.1016/j.jalz.2019.03.004
1552-5260/Crown Copyright © 2019 Published by Elsevier Inc. on behalf of the Alzheimer’s Association. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
Abstract

Introduction: A critical and as-yet unmet need in Alzheimer’s disease (AD) is the discovery of peripheral small molecule biomarkers. Given that brain pathology precedes clinical symptom onset, we set out to test whether metabolites in blood associated with pathology as indexed by cerebrospinal fluid (CSF) AD biomarkers.

Methods: This study analyzed 593 plasma samples selected from the European Medical Information Framework for Alzheimer’s Disease Multimodal Biomarker Discovery study, of individuals who were cognitively healthy (n = 242), had mild cognitive impairment (n = 236), or had AD-type dementia (n = 115). Logistic regressions were carried out between plasma metabolites (n = 883) and CSF markers, magnetic resonance imaging, cognition, and clinical diagnosis.

Results: Eight metabolites were associated with amyloid β and one with t-tau in CSF, these were primary fatty acid amides (PFAMs), lipokines, and amino acids. From these, PFAMs, glutamate, and aspartate also associated with hippocampal volume and memory.

Discussion: PFAMs have been found increased and associated with amyloid β burden in CSF and clinical measures.

Keywords: EMIF-AD; Alzheimer’s disease; Dementia; Amyloid; Tau; CSF; Brain volume measurements; Cognitive function measurements; Metabolomics; Biomarkers

1. Background

Neurodegenerative dementias are characterized by a progressive decline in cognitive function and memory performance. Alzheimer’s disease (AD) is the most common of the neurodegenerative dementias making it a major source of global morbidity and mortality [1]. The World Alzheimer’s Report has estimated that there are more than 46 million people diagnosed with AD-type dementia and with an aging world population this figure is expected to increase to more than 130 million by 2050 [2]. In addition to a major human cost, AD also poses a significant economic cost estimated to increase to $1 trillion by 2018 [2].

Current clinical diagnosis of AD-type dementia relies on experienced clinicians using a battery of cognitive tests combined with various structural and functional imaging and cerebrospinal fluid (CSF) biomarkers to inform a judgment-based decision, with a definitive AD-type dementia diagnosis only possible at postmortem. Histologic examination of brain tissue during autopsy should contain significant evidence of extracellular amyloid β (Aβ) plaques and intracellular neurofibrillary tangles of hyperphosphorylated tau.

The deposition of Aβ plaques has been shown to start up to 20 years before the onset of symptoms [3–5]. There have been numerous drug candidates that have failed clinical
trials in symptomatic patients, these have been unsuccessful in producing a reversal of symptoms or a slowing of the progression of the disease [6]. It is thought that one of the reasons for the failure of these candidates is that they were not administered during the preclinical phase of the disease. This introduces the challenge of diagnosing people during the preclinical phase of the disease, when they are cognitively normal. For this to be possible, it is necessary to discover biomarkers that can identify individuals at high risk of developing clinical AD.

Metabolomics is the study of the complete complement of all low molecular weight metabolites (<1500 atomic mass units, Da) [7,8]. In essence, the metabolome represents metabolism in real time, the interaction of both genomic and environmental exchanges. To date there have been numerous AD metabolomic studies performed with different metabolomic platforms (i.e., different metabolome coverage and measurements) that aimed to identify panels of blood biomarkers in AD [9–13]. A handful of studies have included subjects with mild cognition problems who went on to develop AD during follow-up to find early disease biomarkers. These studies have uncovered metabolite panels with potential that are awaiting validation [14,15].

Here, we aimed to identify blood metabolites associating with CSF measures of amyloid and tau (phosphorylated and total). The abundance of metabolites was measured using liquid chromatography–mass spectrometry to cover ca. 800 metabolites. The selected metabolites were then compared with clinical cognition measures and rate of cognition decline, brain volumes, and diagnosis.

2. Methods

2.1. Subjects

This study used plasma samples from European Medical Information Framework for Alzheimer’s Disease (EMIF-AD) Multimodal Biomarker Discovery study [16]. EMIF-AD Multimodal Biomarker Discovery is a cross-cohort study consisting of collated data from 11 European cohorts that aims to discover novel diagnostic and prognostic markers for AD-type dementia by performing analyses in multiple biomarker modalities. More details on EMIF-AD participants can be found in Section 1 of Supplementary methods and in Bos et al. [16].

2.2. Clinical and cognitive data

In the present study, the 593 plasma samples were from 242 normal cognition (NC), 236 mild cognitive impairment (MCI), and 115 AD patients at sampling (Table 1). Of 236 MCI participants, 83 were later diagnosed with AD-type dementia (defined as AD converting MCI [cMCI]), whereas 78 remained as MCI (defined as stable MCI [sMCI]). The average follow-up length was 2.49 years.

The Mini-Mental State Examination (MMSE) score and the rate of cognitive decline (ROD) were available for 590

Table 1

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Sample size</th>
<th>NC</th>
<th>MCI</th>
<th>AD</th>
<th>Difference</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Size</td>
<td>593</td>
<td>242</td>
<td>236</td>
<td>115</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>Gender (M/F)</td>
<td>281/312</td>
<td>50/65</td>
<td>116/120</td>
<td>115/127</td>
<td>6.06 × 10^−01, y</td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>593</td>
<td>65.06 (7.93)</td>
<td>70.44 (7.86)</td>
<td>69.55 (8.51)</td>
<td>5.61 × 10^−04</td>
<td></td>
</tr>
<tr>
<td>MMSE (−/+−)</td>
<td>306/281</td>
<td>103/137</td>
<td>131/101</td>
<td>72/43</td>
<td>6.93 × 10^−13, y</td>
<td></td>
</tr>
<tr>
<td>Aβ (−/−/−)</td>
<td>590</td>
<td>28.80 (1.13)</td>
<td>26.50 (2.82)</td>
<td>21.07 (4.87)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>t-tau (−/−/−)</td>
<td>538</td>
<td>0.10 (0.90)</td>
<td>0.84 (1.35)</td>
<td>1.28 (1.67)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>p-tau (−/−/−)</td>
<td>538</td>
<td>0.06 (0.83)</td>
<td>0.90 (1.22)</td>
<td>1.73 (1.63)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>MMSE</td>
<td>590</td>
<td>28.80 (1.13)</td>
<td>25.60 (2.82)</td>
<td>21.07 (4.87)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>ROD (−/−/−)</td>
<td>405</td>
<td>0.86 (0.52)</td>
<td>0.22 (0.75)</td>
<td>−1.07 (1.07)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Attention (−/−/−)</td>
<td>563</td>
<td>0.24 (1.13)</td>
<td>−0.89 (1.58)</td>
<td>−1.77 (1.96)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Executive (−/−/−)</td>
<td>343</td>
<td>0.20 (1.10)</td>
<td>−0.81 (1.90)</td>
<td>−2.46 (2.07)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Language (−/−/−)</td>
<td>572</td>
<td>−0.14 (1.01)</td>
<td>−0.98 (1.26)</td>
<td>−2.13 (1.34)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Memory delayed (−/−/−)</td>
<td>463</td>
<td>0.07 (1.15)</td>
<td>−1.26 (1.18)</td>
<td>−2.29 (1.00)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Memory immediate (−/−/−)</td>
<td>537</td>
<td>−0.47 (1.88)</td>
<td>−1.43 (1.29)</td>
<td>−2.24 (1.29)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Visuosconstruction (−/−/−)</td>
<td>346</td>
<td>0.18 (1.20)</td>
<td>0.13 (1.43)</td>
<td>−1.19 (2.20)</td>
<td>1.41 × 10^−08</td>
<td></td>
</tr>
<tr>
<td>Hippocampal left</td>
<td>387</td>
<td>4411.38 (441.14)</td>
<td>3294.23 (634.68)</td>
<td>3042.18 (463.92)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Hippocampal right</td>
<td>387</td>
<td>3868.75 (429.20)</td>
<td>3413.52 (628.90)</td>
<td>3197.99 (496.24)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Hippocampal sum</td>
<td>387</td>
<td>7626.65 (837.45)</td>
<td>6707.78 (1213.79)</td>
<td>6242.20 (884.14)</td>
<td>&lt;2.00 × 10^−16</td>
<td></td>
</tr>
<tr>
<td>Cortical thickness in whole brain</td>
<td>351</td>
<td>2.29 (0.12)</td>
<td>2.30 (0.11)</td>
<td>2.28 (0.11)</td>
<td>5.13 × 10^−01, y</td>
<td></td>
</tr>
<tr>
<td>Cortical thickness in AD regions</td>
<td>351</td>
<td>2.65 (0.17)</td>
<td>2.64 (0.16)</td>
<td>2.57 (0.17)</td>
<td>7.12 × 10^−03, y</td>
<td></td>
</tr>
<tr>
<td>Taking AChEI, yes/no</td>
<td>210</td>
<td>1/40</td>
<td>51/66</td>
<td>28/24</td>
<td>4.90 × 10^−07, y</td>
<td></td>
</tr>
<tr>
<td>Taking other AD medications, yes/no</td>
<td>210</td>
<td>0/41</td>
<td>16/101</td>
<td>7/45</td>
<td>4.27 × 10^−02, y</td>
<td></td>
</tr>
</tbody>
</table>

NOTE. Results are mean (standard deviation) for continuous variables.
Abbreviations: Aβ, amyloid β; AChEI, acetylcholine esterase inhibitor; AD, Alzheimer’s disease; APOE4, apolipoproteinE e4; MCI, mild cognitive impairment; MMSE, Mini-Mental State Examination; NC, normal cognition; ROD, rate of cognitive decline.

*x2 test.
One-way analysis of variance test.
and 405 participants, respectively (n = 405). Neuropsychological tests measuring five different cognitive domains were also available: memory (delayed, n = 452 and immediate, n = 537), language (n = 572), attention (n = 543), executive functioning (n = 434), and visuoconstruction (n = 346). More details on how ROD and neuropsychological test (z score) values were collected can be found in Bos et al. [16] and in Section 2 of Supplementary methods. The clinical design is explained in detail in Bos et al. [16,17]. Details on the amyloid and tau level measurements and magnetic resonance imaging (MRI) and genetic analyses can be found in Section 3 of Supplementary methods.

2.3. Metabolomics data acquisition and treatment

Relative levels of 883 plasma metabolites were measured in fasting blood samples using three different mass spectrometry methods. Details on the analytical method and data treatment can be found in Section 4 of Supplementary methods.

2.4. Statistical analyses

Before statistical analyses, baseline characteristics were compared between the diagnostic group using the \( \chi^2 \) test for categorical variables and the analysis of variance for continuous variables (Table 1). To investigate the association of each metabolite with AD clinical variables, regression models were applied, adjusting for age at sampling, gender, and presence of apolipoprotein E (APOE) ε4. Adjustment for multiple testing was applied using a Bonferroni correction \( P \text{ value} < 7.72 \times 10^{-5} (=0.05/648) \), where 648 is the number of metabolites. All associations are reported as the change per one metabolite standard deviation.

A schematic workflow of the primary data analysis used in this study can be found in Supplementary Fig. 1. More details on statistical analyses and the network analysis can be found in Section 5 of Supplementary methods. More details on statistical analyses can be found in Section 5 of Supplementary methods. Finally, a data-driven network analysis was performed post hoc to identify independent associations between all pairs of variables. Missing values were imputed with the k-nearest neighbor algorithm (impute package for R [18]) after standardizing to zero-mean and unit-variance. To infer the partial correlation network between variables, graphical least absolute shrinkage and selection operating algorithm was used [19]. The optimal complexity network was selected with extended Bayesian information criterion (huge package for R [20]) and visualized with qgraph package for R [21]. All statistical analyses were performed using R Statistical Software (version 3.4.1).

3. Results

3.1. Demographic and cognitive comparisons

The current data set comprises 593 participants divided in three diagnostic groups: NC (n = 242), MCI (n = 236), and AD-type dementia (n = 115). Demographic and clinical data are presented in Table 1. There were no differences in gender between the three diagnostic groups. The MCI and AD participants were older when compared with NC participants. AD participants were more frequently APOEε4 carriers and presented higher CSF \( A\beta \), p-tau, and t-tau z score levels (all, \( P < .01 \)). The z scores for \( A\beta \) have inverted positive values, this means lower CSF \( A\beta_{42} \) and CSF \( A\beta_{42/40} \) ratio. All cognitive tests showed values that were lower in AD participants when compared with MCI and NC participants (all, \( P < .01 \)). Brain volume measurements by MRI analyses showed lower hippocampus volumes (left, right, and sum) and average cortical thickness in AD signature regions in AD participants (all, \( P < .01 \)). No differences could be observed between the three diagnostic groups for average cortical thickness across the whole brain (\( P > .05 \)).

3.2. Association of blood metabolites with measures of amyloid and tau in CSF

First, we aimed to identify metabolites that would associate with AD pathologic features, namely \( A\beta \), p-tau and t-tau z scores. For this, we built linear regression models between 648 metabolite levels and (1) \( A\beta \) levels in participants in all three diagnostic groups (NC + MCI + AD), (2) \( A\beta \) in MCI participants, (3) p-tau in participants in all three diagnostic groups (NC + MCI + AD), (4) p-tau in MCI participants, (5) t-tau in participants in all three diagnostic groups (NC + MCI + AD), and (6) t-tau in MCI participants. Associations in MCI participants only were performed to identify metabolites associating with \( A\beta \) and tau changes in the early stages of AD.

After adjusting for age at sampling, gender, and APOEε4, five metabolites were found to associate with \( A\beta \) levels across all participants at \( P < 7.72 \times 10^{-5} \). These five metabolites were palmitamide, oleamide, linoleamide, stearamide, and aspartate. Palmitamide, oleamide, linoleamide, and stearamide are primary fatty acid amides (PFAMs) and they were increased with higher \( A\beta \) levels (\( \beta = 0.21 \), \( \beta = 0.19 \), \( \beta = 0.18 \), and \( \beta = 0.16 \)) whereas aspartate was decreased with \( A\beta \) levels (\( \beta = -0.18 \)). The \( P \) value for each association can be found in Table 2. In addition, a regression model in a subset (N = 467) investigated metabolite associations with CSF \( A\beta_{42/40} \) values (Supplementary Fig. 2). Palmitamide, oleamide, aspartate, linoleamide, stearamide, aspartate, and glutamate associated with CSF \( A\beta_{42/40} \) at \( P < 7.72 \times 10^{-5} \) level.

Next, we investigated metabolites associating with \( A\beta \) levels in MCI participants. Linear regression models were built for all 648 metabolites, eight metabolites were found to associate with \( A\beta \) at \( P < 7.72 \times 10^{-5} \) level. The eight metabolites included the five previous metabolites (palmitamide, oleamide, aspartate, linoleamide, stearamide, and aspartate) and 9,10-DiHOME, 12,13-DiHOME, and glutamate. Palmitamide (\( \beta = 0.35 \)), oleamide (\( \beta = 0.35 \)), linoleamide (\( \beta = 0.27 \)), and stearamide (\( \beta = 0.33 \)) levels were found
### Table 2

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Beta</th>
<th>P-value</th>
<th>T-z score in all Aβ</th>
<th>P-value</th>
<th>T-z score in MCI group</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>12,13-DiHOME</td>
<td>-0.09</td>
<td>0.94</td>
<td>-0.18</td>
<td>0.80</td>
<td>-0.27</td>
<td>0.59</td>
</tr>
<tr>
<td>9,10-DiHOME</td>
<td>0.04</td>
<td>0.63</td>
<td>0.08</td>
<td>0.50</td>
<td>0.01</td>
<td>0.90</td>
</tr>
<tr>
<td>Argininate</td>
<td>0.03</td>
<td>0.76</td>
<td>0.05</td>
<td>0.70</td>
<td>0.00</td>
<td>0.99</td>
</tr>
<tr>
<td>Stearamide</td>
<td>0.02</td>
<td>0.88</td>
<td>0.01</td>
<td>0.98</td>
<td>0.00</td>
<td>0.99</td>
</tr>
</tbody>
</table>

NOTE: Positive beta value denotes that higher metabolite levels associate with lower CSF amyloid and higher cerebrospinal fluid p-tau and t-tau.

Abbreviations: Aβ, amyloid; AD, Alzheimer’s disease; MCI, mild cognitive impairment.

To be increased, whereas aspartate ($\beta = -0.31$), 9,10-DiHOME ($\beta = -0.27$), 12,13-DiHOME ($\beta = -0.28$), and glutamate ($\beta = -0.28$) levels were found to be lower with $A\beta$ levels ($P$ values can be found in Table 2).

We then examined the association of each of the 648 metabolites with the $z$ score of CSF p-tau in all participants ($N = 538$) and in MCI participants ($N = 235$). At $P < 7.72 \times 10^{-5}$ level, neither model showed metabolites associating with p-tau. Furthermore, we investigated association between 648 metabolites and CSF t-tau levels in all participants ($N = 538$) and in MCI participants ($N = 235$). Argininate was found to associate with CSF t-tau levels in all participants at $P < 7.72 \times 10^{-5}$, its level was found decreased with higher CSF t-tau levels ($\beta < -0.22$). No metabolites associated with CSF t-tau in MCI participants only. Next, we tested if these nine metabolites were associated to any of the drugs detected ($n = 2$, hydroquinone sulfate and salicylate), and AD medications ($n = 2$, acetyl-cholinesterase inhibitors or other AD drugs). Regression models showed no associations between the nine metabolites and drugs at $P < 7.72 \times 10^{-5}$ (data not shown).

Overall, we investigated associations between 648 metabolites and three well-established AD pathologic markers, $A\beta$, p-tau, and t-tau in all participants (NC + MCI + AD) and in MCI participants only. From these models, we were able to select nine metabolites, palmitamide, oleamide, linoleamide, stearamide, 9,10-DiHOME, 12,13-DiHOME, glutamate, and aspartate, which were found to associate with $A\beta$ levels, whereas argininate was found to associate with t-tau levels.

Summary of the regression models between the nine metabolites and clinical $A\beta$ and t-tau levels can be seen in Fig. 1 and Table 2.

#### 3.3. Association of blood panel with cognition, ROD, and diagnosis

We next examined if any of these nine metabolites identified in the regression analyses would associate with AD cognitive and diagnosis variables.

First, including all the diagnosis groups we investigated the associations with MMSE scores and the ROD. Linear regression models showed six metabolites associating with MMSE at $P < 7.72 \times 10^{-5}$ level. Linoleamide ($\beta = -0.98$), oleamide ($\beta = -1.00$), and palmitamide ($\beta = -0.94$) levels were lower whereas argininate ($\beta = 0.84$), aspartate ($\beta = 1.04$), and glutamate ($\beta = 0.87$) levels were higher with higher MMSE scores. Only glutamate was found to associate with ROD ($\beta = -0.24$) at $P < 7.72 \times 10^{-5}$ level ($P$ values can be found in Supplementary Table 1).

Then the selected nine metabolites were examined against neuropsychological tests measuring five different cognitive domains. Lower attention levels were found to associate with higher levels of stearamide ($\beta = -0.38$), palmitamide ($\beta = -0.43$), oleamide ($\beta = -0.40$), and
linoleamide ($\beta = -0.40$) at $P < 7.72 \times 10^{-5}$ level. No metabolite associated with executive function. Lower language levels were found to associate with higher oleamide levels ($\beta = -0.25$) and lower levels of glutamate ($\beta = 0.2$), aspartate ($\beta = -0.25$), and argininate ($\beta = -0.28$) levels at $P < 7.72 \times 10^{-5}$ level ($P$ values can be found in Supplementary Table 2).

No metabolite was found to associate with delayed memory. Three metabolites associated with immediate memory, linoleamide ($\beta = -0.49$), oleamide ($\beta = -0.43$), and palmitamide ($\beta = -0.38$). Finally with visuoconstruction scores, no metabolites were found to associate with this function.

We were also interested on how the nine metabolites would associate with clinical diagnosis. Three metabolites associated with AD (vs. NC) at $P < 7.72 \times 10^{-5}$ level, lower levels of argininate (odds ratio [OR] = 0.50), aspartate (OR = 0.50), and glutamate (OR = 0.53) were found to associate with AD. Afterward, we combined cMCI and AD participants into one group as cMCI is often defined as early AD. We then measured cMCI and AD against NC as an outcome, six metabolites were found to be associated at $P < 7.72 \times 10^{-5}$ level. These six metabolites included argininate (OR = 0.49), aspartate (OR = 0.45), glutamate (OR = 0.50), linoleamide (OR = 2.34), oleamide (OR = 2.34), and palmitamide (OR = 2.20). We also looked at AD conversion from MCI participants (cMCI vs. sMCI) and found no association at $P < 7.72 \times 10^{-5}$ level (Supplementary Table 2). A summary of the regression models can be found in Supplementary Tables 1 and 2.

3.4. Associations of blood panel with brain structural measures

In the present study, MRI data were available for 387 participants (hippocampus volumes left, right, and sum) and 351 participants had cortical thickness measures (average across the whole brain and in AD signature regions).

Of the nine metabolites, four metabolites were found to associate with right hippocampus volume at $P < 7.72 \times 10^{-5}$ level. These included higher level of glutamate ($\beta = 146.10$) and lower levels of linoleamide ($\beta = -154.91$), oleamide ($\beta = -145.33$), and palmitamide ($\beta = -132.32$). When sum of hippocampus volume was examined as an outcome, higher level of glutamate ($\beta = 273.10$) and lower levels of linoleamide ($\beta = -274.93$), oleamide ($\beta = -268.55$), and palmitamide ($\beta = -246.38$) were found to associate with higher sum of hippocampus volume ($P < 7.72 \times 10^{-5}$ level). Linear regression analysis with left hippocampus volume or cortical thickness as an outcome showed no metabolite association (Supplementary Table 3).

Summary of all the regression models between the nine metabolites and all AD clinical variables can be found in Fig. 2A as a circos plot depicting 25 classes of metabolites and nine metabolites associated with amyloid or tau (Supplementary Fig. 3 as a heat map). Fig. 2B shows relative levels of plasma oleamide versus amyloid levels, Supplementary Fig. 4, relative levels of plasma oleamide versus diagnostic groups.

3.5. Predictive models of clinical diagnosis

We built receiver operating characteristic (ROC) models for the nine selected metabolites; Fig. 3A depicts how well the panel can discriminate AD-type dementia from control subjects. The resulting area under the curve (AUC) value was 0.7811. We then examined how well other AD clinical variables would discriminate diagnoses groups. Aβ produced AUC value of 0.7814, t-tau produced AUC of 0.7815, p-tau produced AUC of 0.8541, and covariates (known risks of age at sampling and APOEε4 status) produced AUC of 0.7035 (Fig. 3A).

Then we built ROC model (Fig. 3B) for the nine selected metabolites for AD cMCI versus non-cMCI groups. The resulting AUC value was 0.6625. This value was compared with other AD clinical variables, Aβ (AUC = 0.6065), p-tau (AUC = 0.6623), t-tau (AUC = 0.6642), and covariates (AUC = 0.5718).

3.6. Network analysis

To visualize the interactions in the full data set, including metabolites and all clinical variables, a data-driven network was computed selecting the clinical variables and
metabolites that are immediately connected. PFAMs, glutamine, and aspartate were strongly correlated to amyloid. There were two additional metabolites that showed high partial correlation to MMSE and diagnosis, these were tryptophan, betaine, and 2-pyrrolidinone, respectively (see Supplementary Fig. 5).

4. Discussion

At present clinical MCI and AD-type dementia diagnoses are made based on the presentation of symptoms, cognitive assessments, biomarker analyses, and the judgment of the clinician. However, by the time symptoms are present...
irreversible changes have occurred in the structure of the brain. Hence the ideal plasma biomarker or panel would give information about high amyloid levels and risk of conversion to AD to treat at an earlier stage. In fact very few biomarkers have been approved to this end, the positron-emission tomography tracer florbetapir was approved by the Food and Drug Administration in 2012 and it allows for a measurement of amyloid burden in the brain, achieving sensitivity of 82% to 92% and specificity of 95% [22]. CSF measures of amyloid and tau are also widely accepted as biomarkers of AD. A recent meta-analysis of CSF biomarkers (12 cohorts cMCI = 307 and sMCI = 501), using CSF amyloid-42, p-tau, and t-tau, conversion was predicted with average ratios of 0.67, 1.72, and 1.76, respectively [23].

In this study, 883 structurally distinct compounds were measured in 593 plasma samples. We found that metabolites pertaining to three classes of compounds were associated with amyloid or tau measures, namely four PFAMs, two lipid hormones, and three amino acids. The panel predicted AD-type dementia with the same AUC as amyloid (AUC = 0.78 for both) but lower than t-tau for this cohort (AUC = 0.85). For early diagnoses, in the converter group the prediction of the panel was AUC = 0.66, higher than amyloid by itself (AUC = 0.60) but the same as t-tau and p-tau in this cohort.

4.1. PFAMs and AD

In this study, all four PFAMs measured in our analyses were increased with amyloid burden. Oleamide is the most studied and best understood PFAM. Oleamide was first identified in brains of sleep-deprived cats and mice [24] and is an important regulatory lipid in the brain and central nervous system. Oleamide regulates the sleep-wake cycle, memory, locomotion, pain perception, and is anti-inflammatory, anxiolytic, and neuroprotective [25,26]. Administration of oleamide protects against scopolamine-induced cognitive impairment [27] and depression in a rat chronic stress model [28]. Earlier metabolomic studies have shown lower serum concentrations of oleamide in patients with AD and patients with MCI [29,30]. Differences between our results and those reported earlier could arise by using amyloid as the primary outcome.

If oleamide, an endocannabinoid, is neuroprotective and has the ability to induce sleep, it could be synthesized to improve amyloid clearance and restore the sleep-wake cycle disruption that is characteristic of AD [31]. Sleep deprivation and its disorders are suggested to precede and predict dementia [32–34]. During sleep the brain’s interstitial space increases in volume by up to 60% to enable CSF to clear neurotoxic waste into the systemic circulation [35]. It has been shown now that positron-emission tomography amyloid burden increases in the hippocampus with one-night sleep deprivation [36], this is especially interesting because of PFAMs associated with total hippocampal volume in our study. Moreover, it has been shown that early amyloid toxicity can be blocked by the activation of cannabinoid receptors [37].

In addition to oleamide, we find that the plasma levels of three other fatty acid amides, palmitamide, linoleamide, and stearamide, were also increased. Decreased serum palmitamide in AD was reported earlier [38]. In contrast to oleamide, far less is known about the biological function of palmitamide, linoleamide, and stearamide [26]. Alterations in the blood levels of the PFAMs have been reported for other diseases or disease models. Lower serum oleamide has also been found in a rat model for ischemic stroke [39] and in premutation carriers of the fragile X mental retardation 1 (FMR1) gene [40]. Treatment of rats with quinolinic acid increased the serum levels of pentadecanamide, palmitamide, oleamide, and stearamide [41], this is interesting as quinolinic acid is produced by activated microglia [42]. A metabolomic study of the systemic effects of cirrhosis identified increased serum levels of oleamide and stearamide [43].

Alterations in plasma PFAM levels in AD are likely resulted from dysfunction in synthesis, degradation, or transport. Little is known about PFAM transport; the major degradative reaction for the PFAMs is their hydrolysis into fatty acid and ammonia, a reaction catalyzed by fatty acid amid hydrolase (FAAH) [44] and/or N-acylethanolamine-hydrolyzing acid amidase [45]. There are conflicting reports regarding FAAH and AD, one report indicating that FAAH is overexpressed [46] and another that FAAH activity is decreased in AD [47]. An AD-related decrease in FAAH activity could lead to increased plasma levels of the PFAMs. In Fig. 4, the three main routes are explained, decreases in peptidylglycine α-amidating monoxygenase expression in cultured neuroblastoma cells yielded decreases in PFAM levels providing evidence in vivo [48]. One report indicated that peptidylglycine α-amidating monoxygenase activity is lower than normal in the brain and CSF of patients with AD [49].

We also found two lipokines that were decreased in MCI participants with higher amyloid burden in the brain. Although there are no brain studies in which lipokines were detected, in blood, these were negatively correlated with both body mass index and insulin resistance in the obese, and in another study increased after exercise [54,55]. These findings would point out toward a metabolic explanation that could be investigated in future studies.

The amount of glutamate and aspartate was decreased in plasma and associated with amyloid whereas arginine correlated with t-tau. We and others have found in brain that excitatory neurotransmitters glutamate and aspartate were decreased with increasing amyloid and tau burden [56], whereas plasma arginine was not found to correlate with CSF p-tau [57], but it has been consistently found to be decreased in the brain of patients with AD [58,59].

A limitation of this study is that although it is a relatively large study for metabolomics, the number of participants is
still small. In addition, the amounts of PFAMs alone predicted AD-type dementia with an ROC AUC of 0.63 (data not shown), which is much lower than that predicted by the panel together with amino acids and lipokines.

In sum, our data points toward a panel of metabolites including PFAMs, amino acids, and lipokines that could help understand AD pathways and achieve prediction in blood equivalent to that achieved by amyloid measures in CSF. PFAMs are endocannabinoids that could potentially have an anti-inflammatory and sleep-inducing role as amyloid burden increases toxicity in the brain.

Acknowledgments

The authors thank the individuals and families who took part in this research. The authors would also like to thank all people involved in data and sample collection and/or logistics across the different centers, and in particular Marije Benedictus, Wiesje van de Flier, Charlotte Teunissen, Ellen De Roeck, Naomi De Roeck, Ellis Niemantsverdriet, Charisse Somers, Babette Reijs, Andrea Izagirre Otaegi, Mirian, Ecay Torres, Sindre Rolstad, Eva Bringman, Domilé Tautvydaité, Barbara Moullet, Charlotte Evenepoel, Isabelle Cleynen, Bea Bosch, Daniel Alcolea Rodriguez, Moira Marízzoni, Alberto Redolfi, and Paolo Bosco.

Funding: The present study was conducted as part of the EMIF-AD project, which has received support from the Innovative Medicines Initiative Joint Undertaking under EMIF grant agreement no. 115372, resources of which are composed of financial contribution from the European Union’s Seventh Framework Program (FP7/2007-2013) and EFPIA companies’ in-kind contribution. The DESCRIPTA study was funded by the European Commission within the fifth framework program (QLRT-2001-2455). The EDAR study was funded by the European Commission within the fifth framework program (contract no. 37670). The San Sebastian GAP study is partially funded by the Department of Health of the Basque Government (allocation 17.0.1.08.12.0000.2.454.01.41142.001.H).

Fig. 4. PFAM biosynthesis. PFAM biosynthesis route is not clearly defined with possible routes including the direct conjugation of ammonia to a fatty acid as catalyzed by FAAH [50], a pathway that might be unfavorable in vivo [51]. The cytochrome c catalyzed reaction of ammonia with oleoyl-Coenzyme A to produce oleamide and coenzyme A was reported as one possible in vivo route for oleamide [52]. Peptidylglycine α-amidating monooxygenase–catalyzed cleavage of N-fatty acid glycines to the PFAMs has been proven in vivo [53]. Abbreviations: FAAH, fatty acid amide hydrolase; NAAA, N-acylethanolamine-hydrolyzing acid amidase.
D.J.M was partially supported by a grant from the National Institutes of Health (R15-GM107864).

Supplementary data

Supplementary data related to this article can be found at https://doi.org/10.1016/j.jalz.2019.03.004.

**RESEARCH IN CONTEXT**

1. Systematic review: The authors reviewed the literature using PubMed and reported key publications. Most studies were small and relatively heterogeneous in the metabolites identified. Most blood metabolomic studies have highlighted the role of lipid metabolites as important in Alzheimer’s disease (AD). However, we still need to identify peripheral early stage AD biomarkers that reflect pathology and/or that inform on its biochemistry and potential targets. For this, we used a comprehensive range metabolomic approach to identify small molecules in blood associating with pathology as indexed by cerebrospinal fluid AD biomarkers.

2. Interpretation: The results show that primary fatty acid amides (PFAMs) associated with AD pathology and phenotype. There are two important implications to this finding: first, the lipids in question are thought to be synthesized in the brain; and second, they are also thought to be natural endocannabinoids. With the exception of oleamide, the biology behind endogenous PFAMs is largely unknown; it is thought that they are synthesized to induce sedation. Neuropeptidases were also flagged and these could be depleted because of compromised synaptic signaling.

3. Future directions: Results of this study should be integrated with proteomics and genetics to find more about mechanisms involved in blood. Future studies should address whether PFAMs are causally related to AD, and if it is brain or other organs with the brain that are involved in the synthesis of PFAMs.

**References**


[24] Cravatt B, Prosergarcia O, Siuzdak G, Gilula N, Henriksen S, Crapo JD, et al. Toward defining the preclinical stages of Alzheimer’s disease: results of this study should be integrated with proteomics and genetics to find more about mechanisms involved in blood. Future studies should address whether PFAMs are causally related to AD, and if it is brain or other organs with the brain that are involved in the synthesis of PFAMs.


