The role of circulatory systemic environment in predicting interferon-alpha–induced depression: The neurogenic process as a potential mechanism

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\textbf{ABSTRACT}

Interferon (IFN)-\alpha treatment for hepatitis C virus (HCV) is a well-recognized clinical model for inflammation-induced depression, but the brain cellular mechanisms underlying these effects are still not clear. Previous data reported an alteration in peripheral levels of inflammatory and neuroplasticity markers in the blood of depressed versus non-depressed patients. We investigated the \textit{in vitro} effect of serum from depressed and non-depressed HCV patients (at baseline, before IFN-\alpha; and after four weeks of IFN-\alpha), on the apoptotic and neurogenic processes in a human hippocampal progenitor cells model. Results show that higher apoptosis during proliferation observed upon treatment of cells with baseline serum, and lower neuronal differentiation observed upon treatment with serum after 4 weeks of IFN-\alpha, were predictive of later development of IFN-\alpha–induced depression (odds ratio = 1.26, \(p = 0.06\), and 0.80, \(p = 0.01\), respectively). While serum after IFN-\alpha increased neurogenesis compared with baseline serum, a lower increase in neurogenesis was also predictive of later development of depression (odds ratio = 0.86; \(p = 0.006\)). Our results provide evidence for the fundamental role of the systemic milieu (captured by serum samples) in the regulation of hippocampal neurogenesis by inflammation, a putative mechanism involved in the development of neuropsychiatric conditions.

\textbf{1. Introduction}

Approximately 200,000 individuals have chronic hepatitis C virus (HCV) in the UK, and 170 million worldwide (Harris et al., 2012). The current standard treatment for HCV infection involves subcutaneous injection with pegylated-interferon-alpha (peg-IFN-\alpha), which clear the virus in up to 80% of patients while inducing a number of side effects (Wichers and Maes, 2002). Indeed, despite the efficacy of this treatment, IFN-\alpha promotes depression in up to 35% of patients within the first 8 to 12 weeks of the therapy (Asnis and De La Garza, 2006; Capuron and Miller, 2004; Machado et al., 2017), as well as other side effects, including symptoms of anxiety, mania and fatigue (Sockalingam and Abbey, 2009). The brain cellular mechanisms underlying this association have not been completely elucidated yet.

Our work as well as previous evidence have reported that peripheral IFN-\alpha acutely induces the production and release of several innate immune cytokine proteins (Hepgul et al., 2016; Raison et al., 2010). These cytokines are putatively involved in the depressogenic action of IFN-\alpha. Higher serum or plasma levels of pro-inflammatory markers have been shown to be associated with an increased risk of major depressive disorder (Dowlati et al., 2010). Another study from our group provided evidence for increased inflammation in the blood of depressed patients when compared with healthy controls, together with an association between higher cytokine levels and lack of antidepressant response (Cattaneo et al., 2010). However, it is still unclear how these factors contained in peripheral blood can affect the brain and ultimately predispose the individual to the development of depression.

Several pathways through which peripheral blood factors, including inflammatory molecules, can be transmitted from the periphery to the brain have been investigated (Louveau et al., 2015). Previous studies have demonstrated that distinct cytokines, including interleukin-1beta (IL-1\beta), IL-6, and tumor necrosis factor-alpha (TNF-\alpha), known to be upregulated both in the periphery and in the brain of patients with major depressive disorder (MDD), are able to increase blood brain barrier (BBB) permeability (Deli et al., 1995; Didier et al., 2003; Schwanger et al., 1999) and ultimately to cause changes in discrete brain regions, including the hippocampus (Wang et al., 2008). This region is highly involved in the process of neurogenesis, which in humans principally occurs in the subventricular zone (SVZ) of the lateral ventricles and the sub-granular zone (SGZ) of the dentate gyrus (DG) of
the hippocampus (Kempermann et al., 2018). Of relevance, previous research has demonstrated that peripheral administration of IFN-α or other cytokines, such as C-C motif chemokine ligand 11 (CCL11), in research has demonstrated that peripheral administration of IFN-α in the hippocampus (Kempermann et al., 2018). Of relevance, previous research has demonstrated that peripheral administration of IFN-α or other cytokines, such as C-C motif chemokine ligand 11 (CCL11), in research has demonstrated that peripheral administration of IFN-α in the hippocampus (Kempermann et al., 2018).

To investigate the contributing role of the systemic milieu in the regulation of hippocampal neurogenesis we used our previously established human hippocampal progenitor cell (HPC) line (Anacker et al., 2013a; Borsini et al., 2017b, 2018; Zunszain et al., 2012), and examined the effect of treatment with serum samples from patients with HCV, before and after 4 weeks of IFN-α, on the neurogenic process. In particular, we investigated whether changes in proliferation, neurogenesis and apoptosis, caused by treatment with serum samples, were predictive of later development of IFN-α-induced depression.

2. Method

2.1. Patients cohort

The sub-group of patients, from which serum samples derived, is part of a bigger cohort of patients recruited from the outpatient liver department of King’s College Hospital and St George’s Hospital. Eligible patients were adult patients with chronic HCV infection who were due to commence combination antiviral therapy with IFN-α and ribavirin without any additional direct-acting antiviral (boceprevir, telaprevir or simeprevir). The whole cohort of patients received combination therapy for at least 24 weeks. Patients were assessed at baseline when starting IFN-α treatment (TW0) and monthly for the first three months, at TW4, 8, 12, and 16 then two-monthly thereafter until the end of their treatment, at TW24. Combination therapy comprised of weekly subcutaneous IFN-α injections (1.5 µg per kg of body weight) and daily ribavirin tablets (800–1400 mg orally per day in 2 divided doses). Exclusion criteria for the larger cohort included, and for whom we had blood samples at both TW0 and TW4: a total of 33 eligible patients were included. The study was approved by the King’s College Hospital Research Ethics Committee (Ref: 10/H0808/30).

2.2. Demographics and clinical data of the IFN-α-treated HCV cohort

The Medical Research Council (MRC) Socio-demographic Schedule (Mallett et al., 2002) was administered at TW0 to collect socio-demographic data, including age and gender. The Mini-International Neuropsychiatric Interview (MINI) questionnaire (Sheehan et al., 1998) was administered at TW0, in order assess patients for a current depressive episode, as well as at follow-up assessments (TW4, 8, 12 and 24) to detect new onset cases of depression during IFN-α therapy. Demographic data shows that the selected cohort of 33 patients is predominantly characterised by male subjects (78.8%) with no significant difference in terms of age (Table 1). With respect to clinical data, 9 patients (27.3%) developed depression during IFN-α treatment, and 24 patients (72.7%) did not. Most importantly, the 9 patients developed depression between TW8 and TW24 (Table 2), that is, were not yet depressed at the time of the second (TW4) serum sample used for the present study.

2.3. Human hippocampal progenitor cells

For the in vitro experiments with serum from IFN-α-treated HCV patients, multipotent human hippocampal progenitor cell line HPCDA07/03C, kindly provided by ReNeuron Ltd (Surrey, UK) was used. Cells were grown as previously described in RMM consisting of Dulbecco’s Modified Eagle’s Media/F12 (DMEM:F12, Invitrogen, Paisley, UK) supplemented with 0.03% human albumin (Baxter Healthcare, Compton, UK), 100 µg/mL human apo-transferrin, 16.2 µg/mL human putrescine diHCl, 5 µg/mL human recombinant insulin, 60 ng/mL progesterone, 2 mM L-glutamine and 40 ng/mL sodium selenite (Anacker et al., 2013a, 2013b, 2011; Borsini et al., 2017a, 2018; Zunszain et al., 2012). To maintain proliferation, 10 ng/mL human basic fibroblast growth factor (bFGF), 20 ng/mL human epidermal growth factor (EGF) and 100 nM 4-hydroxymethoxifen (4-OHT) were added. Cells were grown in 75 cm² filtered cap culture flasks (Nuncloc, Roskilde, Denmark) at 37 °C in 5% CO2 and regularly passaged at 80% confluence, until being transferred to plates for the differentiation experiments.

2.4. Proliferation, neurogenesis and apoptosis

In order to assess proliferation and apoptosis (during the proliferation stage) progenitor cells were plated on 96-well plates (Nuncloc) at a density of 1.2 × 10⁴ cells per well per well in 100 µL RMM media. Cells were cultured in the presence of EGF, bFGF and 4-OHT for 24 h followed by 2 days incubation with 1% serum samples (the same concentration previously used in the lab as able to induce a profound effect without causing over-confluence or any alteration in the composition of neurons) from depressed or non-depressed IFN-α-treated HCV patients, also in the presence of growth factors containing 0.5 mg/mL penicillin streptomycin. The synthetic nucleotide bromodeoxyuridine (Brdu) (BS0002, Sigma) with a final concentration of 10 µM was added to the media 4hrs before the end of the incubation to label proliferating cells.

Whereas, in order to assess neuronal differentiation and apoptosis (during the differentiation stage) cells were plated on 96-well plates (Nuncloc) at a density of 1.2 × 10⁴ cells per well per well in 100 µL RMM media. Following the proliferation phase previously described, cells were washed twice for 15 min in RMM media (without EGF, bFGF and 4-OHT), and then cultured with 1% serum samples from depressed or non-depressed IFN-α-treated HCV patients in RMM containing 0.5 mg/mL penicillin streptomycin for subsequent 7 days.

| Table 1 | Characteristics of interferon-alpha (IFN-α) treated Hepatitis C (HCV) depressed vs. non-depressed patients assessed at baseline. |
|---|---|---|
| | Depressed (n = 9) | Non-depressed (n = 24) | p value |
| Age (years) | 41.3 ± 5.3 | 41.6 ± 2.5 | p = 0.9* |
| Gender Male | 8 (88.8%) | 18 (75%) | p = 0.4** |

* Student’s t-test. ** Chi squared test.

| Table 2 | Development of depression during interferon-alpha (IFN-α) treatment. |
|---|---|---|
| IFN-α-induced Depression | Total Number of Patients |
| No | Yes | |
| DEPRESSION WEEK | None | 24 | 0 | 24 |
| TW 8 | 0 | 5 | 5 |
| TW 12 | 0 | 2 | 2 |
| TW 16 | 0 | 1 | 1 |
| TW 24 | 0 | 1 | 1 |
| Total Number of Patients | 24 | 9 | 33 |
2.5. Immunocytochemistry

At the end of the total incubation time, 2 days (proliferation) and 9 days (2 days proliferation followed by 7 days differentiation), cells were fixed with 4% paraformaldehyde (PFA) for 20 min at room temperature. Cells were then washed three times in phosphate-buffered saline (PBS) and stored at 4°C in PBS in preparation for immunocytochemistry. Subsequently, PFA-fixed proliferating cells pulsed with BrdU were first incubated with 2 N hydrochloric acid for 25 min at room temperature followed by neutralisation with 0.1 M borate buffer for 10 min and 2 washes with PBS. Subsequently, both proliferating and differentiating cells were incubated in 50 µl of blocking solution constituted of 5% normal donkey serum (D9663, Sigma) in PBS containing 0.3% Triton X-100 (T8787, Sigma) for 1 h at room temperature. Primary antibodies for BrdU and Ki67 were used to assess proliferation (rat anti-BrdU, 1:500, Serotec; rabbit anti-Ki67, 1:500, Abcam). Antibodies for doublecortin (DCX) and microtubulin-associated protein-2 (MAP2) were used to assess neuroblasts and neurons respectively (rabbit anti-DCX, 1:500; mouse anti-MAP2, 1:500, Abcam), whereas antibody for caspase 3 (CC3) (rabbit anti-CC3, 1:500, Cell Signalling) were used to assess apoptosis and were diluted in blocking solution buffer. Finally, 30 µl/well were added and left at 4°C overnight. The next day, cells were washed, incubated in blocking solution for 30 min and then with fluorescently tagged secondary antibodies for BrdU (Alexa Fluor 488 donkey anti-rat, 1:1000), Ki67 and CC3 (proliferation) (Alexa Fluor 555 donkey anti-rabbit, 1:1000), DCX and CC3 (differentiation) (Alexa Fluor 488 donkey anti-rabbit, 1:1000), and MAP2 (Alexa Fluor 555 donkey anti-mouse, 1:1000; all from Life Technologies) at 30 µl/well for 2 h at room temperature. After 3 washes with PBS, cells were stained with 0.02 mg/ml DAPI at 50 µl/well for 5 min and washed 3 more times.

2.6. Automated quantification of immunofluorescence

An unbiased and automated approach was employed to quantify cell number, cell proliferation, differentiation and apoptosis using the

<table>
<thead>
<tr>
<th>IFN-α–Induced Depression</th>
<th>Odds Ratio</th>
<th>p value</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>TW0 BrdU</td>
<td>1.07</td>
<td>0.11</td>
<td>.08</td>
</tr>
<tr>
<td>TW0 Ki67</td>
<td>.97</td>
<td>0.44</td>
<td>.02</td>
</tr>
<tr>
<td>TW0 CC3</td>
<td>1.26</td>
<td>0.03</td>
<td>.15</td>
</tr>
<tr>
<td>TW0 CC3/BrdU</td>
<td>1.54</td>
<td>0.02</td>
<td>.2</td>
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</table>

Legend: treatment week (TW), bromodeoxyuridine (BrdU), caspase 3 (CC3). Analyses were controlled for age. Pseudo R² (Cox & Snell).

Fig. 1. Baseline markers of proliferation and apoptosis as predictors of interferon-alpha (IFN-α)–induced depression. Main predictive outcomes of baseline proliferating and apoptotic markers on later IFN-α–induced depression (a). Representative images of cells treated with baseline serum from depressed and non-depressed patients. Apoptotic cells were stained with caspase 3 (CC3) (red labelling), and apoptotic cells expressing the proliferating marker bromodeoxyuridine (BrdU) (green labelling) were identified as CC3/BrdU (b). Receiver Operating Characteristic (ROC) curves of baseline CC3 and CC3/BrdU predictors of IFN-α–induced depression (c). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)
3.1.1. Proliferation and apoptosis

We first assessed whether any changes in proliferating or apoptotic markers, detected upon treatment with TW0 serum samples, were able to predict later IFN-α-induced depression. When entering separately each marker in the model, results show no effect for any of those markers (Table 4).

3.2. Lower neurogenesis at TW4 predicts later IFN-α-induced depression

Having shown that baseline apoptotic markers are predictors of later IFN-α-induced depression, we subsequently investigated whether changes in the number of DCX+ cells between TW0 and TW4 were able to predict later IFN-α-induced depression. When entering separately each marker in the model, results show no effect for any of those markers (Table 4).

### Table 3

<table>
<thead>
<tr>
<th>IFN-α-induced Depression</th>
<th>Odds Ratio</th>
<th>p value</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>TW0 DCX</td>
<td>1.08</td>
<td>0.3</td>
<td>0.03</td>
</tr>
<tr>
<td>TW0 MAP2</td>
<td>1.08</td>
<td>0.4</td>
<td>0.02</td>
</tr>
<tr>
<td>TW0 CC3</td>
<td>1.03</td>
<td>0.6</td>
<td>0.006</td>
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<tr>
<td>TW0 CC3/MAP2</td>
<td>0.91</td>
<td>0.7</td>
<td>0.003</td>
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</tbody>
</table>

### Treatment week (TW), doublecortin (DCX), mitotubule associated protein 2. (MAP2), caspase 3 (CC3). Analyses were controlled for age. Pseudo R² (Cox & Snell).

81% accurate in predicting later development of depression (Fig. 1c).

3.2.1. Proliferation and apoptosis

As for data at baseline, we then investigated whether changes in proliferating (BrdU and Ki67) or apoptotic (CC3) markers detected upon treatment with TW4 serum samples, were able to predict later IFN-α-induced depression. When entering separately each marker in the model, results show no effect for any of those markers (Table 4).

### Table 4

<table>
<thead>
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<th>Odds Ratio</th>
<th>p value</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>TW4 BrdU</td>
<td>1.02</td>
<td>0.3</td>
<td>0.03</td>
</tr>
<tr>
<td>TW4 Ki67</td>
<td>0.93</td>
<td>0.1</td>
<td>0.08</td>
</tr>
<tr>
<td>TW4 CC3</td>
<td>1.05</td>
<td>0.5</td>
<td>0.01</td>
</tr>
<tr>
<td>TW4 CC3/BrdU</td>
<td>1.01</td>
<td>0.9</td>
<td>0.00</td>
</tr>
</tbody>
</table>

Treatment week (TW), bromodeoxyuridine (BrdU), caspase 3 (CC3). Analyses were controlled for age. Pseudo R² (Cox & Snell).

81% accurate in predicting later development of depression (Fig. 1c).
focused on two proliferating markers BrdU and Ki67, and on the apoptotic marker CC3. Results show no significant effect for any of those markers on later development of depression (Table 5).

### 3.3.2. Differentiation and apoptosis

As before, we then investigated whether changes between TW0 and TW4 also in the number of neurogenic and apoptotic markers were predictive of later depression.

As mentioned above, DCX+ cells overall tended to increase when cells were treated with TW4 serum compared with TW0 serum; however, patients who developed IFN-α–induced depression had a significantly lower increase in the number of DCX+ cells between TW0 and TW4 (see Supplementary Table 1).

When entering separately each marker in the model, results show that a lower increase in DCX+ cells between TW0 and TW4 is predictive of later development of depression. In particular, for a one-unit lower in the number of ΔTW4-TW0 DCX+ cells, there is a 14% increment in the odds of becoming depressed (odds ratio = 0.86, p = 0.006; Fig. 3a and b). In addition, a ROC curve was generated showing that the model with ΔTW4-TW0 DCX as variable is 89% accurate in predicting later development of depression (Fig. 3c).

### 3.4. Low neurogenesis at TW4 and a lower increase in neurogenesis between baseline and TW4 most accurately predict IFN-α–induced depression

Finally, having discovered that a high number of TW0 CC3+, TW0 CC3/BrdU+, and a low number of TW4 DCX+ and ΔTW4-TW0 DCX+ cells, were significantly predictive of IFN-α–induced depression, we subsequently investigated which model (containing those markers as variables) could be defined as the best one in terms of accuracy when predicting IFN-α–induced depression. Pairwise analysis comparing ROC curves previously generated from the apoptotic (TW0 CC3 and TW0 CC3/BrdU) and neurogenic (TW4 DCX and ΔTW4-TW0 DCX) markers do not show any significant difference in the accuracy of predicting IFN-α–induced depression (respectively, 77%, 81%, 88% and 89% accuracy; Fig. 4a and b), even if numerically the TW4 DCX and the ΔTW4-TW0 DCX have the highest predictive values.

### 4. Discussion

Using a model of inflammation-induced depression, in this study we provide the first evidence that blood factors, contained in the systemic milieu of patients receiving chronic treatment with IFN-α, can alter cell apoptosis and the neurogenic process in vitro, and potentially activate cellular processes that ultimately may contribute to the development of IFN-α–induced depression. In particular, we demonstrated that higher apoptosis upon treatment with baseline (TW0) serum, lower neurogenesis upon treatment with TW4 serum, and a lower increase in neurogenesis between treatments with TW0 and TW4 serum, are predictive of later development of depression – the last two models with,

### Table 5

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<th>Odds Ratio</th>
<th>p value</th>
<th>R²</th>
</tr>
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<tbody>
<tr>
<td>ΔTW4-TW0 BrdU</td>
<td>0.91</td>
<td>0.82</td>
<td>0.002</td>
</tr>
<tr>
<td>ΔTW4-TW0 Ki67</td>
<td>0.93</td>
<td>0.61</td>
<td>0.01</td>
</tr>
<tr>
<td>ΔTW4-TW0 CC3</td>
<td>0.95</td>
<td>0.36</td>
<td>0.03</td>
</tr>
<tr>
<td>ΔTW4-TW0 CC3/BrdU</td>
<td>0.7</td>
<td>0.055</td>
<td>0.13</td>
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</table>

Legend: treatment week (TW), bromodeoxyuridine (BrdU), caspase 3 (CC3). Analyses were controlled for age. Pseudo R² (Cox & Snell).
respectively, 88% and 89% accuracy.

Firstly, our findings indicate that baseline peripheral factors can be predictors of depression when measuring their impact on HPC apoptosis. Indeed, although none of the proliferating and neurogenic markers were affected upon treatment with baseline samples, a high number of cells positive for the apoptotic marker CC3 was predictive of later depression. However, using a larger cohort of IFN-α treated HCV patients (from which we have selected all serum samples used in the present study) we have found no differences at TW0 in the levels of key inflammatory cytokines, including IL-1β, IL-6, IL-17 and TNF-α, between those who later develop or do not develop depression (although, interestingly, we found differences in the prediction of fatigue) (Hepgul et al., 2016; Russell et al., 2019). This lack of difference in ‘usual suspect’ cytokines suggests the involvement of other factors contained in serum from depressed patients that are able to contribute to the increase in cell apoptosis detected in our in vitro study. Indeed, our transcriptome analysis conducted in the same larger cohort of IFN-α treated HCV patients revealed the presence of a distinct set of genes differentially expressed at TW0 between depressed and non-depressed patients (Hepgul et al., 2016). Among them, ubiquitin-fold modifier 1 (UFM1) and eukaryotic translation initiation factor 4B (EIF4B) were highly upregulated in patients who developed IFN-α-induced depression (Hepgul et al., 2016). UFM1 and EIF4B gene belong to respectively, the oxidative stress response signalling and the rapamycin signalling pathway, which are associated with regulation of apoptosis, and synaptic plasticity (Chen et al., 2012; Narasimhan et al., 2012), therefore potentially involved in the detrimental effect seen on cell death upon treatment with serum from depressed patients.

In this study we also demonstrate that lower levels of neurogenesis upon treatment with serum at TW4, and a lower increase in neurogenesis between TW0 and TW4, are predictive of later development of depression. Similarly to baseline, results from cytokines analysis conducted in samples from our larger cohort of IFN-α treated HCV patients, and collected at TW4, did not reveal any differences in the expression of the candidate cytokines previously mentioned, either as absolute values at TW4 or as increased between TW0 and TW4 when comparing depressed with non-depressed (Hepgul et al., 2016; Russell et al., 2019). However, data from our aforementioned transcriptome analysis show that, at TW4, patients who developed depression had a higher expression of a distinct set of genes, including cyclin dependent kinase 5

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<tr>
<td>ΔTW4–TW0 DCX</td>
<td>.86</td>
<td>0.006</td>
<td>.34</td>
</tr>
<tr>
<td>ΔTW4–TW0 MAP2</td>
<td>.92</td>
<td>0.32</td>
<td>.01</td>
</tr>
<tr>
<td>ΔTW4–TW0 CC3</td>
<td>.95</td>
<td>0.41</td>
<td>.02</td>
</tr>
<tr>
<td>ΔTW4–TW0 CC3/MAP2</td>
<td>1.04</td>
<td>0.71</td>
<td>.005</td>
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</table>

Legend: treatment week (TW), doublecortin (DCX), microtubule associated protein 2 (MAP2), caspase 3 (CC3). Analyses were controlled for age. Pseudo R2 (Cox & Snell).

Fig. 3. Changes in neurogenic and apoptotic markers between treatment week 0 (TW0) and TW4 as predictors of IFN-α–induced depression. Main predictive outcomes of differences in neurogenic and apoptotic markers between TW0 and TW4 on later IFN-α–induced depression (a). Representative images of cells treated with TW0 and TW4 serum from depressed and non-depressed patients. Neuroblasts were stained with doublecortin (DCX) (green labelling) (b). Receiver Operating Characteristic (ROC) curve of ΔTW4–TW0 DCX predictor of IFN-α–induced depression (c).
in vivo from the scenario of an adult mechanisms occurring in the hippocampal progenitor cells, may di stage during the treatment.

identify potential neuroprotective mechanisms activated only at a later required, especially upon a longer exposure to IFN-α– induced depression (b). Further characterisation of the cellular mechanisms underlying the effect of IFN-α on neurogenesis are required, especially upon a longer exposure to IFN-α, in order to identify potential neuroprotective mechanisms activated only at a later stage during the treatment.

Of course, this immortalized cell line we used in the study, while being of invaluable importance for our understanding of molecular mechanisms occurring in the hippocampal progenitor cells, may differ from the scenario of an adult in vivo environment and the adult neurogenic niche, in particular because of the absence of microglia cells, which are well-known regulators of inflammatory signaling pathways. However, our previous studies with this in vitro model have been successfully replicated in animal studies, including changes in neurogenesis by cortisol, IL-1 and antidepressants, and changes in stress- and antidepressants-regulated genes (Anacker et al., 2013a, 2013b, 2011; Borsini et al., 2017b; Horowitz et al., 2014; Zunszain et al., 2012). Therefore, we are confident that our results are relevant to the human brain.

The number of samples is also very small and collected primarily from male patients, of whom we have relatively few information with respect to their clinical profile. Therefore, the predictive models presented here should be replicated in a large cohort of patients. However, this was an extremely difficult group of patients to recruit and furthermore, the vast majority of patients recruited had to be excluded due to our strict criteria (see Methods). These strict criteria however allowed us to eliminate potential confounding factors, which otherwise might have affected the results. Finally, no control or placebo group was used for this study. However, the main aim in this case was not to test the effect of IFN-α on later depression, but instead to identify novel biological predictors for the development of depression. As such, we used serum samples of patients who did develop IFN-α-induced depression in comparison with serum samples of patients who did not develop IFN-α-induced depression.

5. Conclusions

In summary, our in vitro study demonstrates that hippocampal progenitor cells can be regulated by treatment with serum from IFN-α–treated HCV patients, and that subsequent alteration in neurogenesis and in cell survival is predictive of later development of depression. We believe that these findings may shed light on the pathogenesis of the depressive disorder, and further propose that the set-up employed in the present in vitro study, using a human HPCs experimental model together with a medium-throughput serum screening assay, offers an excellent opportunity for identification of novel therapeutic strategies for treatment of neuropsychiatric disorders.

Acknowledgements

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Declaration of Competing Interest

This work was supported by a Janssen Pharmaceutica PhD Studentship to Dr. Borsini, as part of a larger research grant on depression and inflammation by Johnson & Johnson to Drs. Pariante, Thuret, Mondelli and Zunszain. In addition, Professor Pariante and Dr. Mondelli have received research funding from the Medical Research Council (UK) and the Wellcome Trust for research on depression and inflammation as part of two large consortia that also include Johnson & Johnson, GSK and Lundbeck. All other authors declare no conflict of interest.
References


