Acetaminophen (APAP) is an effective antipyretic and one of the most commonly used analgesic drugs. Unlike antipyretic non-steroidal anti-inflammatory drugs, APAP elicits hypothermia in addition to its antipyretic effect. Here we have examined the mechanisms responsible for the hypothermic activity of APAP. Subcutaneous, but not intrathecal, administration of APAP elicited a dose dependent decrease in body temperature in wildtype mice. Hypothermia was abolished in mice pretreated with resiniferatoxin to destroy or defunctionalize peripheral TRPV1-expressing terminals, but resistant to inhibition of cyclo-oxygenases. The hypothermic activity was independent of TRPV1 since APAP evoked hypothermia was identical in wildtype and \( \text{Trpv1}^{-/-} \) mice, and not reduced by administration of a maximally effective dose of a TRPV1 antagonist. In contrast, a TRPA1 antagonist inhibited APAP induced hypothermia and APAP was without effect on body temperature in \( \text{Trpa1}^{-/-} \) mice. In a model of yeast induced pyrexia, administration of APAP evoked a marked hypothermia in wildtype and \( \text{Trpv1}^{-/-} \) mice, but only restored normal body temperature in \( \text{Trpa1}^{-/-} \) and \( \text{Trpa1}^{-/-} \text{Trpv1}^{-/-} \) mice. We conclude that TRPA1 mediates APAP evoked hypothermia.
neurons and TRPA1 are necessary for the hypothermic response to stimulation with APAP. Earlier significantly different between naïve wildtype and Trpa1−/− mice during RTX treatments in mice fitted with Bio-thermo iDENTICHIPS. The basal body temperature was not significantly altered by activation of TRPA1 channels expressed in the central terminals of sensory neurons. However, a TRPA1 antagonist would rely on some degree of tonic TRPA1 activity as is thought to be the case for TRPV1. Involvement of TRPA1.

Results

Involvement of TRPA1. We monitored the core body temperature in response to pharmacological treatments in mice fitted with Bio-thermo iDENTICHIPS. The basal body temperature was not significantly different between naïve wildtype and Trpa1−/− mice. Subcutaneous (s.c.) injections of APAP evoked a time- and dose-dependent hypothermia in C57BL/6J mice (n = 6–12). (B) APAP (300 mg/kg) reduced the body temperature in Trpa1+/+ mice but was without effect in Trpa1−/− mice (n = 6–11, see panel C for key to the symbols). (C) Intrathecal administration of APAP (100μg) did not reduce body temperature (n = 6). (D) Resiniferatoxin (RTX, 6.3 μg/kg, s.c.) induced a profound hypothermia in Trpa1+/+ and Trpa1−/− mice (n = 6). (E) APAP (300 mg/kg s.c.) evoked hypothermia in non-RTX treated Trpa1+/+ mice, but was without effect in Trpa1−/− and RTX treated Trpa1+/+ mice (n = 6). (F) Indomethacin (10 mg/kg, s.c.) did not alter body temperature and did not inhibit APAP (300 mg/kg) evoked hypothermia (n = 6). (G) Oral administration of the TRPA1 antagonist CHEM5861528 (40–300 mg/kg, 45 min before APAP) produced a dose-dependent inhibition of APAP evoked hypothermia (n = 6). (H) Intrapitoneal injections of the TRPA1 agonist cinnamaldehyde (CA, 125 mg/kg) induced a marked hypothermia in Trpa1+/+ mice, which was significantly reduced in Trpa1−/− mice (n = 6). Data are mean ± sem; p < 0.05, **p < 0.01, ***p < 0.001 compared to vehicle and †p < 0.05, ††p < 0.01, †††p < 0.001 compared to Trpa1+/+.

antagonist was reported to have no hyperthermic or hypothermic effect,[13] but a direct effect of an antagonist would rely on some degree of tonic TRPA1 activity as is thought to be the case for TRPV1.[12]

In the current study we used genetically modified mice and pharmacological tools to determine the contribution of peripheral sensory neurons to the APAP evoked hypothermia. Our data demonstrate that APAP evoked hypothermia in mice is dependent on the presence of functional TRPA1 channels. The hypothermic effect of APAP is lost in Trpa1−/− mice and inhibited by administration of a TRPA1 agonist.
investigations proposed that APAP acts by inhibiting either COX-1 or COX-2. However, pretreatment with the nonselective COX inhibitor indomethacin (10 mg/kg, s.c.) did not alter the body temperature in mice and also failed to influence the hypothermia evoked by APAP, demonstrating that APAP acts independently of COX-1 and COX-2.

Since our earlier investigations identified activation of TRPA1 by NAPQI formed from APAP as an analgesic mechanism of action, we examined whether the APAP induced hypothermia could be prevented by treatment with a pharmacological inhibitor of TRPA1. Treatment with the selective TRPA1 inhibitor CHEM5861528 (40–300 mg/kg, p.o.) had no acute effect on body temperature in wild-type mice (vehicle 37.8 ± 0.1 °C, n = 18; CHEM5861528 40 mg/kg 37.8 ± 0.1 °C, n = 6; 100 mg/kg 38.0 ± 0.1 °C, n = 6; 300 mg/kg 37.7 ± 0.2 °C, n = 6; p > 0.05) but pre-treatment with CHEM5861528 produced a dose-dependent inhibition of the APAP induced hypothermia. At the highest dose tested the effects of APAP were essentially abolished.

APAP is metabolized to NAPQI and benzoquinone which are both TRPA1 agonists. We therefore examined if other TRPA1 agonists evoke hypothermia. Cinnamaldehyde (125 mg/kg) was administered intraperitoneally to Trpa1−/− and wild-type mice. A large reduction in body temperature was noted in wild-type mice, and this was greatly reduced, although not abolished, in the Trpa1−/− mice (Fig. 1H).

Involvement of TRPV1. Since TRPV1 inhibition evokes hyperthermia and TRPV1 activation elicits hypothermia, we re-evaluated the effects of APAP in wild-type mice treated with the TRPV1 antagonist BCTP, and in Trpv1−/− mice. The basal body temperature was not different in Trpv1+/+ and Trpv1−/− mice (Trpv1+/+ 37.8 ± 0.1 °C; Trpv1−/− 37.8 ± 0.1 °C, p > 0.2, n = 11 groups of 6 for each genotype). BCTP (30 mg/kg p.o.) evoked a small degree of hyperthermia in Trpv1+/+ mice but did not inhibit the hypothermia evoked by subsequent administration of APAP (Fig. 2A). In fact the reduction in body temperature was larger in mice treated with the TRPV1 antagonist, since APAP reduced the body temperature to the same level in mice treated with BCTP or vehicle. Similarly APAP exerted a robust hypothermic effect in Trpv1−/− mice that was at least as large as that observed in wild-type littermates (Fig. 2B). These observations confirm the results of Ayoub et al., which demonstrated that TRPV1 is not involved in APAP evoked hypothermia.

Effects on yeast-induced fever. We next investigated the importance of TRPA1 or TRPV1 for the antipyretic activity of APAP. Pyrexia was induced by subcutaneous administration of yeast (2 g/kg in saline). Such a treatment resulted in an increase in body temperature of approximately 1 °C in wild-type mice and in mice lacking functional TRPV1 or TRPA1 (Fig. 3). This differs from the findings of Iida, et al., who found a reduced hyperthermia in Trpv1−/− mice after intraperitoneal injection of lipopolysaccharide. Subsequent administration of APAP reduced body temperature and produced a robust hypothermia in both wildtype and Trpv1−/− mice (Fig. 3A). In contrast, while the strong hypothermic effect of APAP was absent in Trpa1−/− mice, the anti-pyretic effect of APAP was still evident, demonstrating
that APAP produces antipyretic and hypothermic effects through distinct and independent mechanisms (Fig. 3B).

As it is possible that the anti-pyretic effect of APAP observed in \( \text{Trpa1}^{-/-} \) mice was due to an action on TRPV1, we also examined the effects of APAP in yeast treated mice lacking both TRPV1 and TRPA1 (Fig. 4A). The basal body temperature of \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \) mice was slightly, but not significantly, higher than in matched wildtype control groups (\( \text{Trpa1}^{+/+}/\text{Trpv1}^{+/+} \): 37.5 ± 0.2, range 36.9–37.9; \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \): 37.9 ± 0.3, range 37.1–38.6, \( p = 0.19, n = 7 \) groups of 6 mice of each genotype). Yeast administration evoked a hyperthermic response in double knockout (\( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \)) mice that was similar in magnitude to that seen in wild-type mice. Subsequent administration of APAP reduced body temperature to normal levels indicating that the anti-pyretic effects of APAP were still operational in these double knockout mice.

APAP has been proposed to exert some of its actions through inhibition of cyclo-oxygenase enzymes, despite displaying a relatively modest affinity as a COX inhibitor compared to those of NSAIDs. To determine whether the antipyrexia, unlike the hypothermia, can be explained by APAP inhibition of COX, we examined the effect of APAP and indomethacin in \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \) mice made hyperthermic by yeast administration. Wildtype and \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \) mice were injected with indomethacin (10 mg/kg, s.c.) 18 h after administration of yeast and 1 hour before administration of vehicle or APAP (Fig. 4B,C). Indomethacin reduced body temperature by approximately 1 °C in both wild-type and \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \) mice thereby reversing the pyrexia. Subsequent administration of APAP evoked the typical hypothermia in wild-type mice but had no significant effect on body temperature in \( \text{Trpa1}^{-/-}/\text{Trpv1}^{-/-} \) mice. In the double knockout mice a small further reduction in temperature was noted after APAP treatment (Fig. 4C), but a similar reduction was noted in the vehicle treated group (Fig. 4B), indicating that this small hypothermic effect is independent of APAP but may instead be caused by the continued action of indomethacin.

Is TRPA1 responsible for the hypothermic effects of other agents? A number of other hypothermic agents have been reported to act as TRPA1 agonists. These include some cannabinoid receptor ligands, morphine and isoflurane.

A reduction in core body temperature is a well known response to cannabinoid receptor 1 (CB1) agonists and several naturally occurring and synthetic CB1 agonists have been shown to be TRPA1 agonists. We therefore compared the effects of WIN55,212-2 on body temperature in \( \text{Trpa1}^{-/-} \) mice and wild-type littermates. WIN55,212-2 (3 mg/kg s.c.) evoked similar reductions (6–7°C) in body temperature in both genotypes that lasted for at least 3 hours (Fig. 5A).

Morphine can lower body temperature in a number of species, notably at higher doses and at high concentrations acts as a TRPA1 agonist. Sub-cutaneous administration of 10 mg/kg morphine lowered
the body temperature of wild-type mice by 5 °C after 45–60 minutes and the amplitude and duration of the hypothermic response was not significantly different in Trpa1−/− mice (Fig. 5B).

Isoflurane and other irritant general anaesthetics act as TRPA1 agonists30,31 and produce hypothermia in humans and rodents 32–35. We therefore examined body temperature during isoflurane anaesthesia in wild-type and Trpa1−/− mice. Isoflurane elicited a similar hypothermia of short duration in both genotypes (Fig. 5C), consistent with a TRPA1 independent mechanism.

Discussion

The mechanisms responsible for the analgesic and antipyretic actions of APAP have been the subject of investigation for many decades. Recent studies have concluded that APAP metabolites acting on TRP channels are important for the analgesic actions of the drug. Accordingly, APAP analgesia has been ascribed to AM404 activation of TRPV1 in the central nervous system4,5,20 and activation of TRPA1 in the spinal cord by local formation of the electrophilic NAPQI3. Other studies have suggested that the
cannabinoid receptor system contributes to APAP analgesia\(^6,7\). The antipyretic and hypothermic actions of APAP are not well understood. However, the findings that the hypothermic effects of APAP were retained in \(\text{Trpv1}^{-/-}\) mice as well as in mice lacking cannabinoid receptor 1 rule out a role for these pathways in APAP-evoked hypothermia\(^7\).

Our studies demonstrated that APAP produced a profound hypothermia in wild-type mice that was dependent on the presence of functional TRPA1 channels. APAP-evoked hypothermia was absent in \(\text{Trpa1}^{-/-}\) mice and inhibited by prior systemic treatment with a TRPA1 antagonist. The locus of hypothermic action is likely to be TRPA1 expressed in peripheral sensory neurons, since the response could be abolished by RTX mediated chemical ablation of TRPV1 containing sensory neurons. Body temperature is controlled by peripheral sensory input to the CNS as well as by central thermoregulatory mechanisms\(^3,38,39\). Recent studies on TRPV1 antagonists have highlighted the importance of TRPV1 expressing peripheral sensory nerves for temperature regulation. Stimulation of TRPV1 with systemic capsaicin treatment leads to hypothermia\(^3,38\), while administration of many (but not all) TRPV1 antagonists causes hyperthermia\(^12,40-42\). Functional ablation of these peripheral sensory neurons by prior administration of the ultrapotent TRPV1 agonist RTX results in a loss of TRPV1 mediated temperature changes\(^39\).

In our studies the hypothermic effects of APAP were abolished by a similar systemic treatment with RTX. However, the effect of APAP could not be ascribed to TRPV1 agonism as hypothermia was retained in \(\text{Trpv1}^{-/-}\) mice and unaffected by systemic administration of a dose of a TRPV1 antagonist that exerts a maximal analgesic effect\(^4\), in agreement with previous studies\(^3\). TRPA1 is predominantly expressed in TRPV1 containing sensory neurons\(^4\). The functional ablation of the TRPV1-expressing neurons by RTX will therefore have severely compromised or abolished TRPA1 responses, explaining the very marked effect of RTX treatment on APAP evoked temperature changes.

Intraperitoneal administration of the TRPA1 agonist cinnamaldehyde produced a significant hypothermia in wild-type mice that was greatly reduced, although not completely abolished, in \(\text{Trpa1}^{-/-}\) mice. This finding demonstrates that TRPA1 agonism can influence body temperature, almost certainly because TRPA1 is co-localized in TRPV1 containing thermoregulatory sensory nerves. In addition, it is consistent with the hypothesis that activation of TRPA1 by electrophilic APAP metabolites is responsible for the observed hypothermia. Electrophilic APAP metabolites are formed in the liver and liver afferent neurons are therefore prime candidates for the site of action. Importantly, we found that genetic or pharmacological blockade of TRPA1 does not produce hyperthermia in rodents, in agreement with an earlier report\(^4\). This observation indicates that TRPA1, unlike TRPV1, does not have a significant tonic activity in thermosensitive afferent fibres under normal conditions\(^1\).

Although TRPA1 is important for the hypothermic effects of APAP, our studies suggest that other mechanisms contribute to its antipyretic effects. Yeast induced hyperthermia was of a similar magnitude in wild-type, \(\text{Trpa1}^{-/-}\) and \(\text{Trpa1}^{-/-}/\text{Trpv1}^{-/-}\) (double knockout) mice. The absence of APAP evoked hypothermia in both \(\text{Trpa1}^{-/-}\) and \(\text{Trpa1}^{-/-}/\text{Trpv1}^{-/-}\) mice allowed the antipyretic effects to be studied in isolation. APAP reduced the elevated body temperature to normal levels in \(\text{Trpa1}^{-/-}\) and \(\text{Trpa1}^{-/-}/\text{Trpv1}^{-/-}\) mice, indicating that neither TRP channel plays a role in the antipyretic actions of APAP.

We explored the possibility that the antipyretic effect of APAP was mediated by cyclo-oxygenase inhibition by examining the effect of APAP in mice pre-treated with indomethacin. In wild-type and \(\text{Trpa1}^{-/-}/\text{Trpv1}^{-/-}\) mice, indomethacin reversed the pyrexia and normalized body temperature to the pre-yeast levels, indicating that these TRP channels are not involved in the antipyretic effects of cyclo-oxygenase inhibitors. Subsequent APAP administration evoked hypothermia in wild-type mice, which demonstrated that cyclo-oxygenase activity is not important for this effect of APAP. In indomethacin pre-treated \(\text{Trpa1}^{-/-}/\text{Trpv1}^{-/-}\) mice, a continued small slow reduction in body temperature was noted after administration of either APAP or vehicle. The lack of any obvious APAP induced temperature change when the cyclo-oxygenase enzymes are already inhibited could be interpreted as evidence that cyclo-oxygenase inhibition underlies the antipyretic effects of APAP. However, it is possible that APAP had no effect because the pyrexia had been reversed and body temperature normalized by the indomethacin treatment.

Although the TRPA1 agonist cinnamaldehyde evoked TRPA1-dependent hypothermia, a range of other hypothermic agents that have been reported to act as TRPA1 agonists retained the ability to reduce body temperature in \(\text{Trpa1}^{-/-}\) mice. These included the general anaesthetic isoflurane, the cannabinoid receptor 1 agonist WIN55,212-2 and the opioid receptor agonist morphine. Despite the common property of TRPA1 agonism, these diverse agents must reduce body temperature primarily by other mechanisms at the doses used in our experiments.

Conclusions

Our results demonstrate that activation of TRPA1 in sensory neurons mediates the hypothermic effect of APAP in mice. These findings support our previous identification of TRPA1 as a target for the analgesic activity of APAP\(^5\). Importantly, APAP does not reduce body temperature after intrathecal injections, demonstrating that hypothermia is not required for its spinal antinociceptive effect. The hypothermic effect of APAP is completely lost in \(\text{Trpa1}^{-/-}\) mice and resistant to inhibition of cyclo-oxygenase. However, it is possible that APAP exerts its antipyretic activity by cyclo-oxygenase inhibition, since APAP reversed yeast induced hyperthermia in mice lacking both TRPA1 and TRPV1. The TRPA1 mediated
hypothermia may explain the improved antipyretic activity seen with combinations of APAP and the NSAID ibuprofen.

**Methods**

**Animals.** All animal procedures were carried out in accordance with the UK Animals (Scientific Procedures) Act 1986. All experimental procedures were approved by the King's College London Animal Care and Ethics Committee. Data shown are from female C57BL/6J mice or from male and female homozygote Trpa1−/− and Trpv1−/− littermates. C57BL/6J mice were obtained from Charles River UK. TRPA1-null mice and wild-type littermates were bred from heterozygous mice provided by Drs Kelvin Kwan and David Corey (Harvard Medical School, Boston, MA). TRPV1–null mice were bred from homozygous mice obtained from the Jackson Laboratory (Bar Harbor, ME). Trpa1−/−/Trpv1−/− mice were generated in house by crossing the Trpa1−/− and Trpv1−/− mice.

**Temperature measurements.** Under general anaesthesia, the lower back of the mice was shaved. The mice were then implanted subcutaneously in the shoulder region with a small identification chip incorporating a quick reading thermometer accurate to 0.1 degree °C (iDENTICHIP® with Bio-Thermo™, AnimalCare Ltd. UK). The small skin incision was closed using tissue adhesive (Vetbond, 3M).

Bio-Thermo chips have many advantages over other commonly used methods for recording body temperature. Body temperatures can be measured repeatedly without disturbing the animals using a hand-held scanner. As each chip has a unique identity, they enable animals to be group housed in their home cages. It is less stressful than rectal temperature recording and is cheaper than radiotelemetry methods.

During each experiment, body temperature was recorded every 15 min for up to 45 min prior to drug administration to establish baseline temperatures and then post dose for up to 24h.

**Drugs.** Acetaminophen (APAP; 4-Acetamidophenol; Sigma-Aldrich, UK) and morphine (Sigma-Aldrich, UK) were prepared in 0.9% saline (with warming for APAP). WIN55,212-2 (Tocris, UK) was made up in 10% DMSO/saline. Mice were exposed to isoflurane (Abbott Industries Ltd. UK) at 2.5% during each experiment, body temperature was recorded every 15 min for up to 45 min prior to drug administration to establish baseline temperatures and then post dose for up to 24h.

During each experiment, body temperature was recorded every 15 min for up to 45 min prior to drug administration to establish baseline temperatures and then post dose for up to 24h.

**Data analysis.** All data are presented as the mean ± sem. Plots show the change in body temperature from baseline readings or the raw recorded temperature. Statistical analysis was carried out on raw data using repeated measures ANOVA plus post-hoc analysis using Tukey’s HSD test (p < 0.05 was set as the level of statistical significance).

**References**


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Competing financial interests: Additional Information

C.G. performed all experiments. C.G., D.A.A. and S.B. designed and analysed experiments and wrote the paper.

Author Contributions

C.G. performed all experiments. C.G., D.A.A. and S.B. designed and analysed experiments and wrote the paper.

Additional Information

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