Biomarkers of Tolerance in Kidney Transplantation: Are We Predicting Tolerance or Response to Immunosuppressive Treatment?

I. Rebollo-Mesa1,2,3, E. Nova-Lamperti1, P. Mobillo1, M. Runglall4, S. Christakoudi1,2, S. Norris1,5, N. Smallcombe1, Y. Kamra1,6, R. Hilton7, Indices of Tolerance EU Consortium8-14, S. Bhandari15, R. Baker16, D. Berglund17, S. Carr18, D. Game7, S. Griffin19, P. A. Kalra20, R. Lewis21, P. B. Mark22, S. Marks23, I. Macphee24, W. McKane25, M. G. Mohaupt26, R. Pararajasingam27, S. P. Kon28, D. Seron29, M. D. Sinha30, B. Tucker28, O. Viklicky31, R. I. Lechner1,32, G. M. Lord1,4,7 and M. P. Hernandez-Fuentes1,4,*

1Medical Research Council Centre for Transplantation, King’s College London, London, United Kingdom
2Biostatistics, Institute of Psychiatry, Psychology and Neuroscience, King’s College London, London, United Kingdom
3UCB Celltech, UCB Pharma S.A., Slough, United Kingdom
4National Institute for Health Research Biomedical Research Centre, Guy’s and St. Thomas’ National Health Service Foundation Trust, King’s College London, London, United Kingdom
5University College London, London, United Kingdom
6Guy’s and St. Thomas’ NHS Foundation Trust, London, United Kingdom
7Imperial College London, London, United Kingdom
8King’s College Hospital NHS Foundation Trust, London, United Kingdom
9Great Ormond Street Hospital for Children NHS Foundation Trust, London, United Kingdom
10University College Hospital, London, United Kingdom
11Imperial College London, London, United Kingdom
12Institute for Medical Immunology, Université Libre de Bruxelles, Bruxelles, Belgium
13Milenyi Biotec, Bergisch Gladbach, Germany
14University of Nantes, Nantes, France
15Charité, Universitätsmedizin Berlin, Berlin, Germany
16Hull and East Yorkshire Hospitals NHS Trust, Hull, United Kingdom
17St. James’s University Hospital, Leeds, United Kingdom
18Uppsala University Hospital, Uppsala, Sweden
19Leicester General Hospital, Leicester, United Kingdom
20Cardiff and Vale University Health Board, Cardiff, United Kingdom
21Queen Alexandra Hospital, Portsmouth, United Kingdom
22University of Glasgow, Glasgow, United Kingdom
23Great Ormond Street Hospital for Children NHS Foundation Trust, London, United Kingdom
24St. George’s Hospital, London, United Kingdom
25Northern General Hospital, Sheffield, United Kingdom
26INSELSPITAL, Universitätsspital Bern, Klinik für Nephrologie/Hypertonie Abteilung für Hypertonie, Bern, Switzerland
27Manchester Royal Infirmary, Manchester, United Kingdom
28King’s College Hospital NHS Foundation Trust, London, United Kingdom
29Hospital Universitari Vall d’Hebron, Barcelona, Spain
30Evelina London Children’s Hospital, London, United Kingdom
31Transplantaci laborato íIKEM, Prague, Czech Republic
32King’s Health Partners, London, United Kingdom
*Corresponding author: Maria P. Hernandez-Fuentes, maria.hernandez@kcl.ac.uk

We and others have previously described signatures of tolerance in kidney transplantation showing the differential expression of B cell–related genes and the relative expansions of B cell subsets. However, in all of these studies, the index group—namely, the tolerant recipients—were not receiving immunosuppression (IS) treatment, unlike the rest of the comparator groups. We aimed to assess the confounding effect of these regimens and develop a novel IS-independent signature of tolerance. Analyzing gene expression in three independent kidney transplant patient cohorts (232 recipients and 14 tolerant patients), we have established that the expression of the previously reported signature was biased by IS regimens, which also influenced transitional B cells. We have defined and validated a new gene expression signature that is independent of drug effects and also differentiates tolerant patients from healthy controls (cross-validated area under the receiver operating characteristic curve [AUC] = 0.81). In a prospective cohort, we have demonstrated that the new signature remained stable before and after steroid withdrawal. In addition, we report on a validated and highly accurate gene expression signature that can be reliably used to identify patients suitable for IS reduction (approximately 12% of stable patients), irrespective of the IS drugs they are receiving. Only a similar
approach will make the conduct of pilot clinical trials for IS minimization safe and hence allow critical improvements in kidney posttransplant management.

Abbreviations: AUC, area under the receiver operating characteristic curve; Aza, azathioprine; CNI, calcineurin inhibitor; Cyc, ciclosporin A; DMSO, dimethyl sulfoxide; GMCSF, granulocyte macrophage colony-stimulating factor; IS-IE, IS-independent expression, the new signature of tolerance; IS, immunosuppression; KTR, kidney transplant recipient; MMF, mycophenolate mofetil; NFκB, nuclear factor kappa B; PBMC, peripheral blood mononuclear cell; Pred, prednisone; REGGR, glucocorticoid receptor regulatory network; Tac, tacrolimus; TNF, tumor necrosis factor

Received 24 January 2016, revised 12 May 2016 and accepted for publication 08 June 2016

Introduction

Despite improvements in the stratification of therapy in kidney transplant recipients (KTRs), grafts do not display the desired longevity. The immunosuppression (IS) needs of individual patients are poorly defined, and the current approaches are far from an ideal personalized management program. Standard KTRs are maintained with calcineurin inhibitors (CNIs) or sirolimus on doses defined by blood levels and azathioprine (Aza) is adjusted on weight, whereas mycophenolate mofetil (MMF) or prednisone doses are given as population-based results dictate (1,2). As a result, overimmunosuppression is responsible for a high number of patients with a functioning graft dying or suffering from cancer, infections or cardiovascular events (3). For example, prolonged intake of azathioprine has long been associated with increased incidence of tumors, particularly skin cancers in KTRs (4). At the same time, underimmunosuppression remains a clinical problem in that acute rejection is still frequent (5), albeit decreasing in magnitude, and chronic rejection is definitively significant and has a poor outcome (6). This means that there is an important clinical need to characterize biomarker signatures that could reliably identify patients who have developed a tolerant response to their graft. We and others have been trying to address this need (7–11).

The index patient group that is central to identifying tolerance is formed by those kidney transplant recipients (KTRs) who have challenged conventional clinical practice and have discontinued immunosuppressive medication while maintaining good graft function for years (12); these recipients are thus labeled as “tolerant.” In the process of biomarker discovery, these tolerant KTRs—free from IS for years—have always been compared with KTRs receiving various IS regimens, who are representatives of nontolerant patients. These differences in therapy between the groups imply that any previous signatures suffer from a systematic analysis flaw if the effects of the drugs have not been accounted for.

The final proof that any biomarker signature is indeed a signature of tolerance would only arise through a clinical trial whereby patients displaying the chosen signature would be weaned off IS and their graft would maintain good function. This definitive evidence for any signature of tolerance is still missing in the literature. Consequently, the selection of possible signatures of tolerance need to abide by the most stringent quality requirements because testing them in prospective trials would put some patients at risk of late rejection or even at risk of irreversible graft damage if a misdiagnosis is made. To our knowledge, none of the referenced studies have yet attempted to directly address the fact that IS drugs affect gene expression despite available evidence (13,14). Notably, a noninvasive signature of tolerance could also be used to evaluate the effectiveness of tolerance induction therapies that are currently under investigation in clinical trials (15–17).

Gene expression patterns found in the peripheral blood of KTRs can reflect at least two mechanisms: the response of the recipient’s immune system to the presence of a highly immunogenic tissue (the transplanted graft) and the effect of IS treatment used to counteract the rejection process. Edemir et al (18) used a rodent model to describe gene expression patterns that support the spontaneous simultaneous activation of immune effector–related pathways and protective and immune counter-regulatory mechanisms as a response to the allogeneic transplant. In humans, we and others have previously described a dysregulation of B cell–related genes in tolerant recipients—associated with the maintenance or expansion of transitional B cells in peripheral blood (8,9)—that elicited new avenues toward understanding the role of transitional B cells in tolerance (19–21). Differential expression profiles associated with IS treatment have been demonstrated by Erickson et al (22) in the context of transplantation in rats. Thus, when investigating gene expression markers of operational tolerance in humans, we need to ensure that we are isolating the natural counter-regulatory immune mechanisms from those that reflect the IS drug intake, which could disappear after discontinuation of the drug.

We therefore undertook the current study to explore the effects of IS regimens on gene expression in peripheral blood and on our previously described signature of tolerance. Sagoo et al (8) and hypothesized that adjusting for the confounding effects of IS drugs would provide more reliable biomarkers. We also hypothesized that by targeting immune responses via different mechanisms, IS drugs would have a differential effect on lymphocyte populations, particularly that of transitional B cells, which could bias the probability of tolerance estimates if differences in IS drug regimens are not accounted for. As a proof of concept of the effect of IS on lymphocyte subsets and gene expression, we have prospectively collected samples from
patients who underwent steroid withdrawal owing to clinical reasons.

Concise Materials and Methods

Patients and samples

To collect the necessary evidence described in this article, we used samples from three patient cohorts:

- Cohort 1: We performed a reanalysis of the data from 71 European KTRs from the previously published Indices of Tolerance (IoT) study. This patient cohort had been used to discover the original biomarker signature and comprised 11 tolerant recipients, 51 stable patients (30 on standard triple therapy, 10 who had never received a CNI and 11 on low doses of prednisone), 9 patients with biopsy-proven chronic rejection and 19 healthy controls; these patients are all thoroughly described in reference (8).

- Cohort 2: Cohort 2 is a novel observational case-control cohort from the Genetic Analysis of Molecular Biomarkers of Immunological Tolerance (GAMBIT) study (Research Ethics Reference: 09/H0713/12). The cohort comprised tolerant (n = 14; detailed description in Table S3), stable (n = 190) and chronic rejection (n = 36) patients and healthy controls (n = 12). At least two blood samples—6 months apart on average—were obtained from each individual; these are identified in the text as time point 1 and time point 2.

- Cohort 3: Cohort 3 is a prospective cohort from the same GAMBIT study (Research Ethics Reference: 09/H0713/12) that included stable patients who, early posttransplant, were undergoing steroid withdrawal owing to clinical reasons. These patients were selected exclusively based on clinical criteria and were recruited from the London and Portsmouth hospitals. Patient selection and steroid withdrawal were conducted according to local clinical practice. Samples were collected before and 2 to 6 months after complete steroid withdrawal.

Patient characteristics from cohorts 2 and 3 are described in Table 1, and further clinical details are given in Data S1.

The characteristics of the patients in the prospective steroid withdrawal cohort 3 were largely comparable to the observational cohort 2 (Table 1), except that their time posttransplantation was significantly shorter (p < 0.001), none had Donor Specific Antibodies (DSA) and none were receiving azathioprine, whereas all but one were treated with mycophenolate mofetil (Table 1).

RNA isolation, complementary DNA (cDNA) synthesis and reverse transcription quantitative real-time polymerase chain reaction (RT-qPCR)

Peripheral vein blood was drawn directly into Tempus Blood RNA Tubes (Life Technologies, Paisley, UK) and stored at −20°C. RNA isolation, cDNA synthesis, RT-qPCR conditions and primers are described in Data S1.

Fluidigm platform

The expression levels of a set of target genes and three endogenous reference genes were measured in 470 RNA samples on the Fluidigm Bio-Mark quantitative real-time PCR (qPCR) platform (South San Francisco, CA) with a preamplification step to validate the signature rederived from the IoT cohort. Further details are given in Data S1.

Flow cytometry

Peripheral blood mononuclear cells (PBMCs) were isolated and frozen immediately at −80°C. After 24 h, cells were transferred into liquid nitrogen −170°C and kept until use. Antibody panels and acquisition details are described in Data S1.

Statistical analyses

The analysis strategy is depicted in Figure 1. All statistical analyses (preprocessing of RT-qPCR, Fluidigm and array data) were carried out in R software (http://www. R-project.org) (23–25). For individual gene expression, Ct values were normalized using DCT with respect to HPRT (hypoxanthine phosphoribosyltransferase) and were used as log2(2−ΔCt) values.

Associations of gene expression or of predicted probability of tolerance with IS drug intake were examined in linear regression models adjusting simultaneously for confounding by IS drugs other than the drug of interest. Bonferroni correction was applied for multiple comparisons between patients on and off different drugs.

Elucidating the confounding effect of IS on gene expression

Details of the RISET 2.0 array and the preprocessing method have been published elsewhere (8). The array-wide analysis of drug effects was carried out using empirical Bayes moderated linear models implemented in the Limma package (26). The Benjamini–Hochberg (BH) method was used for multiple testing correction. Genes for which expression was not significantly above background were filtered out prior to analysis to increase statistical power (27).

Defining IS-independent expression

The residuals of a linear regression model relating gene expression data from stable and chronic rejector patients to drug intake (binary yes/no) represent the IS-independent expression (IS–IE). The gene expression of tolerant patients and healthy controls was rescaled to IS–IE by subtracting the intercept of that model from the raw expression.

Defining the novel signature of tolerance

We used the regularized multivariate logistic regression method ElasticNet (28,29) to select an optimal set of genes predictive of tolerance (as the number of genes was high relative to the number of patients), and considering many genes were correlated, thus invalidating the application of classical regression models. We compared the estimated IS–IE in tolerant patients versus patients on IS (stable and with chronic rejection). In order to test the stability of the signatures, IS–IE from the time point 2 samples was estimated based on the model trained on time point 1 samples, and classification accuracy was evaluated with the same cutoff.

For the identification of differentially expressed biological pathways, we carried out gene set analysis (30) using the curated list of gene sets from the Broad Institute (www. broadinstitute.org/gsea).

For the evaluation of predictive accuracy, an adjusted area under the receiver operating characteristic curve (AUC) was derived after fitting a linear regression model of the estimated probability of tolerance on drug regimen in IS patients and calculating the AUC for the residuals of this model (31).

Further details regarding the laboratory methods and statistical analysis are provided in Data S1.

Results

Immunosuppressants bias the expression of genes in peripheral blood

To study the effects of commonly used immunosuppressants on the expression of genes measured in peripheral blood, we first carried out a previously unattempted
<table>
<thead>
<tr>
<th>Clinical parameters</th>
<th>Retrospective cohort</th>
<th>Prospective cohort</th>
<th>Steroid withdrawal</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Tolerant(^1)</td>
<td>Stable(^2)</td>
<td>Chronic(^2) rejector</td>
</tr>
<tr>
<td>N</td>
<td>14</td>
<td>190</td>
<td>36</td>
</tr>
<tr>
<td>% female</td>
<td>21.4%(^nn)</td>
<td>32.6%</td>
<td>36.1%</td>
</tr>
<tr>
<td>% deceased</td>
<td>50.0%(^nn)</td>
<td>66.5%</td>
<td>65.7%</td>
</tr>
<tr>
<td>Years posttransplant(^4)</td>
<td>17.5(^*) (2.2, 30.8)</td>
<td>12.5 (4.2, 36.7)</td>
<td>6.9 (1.3, 27.8)</td>
</tr>
<tr>
<td>Age(^5)</td>
<td>48.8(^mm) (15.2)</td>
<td>50.3 (13.4)</td>
<td>43.9 (13.6)</td>
</tr>
<tr>
<td>eGFR(^5)</td>
<td>57.9(^mm) (14.4)</td>
<td>64.1 (23.0)</td>
<td>32.4 (13.0)</td>
</tr>
<tr>
<td>Nr HLA-MM(^6)</td>
<td>3 (0-5)(^mm)</td>
<td>2 (0-6)</td>
<td>3 (0-6)</td>
</tr>
<tr>
<td>% DSA</td>
<td>21.4%(^nn)</td>
<td>9.5%(^nn)</td>
<td>44.4%(^mm)</td>
</tr>
<tr>
<td>IS(^7,8)</td>
<td>–</td>
<td>26.8%</td>
<td>80.6%</td>
</tr>
<tr>
<td>% on Tac</td>
<td>–</td>
<td>46.3%</td>
<td>8.3%</td>
</tr>
<tr>
<td>% on Cyc</td>
<td>–</td>
<td>32.6%</td>
<td>13.9%</td>
</tr>
<tr>
<td>% on Aza</td>
<td>–</td>
<td>44.7%</td>
<td>63.9%</td>
</tr>
<tr>
<td>% on MMF</td>
<td>–</td>
<td>41.6%</td>
<td>69.4%</td>
</tr>
<tr>
<td>% on Pred</td>
<td>–</td>
<td>26.8%</td>
<td>80.6%</td>
</tr>
<tr>
<td>IS dose(^8)</td>
<td>Tac, ug/L</td>
<td>3.8 (2.1)</td>
<td>5.7 (4.1)</td>
</tr>
<tr>
<td></td>
<td>Cyc, ug/L</td>
<td>157 (713)</td>
<td>158 (38)</td>
</tr>
<tr>
<td></td>
<td>Aza, mg/day</td>
<td>87 (36)</td>
<td>105 (45)</td>
</tr>
<tr>
<td></td>
<td>MMF, mg/day</td>
<td>1200 (450)</td>
<td>1112 (482)</td>
</tr>
<tr>
<td></td>
<td>Pred, mg/day</td>
<td>4.8 (1.8)</td>
<td>6.7 (3.1)</td>
</tr>
<tr>
<td>WBC × 10(^{-9})(^5)</td>
<td>7.3(^mm) (1.8)</td>
<td>7.2 (2.2)</td>
<td>7.12 (2.5)</td>
</tr>
<tr>
<td>Lymph × 10(^{-9})(^5)</td>
<td>1.8(^&quot;) (0.6)</td>
<td>1.5 (0.6)</td>
<td>1.4 (0.9)</td>
</tr>
<tr>
<td>n with two samples</td>
<td>11</td>
<td>119</td>
<td>29</td>
</tr>
</tbody>
</table>

Healthy controls were age and gender matched to tolerants.

Aza, azathioprine oral dose; Cyc, cyclosporine A trough levels; DSA, Donor Specific Antibodies; eGFR, estimated Glomerular Filtration Rate; GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance; IS, immunosuppression; Lymph, Lymphocytes; MM, mycophenolate mofetil; MMF, mycophenolate mofetil oral dose; Nr HLA-MM, Number of Human Leucocyte Antigen mismatches (out of 6); Pred, prednisone oral dose; Tac, tacrolimus trough levels; WBC, White blood Cells.

\(^1\)Statistical significance of the differences between tolerant patients and the combined group of stable and chronic rejectors.

\(^2\)Statistical significance of the difference compared to tolerant patients.

\(^3\)Statistical significance of the difference compared to the total retrospective cohort.

\(^4\)Median (minimum, maximum).

\(^5\)Mean [standard deviation (SD)].

\(^6\)Median sum of HLA-A, HLA-B, and HLA-DR mismatches (minimum and maximum).

\(^7\)Percentage (%) from the total number of patients in the group (information on IS drugs was absent for eight stable patients [4.2%] and two chronic rejector patients [5.6%]).

\(^8\)Mean (SD) doses at recruitment in mg are displayed.

The statistical significance of the difference between stable and chronic rejectors and the prospective steroid withdrawal cohort. ns > 0.05, *<0.05; ***<0.001; nonsignificant p-values: 0.05 to 0.1; 0.1 to 0.3; 0.3 to 0.5; 0.5 to 0.7; 0.7 to 0.9; 0.9 to 1.0; 1.0 to 1.
1. Confounding effect of IS on the expression of genes

Array-wide Empirical Bayes moderated linear models analysis
LIMMA [26] + FDR a
IoT patients (Cohort 1)
IoT custom array RISET2.0 Agilent (5069 probes, 4607 genes)

RT-qPCR analysis of IoT signature
GAMBIT patients (Cohort 2, time point 1)

Proof of Concept
RT-qPCR analysis of IoT signature
GAMBIT patients (Cohort 3)

2. Effects of Drugs on Lymphocyte subset distribution

Flow cytometry
GAMBIT patients (Cohort 2)

Proof of Concept
Flow cytometry
GAMBIT patients (Cohort 3)

3. Immunosuppression-Independent Expression (IS-IE)

Multiple Linear Regression residual estimates b
IoT patients (Cohort 1)
RISET2.0 array gene expression (1989 probes post QC)

Gene Set Analysis
GSA [30] c

Feature selection using IS-IE
Regularized Multivariate Logistic Regression – Elastic Net
GLMNET [25] d
(28-gene selection)

Validation

Multiple Linear Regression residual estimates (IS-IE)
GAMBIT patients (Cohort 2)
Fluidigm RT-qPCR (26 validated array-selection genes)

Feature selection using IS-IE
Regularized Multivariate Logistic Regression – Elastic Net
GLMNET [25] e
(9-gene signature)
GAMBIT patients (Cohort 2, time point 1)

Evaluation of Signature Stability
(9-gene signature)
GAMBIT patients (Cohort 2, time point 2)

Figure 1: Analysis strategy to demonstrate confounding effect of IS and how to avoid it. A complete description can be found in Data S1. In summary: aProbes with nonsignificant expression above background in more than 20% of the samples were filtered out. The effects of three factors were estimated: CNI: (ciclosporin [Cyc], tacrolimus [Tac], none), MMF/Aza (mycophenolate mofetil, azathioprine, none) and prednisone-Pred-(Pred; on steroids, off steroids). bThe model for the ith patient and the jth Gene Set Analysis gene was

\[ y_{ij} = a_j + \beta_{Cyci} \times \text{Tac}_j + \beta_{Azip} + \text{MMFi} + \beta_{Predi} + e_{ij} \]

This was estimated using data from stable and chronic rejector patients. The resulting estimated residual \( e_{ij} \) represents the IS-independent gene expression (IS-IE). The gene expression of tolerant patients and healthy controls was rescaled to IS-IE by subtracting the intercept \( a_j \) from the raw expression. cGSA was used for the identification of differentially expressed biological pathways based on the curated list of gene sets from the Broad Institute (www.broadinstitute.org/gsea). Gene sets with an associated FDR below 10% were considered differentially expressed. dIn ElasticNet regression, a penalty is imposed on the regression coefficients, which is a combination of the penalties used in lasso and ridge regression. ElasticNet enables selection of genes (unlike ridge regression, which would preserve all genes) as well as gene groups irrespective of correlation (unlike lasso, which would select only one of a group of correlated genes). Model parameters were tuned using leave-group-out cross-validation with a 85% training set and 100 resampling iterations, with the AUC as an accuracy measure, via the caret package in R [29]. Prior to model estimation, missing values were imputed using K-nearest neighbors. Genes for which the expression was not significantly above background (p > 0.01) in at least 80% of the samples were filtered out prior to analysis (3081 out of 5070 probes) to increase statistical power (10). eClassification cutoffs were selected to ensure specificity above 0.85 while retaining sensitivity above 0.70. IS, immunosuppression; CNI, calcineurin inhibitors; Aza, azathioprine; MMF, mycophenolate mofetil; IS-IE, IS-independent expression; GLMNET, Lasso and Elastic-Net Regularized Generalized Linear Models; GSA, Gene Set Analysis; FDR, false discovery rate; AUC, area under the receiver operating characteristic curve; GAMBIT, Genetic Analysis and Monitoring of Biomarkers of Immunological Tolerance; IoT, Indices of Tolerance; RT-qPCR, reverse transcription quantitative real-time polymerase chain reaction; RISET, Reprogramming the Immune System for the Establishment of Tolerance.
array-wide analysis of drug effects using the expression measured on the original data from the IoT study (8). This revealed that 119 genes were differentially expressed in association with CNI drug intake and that 83 genes were associated with MMF and azathioprine intake, whereas only one gene was exclusively affected by steroid intake (Table S1).

To confirm the effect of IS, we then assessed the effect of IS on the expression of the previously published 10 genes of the signature using samples from stable patients on IS from cohort 2 (time point 1) of the GAMBIT study (8). A summary of the different IS regimens the patients were under is provided in Table S2. Prednisone and azathioprine showed statistically significant effects on the expression of 7 out of 10 of the previously described individual genetic markers of tolerance (Table 2). The effects of each drug were adjusted for the intake of other IS drugs.

To measure the magnitude of these effects, we calculated the percentage of variance in the expression of each gene explained by drug effects ($R^2$ values from linear regression models based on data from the GAMBIT cohorts 2 and 3; Table S4). For 5 genes among the 10 in the original IoT signature, the percentage of expression explained by drugs was at or higher than 10% and up to 27% ($CD79b$, $TCL1A$, $SH2DB1$, $FCRL1$, and $MS4A1$), further strengthening the argument that the expression of genes included in the IoT signature was influenced by IS drug regimens.

We then calculated the probability of tolerance for each patient based on the RT-qPCR expression in peripheral blood samples. We used the gene expression from the previously published IoT signature, as described (8) for patients from cohort 2, time point 1, of the GAMBIT study. We observed that this probability was undeniably and significantly associated with the drug exposure of the patients (Figure 2). The probability was significantly lower in stable patients treated with azathioprine compared to patients off antiproliferative ($p < 0.0001$) and patients on MMF ($p < 0.0001$) (Figure 2A). This probability was not influenced by the intake of CNIs once the effects of prednisone and azathioprine were accounted for (Figure 2B). Similar to azathioprine, the administration of prednisone was associated with a significantly lower estimated probability of tolerance ($p < 0.0001$) in stable patients in GAMBIT cohort 2 (Figure 2C; note that the patients shown in this plot are all off azathioprine).

To further confirm this observation, we used the gene expression from samples in the prospective GAMBIT cohort 3 ($p = 0.008$). Indeed, the estimated probability of tolerance significantly increased after steroid withdrawal (Figure 2D; please note that no patient in this group was receiving azathioprine).

The AUC for the probability of tolerance estimates from the comparison between tolerant and IS-treated patients from GAMBIT cohort 2, based on the IoT signature, was 0.89 (95% confidence interval [CI]: 0.83–0.94). When the AUC was adjusted for the effects of IS drugs, it became significantly lower: 0.77 (95% CI 0.67–0.86; $p = 0.032$). This provided further evidence for a confounding effect of IS regimens in the expression of these genes.

Therefore, we have demonstrated, in three completely independent cohorts (1, 2 and 3), evidence of drug confounding or bias in the gene expression of our previously identified tolerance signature.

**Immunosuppressants affect the transitional B cell subset**

Whole blood gene expression data are greatly influenced by the repertoire of circulating lymphocyte subsets. To

### Table 2: Effects of IS drugs on the published signature in stable patients from the GAMBIT study

<table>
<thead>
<tr>
<th>Gene Symbols</th>
<th>Pred</th>
<th>Cyc</th>
<th>Tac</th>
<th>Aza</th>
<th>MMF</th>
</tr>
</thead>
<tbody>
<tr>
<td>PNOC</td>
<td>0.11</td>
<td>0.10</td>
<td>0.04</td>
<td>0.76</td>
<td>1.00</td>
</tr>
<tr>
<td>CD79b</td>
<td>$2.1 \times 10^{-04}$</td>
<td>1.00</td>
<td>0.12</td>
<td>$8.1 \times 10^{-04}$</td>
<td>0.94</td>
</tr>
<tr>
<td>TCL1A</td>
<td>$1.9 \times 10^{-06}$</td>
<td>0.17</td>
<td>0.02</td>
<td>$6.7 \times 10^{-16}$</td>
<td>1.00</td>
</tr>
<tr>
<td>H3ST1</td>
<td>$1.3 \times 10^{-04}$</td>
<td>0.30</td>
<td>0.14</td>
<td>$3.6 \times 10^{-05}$</td>
<td>0.20</td>
</tr>
<tr>
<td>SH2DB1</td>
<td>0.42</td>
<td>1.00</td>
<td>1.00</td>
<td>$&lt;2.0 \times 10^{-16}$</td>
<td>0.11</td>
</tr>
<tr>
<td>TLR5</td>
<td>$4.0 \times 10^{-03}$</td>
<td>1.00</td>
<td>0.09</td>
<td>1.00</td>
<td>1.00</td>
</tr>
<tr>
<td>MS4A1</td>
<td>$3.0 \times 10^{-03}$</td>
<td>0.73</td>
<td>0.18</td>
<td>$1.1 \times 10^{-04}$</td>
<td>1.00</td>
</tr>
<tr>
<td>FCRL1</td>
<td>$1.7 \times 10^{-04}$</td>
<td>1.00</td>
<td>0.73</td>
<td>$1.1 \times 10^{-10}$</td>
<td>1.00</td>
</tr>
<tr>
<td>FCRL2</td>
<td>$5.7 \times 10^{-04}$</td>
<td>1.00</td>
<td>0.15</td>
<td>$1.6 \times 10^{-05}$</td>
<td>1.00</td>
</tr>
<tr>
<td>FoxP3/AMann</td>
<td>0.69</td>
<td>0.16</td>
<td>$9.0 \times 10^{-03}$</td>
<td>1.00</td>
<td>1.00</td>
</tr>
</tbody>
</table>

p-values for comparisons of stable patients on and off each drug are derived after adjustment in a linear regression model for all other drugs/drug groups. The p-values for the Cyc/Tac group (subgroups No-Cyc/Tac, Cyc, and Tac) and for the Aza/MMF group (subgroups No-Aza/MMF, Aza, and MMF) were adjusted for multiple comparisons with Bonferroni correction.

Aza, azathioprine oral dose; Cyc, cyclosporine A trough levels; GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance; IS, immunosuppression; MMF, mycophenolate mofetil oral dose; Pred, prednisone oral dose; Tac, tacrolimus trough levels.
determine whether the previously described association of tolerance with a relative increase in circulating transitional B cells in tolerant recipients might also be confounded by IS, we performed flow cytometry analysis of peripheral blood cells from the patients included in GAMBIT cohort 2 (for gating, see Figure S2). The effect of IS drugs was clearly evident in the percentage of transitional B cells within the naive B cell population (Figure 3), and this effect was similarly observed when measuring absolute numbers of cells in a subset of the recipients (Figure S3). Notably, the pattern of the changes closely resembled the effect of IS drugs on the estimated probability of tolerance based on the IoT signature. Stable patients on azathioprine (Figure 3A) and those on prednisone (Figure 3C) showed lower percentages of transitional B cells than patients off each of these drugs, whereas CNI drugs showed no effect (Figure 3B). The effect of prednisone showed a clear dose–response relationship (Figure 3D), and this was confirmed in the prospective steroid withdrawal GAMBIT cohort 3.
The percentage of total B cells in periphery significantly decreased only for those patients taking azathioprine (data not shown). Changes in the opposite direction were observed in the percentages of T cells from lymphocytes. These were increased in stable patients taking azathioprine or prednisone but were unaffected by CNI drugs (Figures S1A, B and D). However, the evidence for a dose–response effect of prednisone was very weak, and there were no differences in the percentages of peripheral blood T cells in the prospective steroid withdrawal cohort 3 (Figures S1E and F).

**Development of a novel immunosuppression-independent gene expression signature of tolerance**

Having demonstrated that the IS drug regimen is a confounder for the association between gene expression levels and the predicted probability of tolerance, we concluded that for a predictive test of tolerance to be clinically applicable and unbiased, IS effects on gene expression must be accounted for in the predictive algorithm. We used the array data from the IoT study (cohort 1) as a “discovery” set. We obtained the residuals of a multivariate linear regression model for IS drugs per each gene in the array. We used this to estimate the IS-independent gene expression (IS-IE) per gene.

We used the estimated IS-IE to select an optimal set of genes predictive of tolerance. To enable applicability of the signature in a clinical setting, we restricted the selection to a maximum of 30 genes. The resulting set of 28 IS-independent genes provided excellent predictive
The accuracy of the new signature, as evaluated by the AUC, was 0.93 (95% CI: 0.86–1.0) and 0.81 after optimism correction via cross-validation. A classification cutoff of 0.54 was selected to maximize specificity for patient safety (sensitivity of 0.92, specificity of 0.88).

Table 3 shows the validated IS-IE nine-gene list. Please note that drugs explained very little of the variance of their expression (see R² in Table S5). No gene overlapped with the previous IoT signature.

The new IS-IE estimated probability of tolerance was independent of IS regimen—and therefore unconfounded—in stable patients (p > 0.05 for all drugs) from GAMBIT cohort 2 (Figures 5A, B and C). Further, in the prospective GAMBIT cohort 3, we could demonstrate that steroid withdrawal does not affect the expression of this new gene signature (Figure 5D).

Figure 4: Discovery of a new set of IS-independent markers of tolerance. (A) Heat map showing patterns of residual gene expression from 28 genes selected using ElasticNet on array data from the IoT cohort, comparing tolerant versus immunosuppressed recipients. (B) Predictive accuracy of multivariate predictive sets of genes. GLMNET refers to the 28-gene set discovered via ElasticNet analysis. The remaining sets were selected from those molecular pathways shown to be differentially expressed via gene set analysis. AUC, area under the receiver operating characteristic curve; CV-AUC, cross-validated AUC; IoT, Indices of Tolerance; Sens, sensitivity; Spec, specificity; GLMNET, Elastic Net Regularized General Linear Model; REGGR, Glucocorticoid receptor regulatory network; TNF, Tumor Necrosis Factor Pathway; CD40, CD40L Signaling Pathway; NFκB, NF-kappaB Signaling Pathway; GMCSF, GMCSF-mediated signaling events.

Validation of the new signature on Fluidigm platform in samples from the GAMBIT study
We chose to validate and further refine the new IS-IE gene set using the Fluidigm platform, an RT-qPCR-based assay, in samples from patients in GAMBIT cohort 2 (time point 1). Quality control criteria were met by 26 genes.

A set of 9 genes out of the 26 was selected by ElasticNet as optimal to predict tolerance (see Data S1). Table 3 shows the validated IS-IE nine-gene list. Please note that drugs explained very little of the variance of their expression (see R² in Table S5). No gene overlapped with the previous IoT signature.
Based on the IS-IE signature, 20 out of 173 patients from cohort 2 on IS with stable function were identified as “probably tolerant” (11.6%). These patients were different individuals from those identified with the IoT signature (n = 25) (only two overlapping). Comparison of the two groups (Tables S6 and S7; Data S1) revealed, importantly, that the IS-IE selection had significantly longer time posttransplantation, making them more comparable to the index group of tolerant patients.

Evaluation of the stability of the signature developed in cohort 1 (by estimating its predictive accuracy in two time points in cohort 2) showed satisfactory performance (AUC time point 2 = 0.83; 95% CI: 0.67–0.99). Additionally, a nonsignificant McNemar’s test indicated that binary classification is stable across repeated samples (p = 0.095), meaning that classifications at one time point are not significantly different from those at a subsequent time point. Furthermore, the continuous predicted probability of tolerance does not change significantly between time points (Figure S4).

The genes included in the new signature participate in cellular pathways such as regulation of nucleic acid metabolism (GEMINI7, NFκB1A, TNFIP3), cell communication activities (RAB40C) and transcription factor activity (EEF1A1, NFκB1A, TNFIP3). Still, the protein expression derived from three of the genes has been observed in B cells and in other cells (BCL2A1, EEF1A1) or in B cells exclusively (IGLC1; see Table 3).

Based on the IS-IE signature, 20 out of 173 patients from cohort 2 on IS with stable function were identified as “probably tolerant” (11.6%). These patients were different individuals from those identified with the IoT signature (n = 25) (only two overlapping). Comparison of the two groups (Tables S6 and S7; Data S1) revealed, importantly, that the IS-IE selection had significantly longer time posttransplantation, making them more comparable to the index group of tolerant patients.

Evaluation of the stability of the signature developed in cohort 1 (by estimating its predictive accuracy in two time points in cohort 2) showed satisfactory performance (AUC time point 2 = 0.83; 95% CI: 0.67–0.99). Additionally, a nonsignificant McNemar’s test indicated that binary classification is stable across repeated samples (p = 0.095), meaning that classifications at one time point are not significantly different from those at a subsequent time point. Furthermore, the continuous predicted probability of tolerance does not change significantly between time points (Figure S4).

The genes included in the new signature participate in cellular pathways such as regulation of nucleic acid metabolism (GEMINI7, NFκB1A, TNFIP3), cell communication activities (RAB40C) and transcription factor activity (EEF1A1, NFκB1A, TNFIP3). Still, the protein expression derived from three of the genes has been observed in B cells and in other cells (BCL2A1, EEF1A1) or in B cells exclusively (IGLC1; see Table 3).

**The new IS-independent signature is differentially expressed in healthy controls**

An important limitation of the previously described signatures is their inability to differentiate between tolerant recipients and healthy controls (as illustrated in Figure 6A for the IoT signature). Similarly, we have observed no difference in the transitional B cell percentages in peripheral blood between these two groups (Figure 6B). Importantly, the predicted probability of tolerance based on the new signature was higher in tolerant patients compared to healthy controls (Figure 6C). This suggests that the method proposed herein and the resulting novel signature indeed capture an underlying predisposition to tolerance rather than the absence of IS drugs. Comparisons of the predicted probabilities of tolerance in tolerant patients and healthy controls with the corresponding cutoffs for the
IoT and the IS-IE signatures are shown in Table S8 and are discussed in Data S1.

**Discussion**

A number of studies, including our own, have suggested gene expression signatures of tolerance in kidney transplantation (7–9,11,32) or have described differential expression of smaller gene sets (11,33,34). None had assessed the confounding effect of IS. Baron et al (10) recently completed a comprehensive review of all public data and concluded that the expression of a set of 20 genes—mostly expressed by B cells—may be used as a standardized tool for personalized medicine in KTRs. Four genes from our IoT signature (TCL1A, MS4A1, FCRL2, and CD79B) were included in this 20-gene set, but we now demonstrate that their expression is highly affected by azathioprine and prednisone. Similarly, it has been shown, in an independent cohort, that azathioprine affects the three-gene signature proposed by the Immune Tolerance Network (35). We have shown in our results, using the published signature of tolerance, that IS confounds the assessment of tolerance in tissue biopsy samples. The TOL expression was compared across different groups and in paired samples pre- and post-steroid withdrawal. The p-values for each statistical comparison were derived after adjustment in a linear regression model for all other drugs/drug groups. The p-values for CNI drugs and for antiproliferative drugs were adjusted for multiple comparisons with Bonferroni correction. The p-values for comparisons pre- and post-steroid withdrawal were derived from a Wilcoxon matched pairs test. Cyc, ciclosporin; Tac, tacrolimus; Pred, prednisone/prednisolone; Aza, azathioprine; IS, immunosuppression; IoT, Indices of Tolerance study; GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance; MMF, Mycophenolate Mofetil; CNI, Calcineurin Inhibitors. Probability of tolerance cutoff was 0.54.
KTRs who were maintained off steroids, on MMF and on tacrolimus were being identified more frequently as possible tolerant recipients.

Therefore, we asked, “Was this tolerance or response to immunosuppressive therapy?” We have now demonstrated that it is highly likely that it was the latter. Consequently, we have developed and validated a new noninvasive gene expression signature of tolerance that is independent of IS drug effects and that additionally differentiates tolerant patients from healthy controls. This dictates that further analysis of tolerance signatures using the correction proposed herein (or a similar method) needs to be considered.

Additionally, we have demonstrated that the percentages of transitional B cells in peripheral blood, which had been described as characteristic of tolerant recipients, were also significantly affected by IS drugs. We have not addressed the specific mechanism by which each immunosuppressant affects the intracellular pathways in individual lymphocyte subsets, as this would require longer-term studies and is beyond the scope of the current one. However, commonly used immunosuppressants, such as CNIs, which are fundamentally aimed at inhibiting T cell activation and have only an indirect effect on B cell activation, consistently showed little effect on gene expression, the percentage of transitional B cell subset size or the estimated probability of tolerance in our study after adjustment for intake of other IS drugs. When specifically addressed, transitional B cells exhibit the capacity to decrease anti-inflammatory responses and produce anti-inflammatory cytokines (21,36). Therefore, we do not question the important functionality that transitional B cells may play in transplantation tolerance, but we believe that the evidence of the role of B cells in tolerance needs further scrutiny, particularly regarding the immunosuppression effects.

While there is no clear major genetic pathway connecting the genes included in the new IS-independent signature, literature reports suggest that at least some of these genes have a mechanistic relevance to tolerance. For example, a polymorphism in the NFκBIA gene resulting in upregulation has been associated with higher rates of acute liver transplant rejection (37), and BCL2A1 has been shown to be a transcriptional target for NFκB (38). In our study, both genes were downregulated in tolerant patients. IGLC1 has been included in the expansion of the B cell signature of tolerance in KTRs (39). Upregulation of MS4A4A and RAB40C (upregulated in tolerant patients in our study) has been associated with macrophage activation (40,41). TNFAIP3 (A20) is an NFκB regulatory protein and its expression has been associated with outcome prediction in kidney transplantation (42), but the regulation of its expression and function in inflammatory responses has been shown to be complex (43,44). The association found herein of the downregulation of this gene with operational tolerance merits further investigation.

External validation of the presented signature in other independent cohorts would strengthen the confidence in the generalizability of the results and would allow final calibration before translation into clinical practice. Such studies would require the collection of detailed clinical phenotype data in parallel to the gene expression data.
In conclusion, this study emphasizes the importance of assessing and correcting for the effect of diverse IS regimens on gene expression–based biomarker signatures. Using this correction, we identify a novel—and, arguably, a more clinically robust—signature of operational tolerance, which we have validated in independent and extensive cohorts of KTRs. Moreover, in our prospective validation cohort, the estimated probability of tolerance remained unchanged after steroid withdrawal, supporting the view that the new signature highlights natural counter-regulatory mechanisms and excludes transient alterations of the immune effector pathways by IS drugs. Further evidence that our approach is uncovering tolerance-related responses is the fact that the estimated probability of tolerance in tolerant patients is higher than that of healthy controls, in agreement with studies demonstrating the involvement of an active immune response in tolerance (47).

We are aware that these results do bring into question previous published evidence, and we have demonstrated the effect of IS drugs on our own published signature. Transplant physicians will require confidence in any novel clinical-grade biomarker set of kidney transplantation tolerance, such as the new one described in this article, prior to embarking on clinical trials of IS weaning or minimization. Such trials are critical if we aim to reduce cancer risk and increase long-term survival with improved quality of life for KTRs. In the current climate of stratified medicine, these findings may also be relevant to autoimmune diseases and other disorders in which IS is a prevalent drug therapy.

Acknowledgments

The authors acknowledge financial support from the Medical Research Council (MRC) (grant G0801537/1:D: 88245), the MRC Centre for Transplantation (MRC grant MR/J006742/1) and Guy’s and St. Thomas’ Charity (grants R080530 and R090782). E.N.-L. was funded by a scholarship from CONICYT Bicentennial Becas-Chile, Chile. Authors M.P.H.F., I.R.-M., and S.C. were funded by EU project BIO-DrIM (FP7-HEALTH-2012-INNOVATION-1 project number 305147). M.P.H.F. received funding from the European Union, Seventh Framework Programme (FP7/2007–2013), under grant agreement HEALTH-F5–2010–260687: The ONE Study. The research was funded and supported by the National Institute for Health Research (NIHR) Biomedical Research Centre based at Guy’s and St. Thomas’ National Health System (NHS) Foundation Trust and King’s College London. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. All UK-based centers received service support through Clinical Research Networks, study portfolio 7521. This allowed the support of a large number of research nurses in different centers, whose dedication allowed the samples and clinical information to be collected. The authors are in great debt to Florence Delaney for data management and organizational support. Thomas Lewis and Akiko Tsutsui were responsible for project management and Andy Thomason for English review. We would also like to thank the patients and donors who graciously donated their samples and time to this project. The array data from the Indices of Tolerance study were deposited in National Centre for Biotechnology Information’s Gene Expression Omnibus (48) and are accessible through Gene Expression Omnibus Series accession GSE14655 (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE14655).

Author Contributions

I.R.-M. performed all the statistical analysis, wrote the main part of the manuscript and contributed to the acquisition of funds. E.N.-L. worked specifically on the B cell flow cytometry studies and contributed to the review of the manuscript. P.M., M.R., S.N., Y.K., and N.S. significantly contributed to the experiments and experiment planning. S.C. contributed to statistical analysis and the writing of the manuscript. R.H., S.B., R.B., D.B., S.C., D.G., S.G., R.L., P.K., P.B.M., S.M., I.M., W.M., M.M., R.P., S.P.K., D.S., M.D.S., B.T., and O.V. contributed the clinical and follow-up data of the patients and contributed to the review of the manuscript. R.I.L. and G.M.L. partially contributed to the funding provisions, mentoring, project overview and review of the manuscript. M.P.H.F. designed the study, obtained funding, managed the team, and contributed to and supervised the writing of the manuscript.

Disclosure

The authors of this manuscript have no conflicts of interest to disclose as described by the American Journal of Transplantation.

References

Rebollo-Mesa et al


Supporting Information

Additional Supporting Information may be found in the online version of this article.

Data S1: Materials and methods.

Table S1: Genes whose expression is affected by immunosuppressive drugs (Excel file attached at the end).

Table S2: Distribution of drug regimen in stable patients.

Table S3: Detailed clinical description of GAMBIT tolerant patients. GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance.

Table S4: Assay list used for the RT-qPCR test of the Indices of Tolerance gene list. RT-qPCR, reverse transcription quantitative real-time polymerase chain reaction.

Table S5: Gene list (reliable signal) with assay description used in the Fluidigm platform for ElasticNet selection of the immunosuppression-free signature.

Table S6: Clinical and demographic characteristics of stable patients from the GAMBIT study classified as tolerant with the IoT and IS-IE signatures (classification match only in two patients). GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance; IoT, Indices of Tolerance; IS-IE, Immunosuppression-independent expression.

Table S7: Effects of IS drugs on predicted probability of tolerance according to the IoT and IS-IE signatures in stable patients from the GAMBIT study. IS, Immunosuppression; IoT, Indices of Tolerance; IS-IE, Immunosuppression independent gene expression: GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance.

Table S8: Predicted probability of tolerance in tolerant patients and healthy controls in relation to the cutoff.

Figure S1: Percentage of T cells (CD3+ cells within the live lymphocyte gate) in peripheral blood of patients from the GAMBIT cohort. GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance.

Figure S2: Gating strategy for transitional B cells in the flow cytometer.

Figure S3: Absolute number of CD24hiCD38hi (transitional B cells) within the live CD20+ CD19+ B lymphocytes and CD27-IgD+IgM+ gate in peripheral blood of patients from the GAMBIT study, cohort 2. GAMBIT, Genetic Analysis of Molecular Biomarkers of Immunological Tolerance.

Figure S4: Estimated probability of tolerance is stable over time.