Heterogeneity of respiratory disease in children and young adults with sickle cell disease

Alan Lunt¹, Lucy Mortimer¹, David Rees², Sue Height², Swee Lay Thein²,³, Anne Greenough¹,⁴

¹Division of Asthma, Allergy and Lung Biology, MRC & Asthma UK Centre in Allergic Mechanisms of Asthma, King's College London, London, United Kingdom

² Department of Paediatric Haematology, King’s College Hospital NHS Foundation Trust, London, UK

³ Molecular Haematology, Division of Cancer Studies, King’s College London, UK.

⁴NIHR Biomedical Research Centre at Guy’s and St Thomas’ NHS Foundation Trust and King's College London, United Kingdom

Address for correspondence: Professor Anne Greenough, Neonatal Intensive Care Unit, 4th Floor Golden Jubilee Wing, King’s College Hospital, Denmark Hill, London, SE5 9RS, United Kingdom. Tel: 0203 299 3037; fax: 0203 299 8284

Email: anne.greenough@kcl.ac.uk

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ABSTRACT

To detect and characterise different phenotypes of respiratory disease in children and young adults with sickle cell disease (SCD), eleven lung function and haematological biomarkers were analysed using k-means cluster analysis in a cohort of 114 SCD subjects aged between 5 and 27 years. Three clusters were detected: cluster one had elevated pulmonary capillary blood volume, mixed obstructive/restrictive lung disease, hypoxia and moderately severe anaemia; cluster two were older patients with restrictive lung disease; cluster three were younger patients with obstructive lung disease, elevated serum lactate dehydrogenase and bronchodilator reversibility. These results may inform more personalised management strategies to improve outcomes.
INTRODUCTION

Sickle cell disease (SCD) is one of the commonest inherited disorders worldwide, affecting an estimated 300,000 new-borns every year. The majority of SCD children in developed countries will survive to adulthood, but may then suffer multiorgan damage, including pulmonary complications. Whilst lung function abnormalities are common in both children and adults, there is not a consistent picture. Furthermore, the aetiology and clinical consequences of the lung function abnormalities are unclear. It seems likely that the respiratory disease is not homogenous [1]. Identification of different phenotypes of respiratory disease may provide a basis to stratify patients with regard to management. Cluster analysis is a method of detecting subgroups within multidimensional (i.e. comprising many and/or varied measurements) datasets.[2] We hypothesised that cluster analysis of respiratory function and haematological biomarkers would reveal different phenotypes in patients with SCD.

METHODS

Study design

A secondary analysis of data from children and young adults with sickle cell anaemia (SCA) (homozygous for sickle cell haemoglobin (HbSS)) and ages ranging from 5 to 27 years was conducted.[3-5] The studies were approved by King’s College Hospital Research Ethics Committee and participants or their parents (in the case of children) gave informed written consent to take part.

Lung function measurements (see online supplement)

Respiratory system resistance at a frequency of 5Hz ($R_{ns5}$) was measured, then
spirometry (FEV1:VC), static lung volumes using whole-body plethysmography (TLC) and gas transfer for carbon monoxide (DLCO) were assessed according to American Thoracic Society/European Thoracic Society criteria.[6-8] The transfer factor for nitric oxide (DLNO) was obtained by the addition of 40ppm NO to the inspired gas mixture prior to commencing the single-breath DLCO measurement and used to estimate the pulmonary capillary blood volume (PCBV).[3] Arterial oxygen saturation (SpO2) was measured using pulse oximetry (Masimo Radical 7, Masimo, CA).

**Haematological data and SCD complications**

The haemoglobin concentrations (Hb), lactate dehydrogenase (LDH) levels, reticulocyte percentage (reticulocyte%) and white cell counts (WCC) were obtained from routine blood tests undertaken within three months of lung function testing. The medical records were reviewed and an ACS episode was diagnosed if the child had suffered chest pain, dyspnoea and pyrexia together with a new pulmonary infiltrate on chest radiograph. To determine the impact of cluster membership on future morbidity, the medical records were reviewed for the period subsequent to the lung function testing and the occurrence of individual ACS episodes and non-elective hospital admissions were recorded and expressed as events/year.

**Statistical analysis**

Variables included in the cluster analysis were FEV1:VC, TLC, Rrs, DLCO, PCBV, [Hb], SpO2, LDH, WCC, reticulocyte% and age at time of testing. The number of clusters was selected using the CritCF index [9] which ranks partitions comprising different numbers of clusters; a higher CritCF indicates a better clustering.
RESULTS

Data from 114 patients with complete lung function results and a median age 14.5 (5.0-27.1) years were analysed (see online supplement Table A). A three-cluster solution was found to be optimal (see online supplement Figure S1). The clustering solution was robust under multiple imputations with the same three-cluster partition occurring in 96.4% of imputed datasets (see online supplement Figure S1). For all lung function and haematological variables, the proportion of patients with obstructive, restrictive or mixed lung function defects and the prevalence of bronchodilator reversibility, but not the proportion taking hydroxyurea, receiving regular transfusions or with a history of ACS episodes, differed significantly between clusters (Table 1). Clusters were well separated, with little overlap on a discriminant coordinates plot (see online supplement Figure S2). Physiological profiles for the three clusters are shown graphically (see online supplement Figure S3).

Cluster one had the largest proportion with mixed obstructive/restrictive lung disease and both the highest respiratory system resistance and pulmonary capillary blood volume. Cluster two comprised older patients with the highest FEV₁:VC ratio and a low TLC (i.e. restrictive pattern), the lowest gas transfer gas transfer, but highest haemoglobin. Cluster three comprised younger patients with the highest incidence of bronchodilator reversibility and the highest LDH levels (Table 1). Cluster three has a greater frequency of hospital admissions for vaso-occlusive crises, but not ACS episodes, than clusters one and two (both p<0.05) see online supplement.

A conditional inference tree analysis was performed on the subset of patients assigned to the training set (n=85). A model using three variables (PCBV, DLCO and LDH) (Figure 1) was found to be optimal and assigned 90% of patients from the unseen validation set to the
correct cluster. Classification success rates for clusters one, two and three were 80%, 93% and 90%, respectively.

DISCUSSION

Analysis of lung function and haematological data detected three distinct clusters in children and young adults with SCD. Cluster one had the highest respiratory system resistance and pulmonary capillary blood volume which suggest they may have peripheral airways disease mediated by pulmonary vascular engorgement. We speculate that this interaction may arise from direct compression of distal airways by adjacent pulmonary vessels within peribronchial sheaths or from bronchovascular congestion due to elevated pulmonary venous pressure. Haemoglobin levels were significantly higher in cluster two which suggests that their restrictive lung disease might be related to increased blood viscosity leading to impaired microvascular blood flow and an increased likelihood of occult vaso-occlusion in the pulmonary capillary plexus with consequent cumulative lung injury. Cluster three had the highest LDH levels and the highest incidence of bronchodilator reversibility. Those findings are consistent with the observation that airway hyper-reactivity, as assessed by methacholine challenge testing, was associated with elevated LDH levels.[10] Serum LDH, but not reticulocyte count or haemoglobin concentration was discriminative for cluster three. This may reflect LDH had a greater sensitivity as a marker of haemolysis. LDH, however, is thought to primarily reflect intravascular haemolysis, thus in cluster three compared to the other two clusters perhaps a greater proportion of overall haemolytic activity was occurring in the intravascular compartment.

Conditional tree analysis demonstrated that a subset of the variables could reliably predict cluster membership. This suggests that a method for phenotyping of SCD respiratory subclasses could be based on those biomarkers.
ACKNOWLEDGEMENTS

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Authors contributions: AL and AG designed the study, AL, LM, DR and SH collected the data. AL analysed the data. All authors were involved in the development of the manuscript and approved the final version.

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REFERENCES


Table 1 Subject characteristics by cluster.

*Lung function tests are expressed as the percentage predicted for age and/or height.

†Supplementary data not used in the clustering.

<table>
<thead>
<tr>
<th></th>
<th>Cluster 1 (n = 14)</th>
<th>Cluster 2 (n = 55)</th>
<th>Cluster 3 (n = 45)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>FEV₁:VC *</td>
<td>92.4 (88.05 – 100.9)</td>
<td>97.3 (91.1 – 102.6)</td>
<td>90.7 (85.9 – 96.2)</td>
<td>0.002</td>
</tr>
<tr>
<td>Rs5 *</td>
<td>165.8 (140.8 – 186.5)</td>
<td>131.6 (112.9 – 160.3)</td>
<td>133.4 (110.2 – 153.6)</td>
<td>0.020</td>
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<td>TLC *</td>
<td>84.8 (82.2 – 86.8)</td>
<td>85.6 (77.4 – 91.6)</td>
<td>94.1 (85.7 – 99.6)</td>
<td>0.003</td>
</tr>
<tr>
<td>DLCO *</td>
<td>98.1 (92.5 – 106.2)</td>
<td>77.8 (72.2 – 83.8)</td>
<td>96.6 (88.9 – 108.3)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>PCBV (ml/L)</td>
<td>44.9 (41.4 – 47.0)</td>
<td>21.1 (17.9 – 23.5)</td>
<td>26.4 (23.4 – 29.7)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>[Hb] (g/dl)</td>
<td>8.2 (7.4 – 8.9)</td>
<td>9.9 (8.7 – 10.8)</td>
<td>8.4 (5.7 – 9.2)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>SpO₂ (%)</td>
<td>95.8 (95.0 – 97.8)</td>
<td>97.0 (95.8 – 99.0)</td>
<td>95.5 (93.0 – 98.0)</td>
<td>0.0327</td>
</tr>
<tr>
<td>LDH (IU/L)</td>
<td>495 (441 – 622)</td>
<td>399.5 (303.5 – 474.8)</td>
<td>639 (542 – 732.0)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>WCC (x10⁹/L)</td>
<td>10.7 (10.3 – 12.4)</td>
<td>9.2 (7.3 – 10.8)</td>
<td>11.0 (8.8 – 12.3)</td>
<td>0.015</td>
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<tr>
<td>Reticulocyte%</td>
<td>12.7 (10.6 – 14.2)</td>
<td>8.5 (6.5 – 10.5)</td>
<td>10.4 (7.7 – 12.8)</td>
<td>0.018</td>
</tr>
<tr>
<td>Age (yrs)</td>
<td>14.2 (10.3 – 17.0)</td>
<td>17.5 (13.3 – 21.6)</td>
<td>11.3 (9.0 – 14.5)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Obstructive†</td>
<td>0%</td>
<td>9.1%</td>
<td>26.7%</td>
<td>0.027</td>
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<tr>
<td>Restrictive†</td>
<td>14%</td>
<td>29.1%</td>
<td>4.4%</td>
<td>0.023</td>
</tr>
<tr>
<td>Mixed†</td>
<td>21%</td>
<td>5.5%</td>
<td>4.4%</td>
<td>0.028</td>
</tr>
<tr>
<td>ACS ever†</td>
<td>35.7%</td>
<td>33.3%</td>
<td>33.3%</td>
<td>0.9231</td>
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<td>BDR %†</td>
<td>7.1%</td>
<td>2.0%</td>
<td>15.6%</td>
<td>0.0338</td>
</tr>
<tr>
<td></td>
<td>14.3%</td>
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<tr>
<td><strong>Hydroxyurea</strong></td>
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<td></td>
</tr>
<tr>
<td>%</td>
<td>7.1%</td>
<td>20.0%</td>
<td>31.1%</td>
<td>0.1633</td>
</tr>
</tbody>
</table>

† BDR: bronchodilator reversibility.
FIGURE LEGENDS

Figure 1: Stratification algorithm to predict cluster membership