Local activation time sampling density for atrial tachycardia contact mapping: how much is enough?

Steven E. Williams1*, James L. Harrison1, Henry Chubb1, John Whitaker1, Radek Kiedrowicz2, Christopher A. Rinaldi2, Michael Cooklin2, Matthew Wright1, Steven Niederer1, and Mark D. O’Neill1

1Division of Imaging Sciences and Biomedical Engineering, King’s College London, 4th Floor North Wing, St Thomas’ Hospital, 249 Westminster Bridge Road, SE1 7EH London; and 2Department of Cardiology, Guy's and St Thomas’ NHS Foundation Trust, St Thomas’ Hospital, 249 Westminster Bridge Road, SE1 7EH London, UK

Received 14 June 2016; editorial decision 1 February 2017; accepted 12 February 2017; online publish-ahead-of-print 3 April 2017

Aims

Local activation time (LAT) mapping forms the cornerstone of atrial tachycardia diagnosis. Although anatomic and positional accuracy of electroanatomic mapping (EAM) systems have been validated, the effect of electrode sampling density on LAT map reconstruction is not known. Here, we study the effect of chamber geometry and activation complexity on optimal LAT sampling density using a combined in silico and in vivo approach.

Methods and results

In vivo 21 atrial tachycardia maps were studied in three groups: (1) focal activation, (2) macro-re-entry, and (3) localized re-entry. In silico activation was simulated on a 4x4cm atrial monolayer, sampled randomly at 0.25–10 points/cm² and used to re-interpolate LAT maps. Activation patterns were studied in the geometrically simple porcine right atrium (RA) and complex human left atrium (LA). Activation complexity was introduced into the porcine RA by incomplete inter-caval linear ablation. In all cases, optimal sampling density was defined as the highest density resulting in minimal further error reduction in the re-interpolated maps. Optimal sampling densities for LA tachycardias were 0.67 ± 0.17 points/cm² (focal activation), 1.05 ± 0.32 points/cm² (macro-re-entry) and 1.23 ± 0.26 points/cm² (localized re-entry), P = 0.0031. Increasing activation complexity was associated with increased optimal sampling density both in silico (focal activation 1.09 ± 0.14 points/cm²; re-entry 1.44 ± 0.49 points/cm², P < 0.0001) and in vivo (porcine RA pre-ablation 0.45 ± 0.13 vs. post-ablation 0.78 ± 0.17 points/cm², P = 0.0008). Increasing chamber geometry was also associated with increased optimal sampling density (0.61 ± 0.22 points/cm² vs. 1.0 ± 0.34 points/cm², P = 0.0015).

Conclusion

Optimal sampling densities can be identified to maximize diagnostic yield of LAT maps. Greater sampling density is required to correctly reveal complex activation and represent activation across complex geometries. Overall, the optimal sampling density for LAT map interpolation defined in this study was ~1.0–1.5 points/cm².

Keywords

Electroanatomic mapping systems • Atrial arrhythmias • Sampling density • Local activation time mapping

Introduction

Local activation time (LAT) mapping forms the cornerstone of atrial tachycardia diagnosis. The creation of LAT maps by electroanatomic mapping (EAM) systems involves four key steps: (1) creation of a three dimensional anatomy model, (2) analysis of intracardiac electrograms to identify activation times relative to a fixed temporal reference, (3) interpolation of the activation times across the anatomical model, and (4) representation of the interpolated data, usually in the form of a colour map. Anatomic accuracy of LA EAM chamber models has been compared to cross sectional imaging...
What’s new

- Electroanatomic mapping advances allow many hundreds, or possibly thousands, of activation time points to be collected within a single local activation time (LAT) map.
- Whether such an exponential increase in LAT sampling density results in similar increases in LAT map accuracy is not known.
- This manuscript defines the optimal sampling density for atrial tachyarrhythmias as the maximal density beyond which further sampling produces minimal further improvement in interpolated LAT map accuracy.
- Optimal sampling densities are thereby determined for a variety of tachycardia mechanisms, and are found to be in the range of 1.0–1.5 points/cm².
- Therefore, we propose that when a diagnosis is not reached at this density, further sampling may be less likely to improve map accuracy than a change of mapping strategy (e.g. a different window of interest, reference electrogram or LAT assignment technique).

Methods

In vivo procedures

A total of 37 electroanatomic maps were included in the analysis. Isochronal local activation time maps were created using the Carto3 mapping system (Biosense Webster, Diamond Bar, CA, USA) by taking the activation time at each acquired point as the time from reference activation (typically an atrial coronary sinus electrogram) to the earliest rapid deflection on the bipolar mapping catheter signal.

Mapping data was analysed from 21 patients undergoing clinically-indicated ablation for left atrial tachycardia. Cases with high quality maps (defined as maps with complete LA coverage) were identified retrospectively and all identified cases were included in the analysis. Following transseptal puncture, a 20-pole circular mapping catheter (Lasso Nav, Biosense Webster, Diamond Bar, CA, USA) was used to create a 3D geometry of the left atrium. High-density local activation time maps were constructed with the multipolar mapping catheter during tachycardia. Local activation times were automatically assigned and all points were re-annotated offline to ensure correct identification of the first component of each electrogram. Correct tachycardia diagnosis was confirmed by LAT map appearance, response to entrainment manoeuvres and response to ablation. Informed written consent was obtained from all patients for invasive procedures.

Animal studies complied fully with Danish law on animal experiments. Eight Götttingen mini-pigs (41.2 ± 7.2 kg) were pre-sedated with intramuscular azaperone (4 mg/kg) and midazolam (0.5 mg/kg). General anaesthesia was induced with intravenous ketamine (5 mg/kg) and midazolam (0.5 mg/kg). Anaesthesia was maintained with a continuous intravenous infusion of propofol (3 mg/kg/h) and fentanyl (15 μg/kg/h). Two 8F sheaths were placed percutaneously in the right femoral vein, followed by an intravenous injection of 100 IU/kg heparin. Fluoroscopy was used to position a 6F decapolar reference catheter in the coronary sinus (CS) and a 7.5F ablation catheter was advanced to the RA and used to create a LAT map during proximal CS pacing. Incomplete inter-caval linear ablation (confirmed with pace mapping) was performed, with a deliberate 5 mm gap left between cranial and caudal aspects of the lesion, before repeat LAT mapping of the new activation detour was performed.

Atrial monolayer model

A 4 x 4 cm atrial tissue model was meshed using triangular elements (84 346 triangles, 42 511 nodes, average edge length Δ1=0.2 mm). To simulate electrical activation of atrial tissue by an extracellular stimulus (1000 mV, 1 ms duration), the period of electrical activation was modelled using the bi-domain equations given by:

\[
\nabla \cdot (\sigma \nabla \phi) = \beta I_m
\]

\[
\nabla \cdot (\sigma_e \nabla \phi_e) = - \beta I_m
\]

\[
I_m = C_m \frac{\partial V_m}{\partial t} + I_{on}
\]

\[
V_m = \phi_i - \phi_e
\]

where \( \sigma \) and \( \sigma_e \) are the intra and extracellular conductance tensors, respectively, \( \beta \) is the surface to volume ratio (1400 cm⁻¹), \( \phi_i \) and \( \phi_e \) are intra and extra cellular potentials, respectively, \( I_{on} \) is the transmembrane current, \( C_m \) is the membrane capacitance (0.01 μF mm⁻²), \( V_m \) is the transmembrane potential and \( I_m \) is the cellular ionic current simulated using the Nygren et al., atrial electrophysiology cellular model. The conduction tensors are aligned with the local tissue microstructure orientation with intracellular fibre and cross-fibre conduction set to 0.28 and 0.026 S/m, respectively. The extracellular fibre and cross-fibre conduction were set to 0.22 and 0.13 S/m, respectively, for linear and focal activation patterns. In the tissue model the fibre direction was aligned parallel to the x-axis. Isotropic conduction with equal intra and extracellular conductivities of 0.15 S/m was employed for re-entrant and spiral wave activation patterns in order to facilitate the production of stable re-entrant waves within the 4 x 4 cm monolayer domain. Following stimulation to reduce computational load, the model reverts to the mono-domain equations:
\[ \nabla \cdot (\mathbf{\sigma} \nabla V_m) + I_{\text{ion}} = \beta \left( C_m \frac{\partial V_m}{\partial t} + I_{\text{ion}} \right), \]

where \( \mathbf{\sigma} \) is the conductivity tensor and the fibre and cross fibre conductivity is set to the harmonic mean intra and extra cellular conductivities giving fibre and cross fibre values of 0.1232 and 0.0217 S/m, respectively (or 0.075 S/m for isotropic conduction). Electrical propagation was determined by solving the bidomain and monodomain equations using the finite element method. The model was discretized in space with linear finite elements; the non-linear term describing the ionic current was treated with a splitting technique. The ionic model was discretized in time with a forward-Euler scheme and a time step of \( \Delta t = 5 \mu s \), while the partial differential equations were solved with a time step of \( \Delta t = 10 \mu s \). The numerical simulations were performed with the Cardiac Arrhythmia Research Package (CARP) solver.

To create uniform linear activation, a 2 mm region at the left edge of the tissue was stimulated. To create focal activation a 1 x 1 mm region close to the centre of the monolayer was stimulated. To create re-entry, a circular non-conducting region (diameter 1.5 cm) was introduced in the monolayer and a dual stimulus protocol was used to initiate re-entry. To create spiral wave activation, two linear stimuli were used. The first stimulus activated the entire extent of the leftmost edge of the monolayer, whilst the second critically timed stimulus initiated spiral wave re-entry. In silico isochronal maps (see leftmost column, Figure 6) were constructed from the transmembrane potential fields by calculating the time elapsed between a fixed temporal reference and the time at which the transmembrane potential exceeded a pre-defined voltage threshold (-20 mV).

### Local activation map construction

Re-interpolated LAT maps were created by down-sampling the number of acquired points and performing linear interpolation of the resulting activation field across the 2D domain for the simulated activation patterns or 3D geometry (i.e. electroanatomic shell) for the clinical arrhythmias. For locations beyond the outermost re-sampled points linear extrapolation based on boundary gradients was used to calculate activation times. The scatteredInterpolant class (MATLAB 8.2, The Mathworks Inc., Natick, MA) was used to implement interpolation and extrapolation methods. When performing re-interpolation on the clinical datasets, a ‘region of electrical interest’ was defined as all the nodes of the electroanatomic map lying within 10 mm of an original electrical mapping point. During re-interpolation, the interpolant was only interrogated for mesh nodes lying within this region. Points out-with this region were assigned indeterminate local activation times and are represented as grey in the accompanying figures.

### Optimal LAT sampling density measurement

For simulated activation patterns, we defined the reference map as the high-resolution isochronal map created from the activation times calculated at every mesh node. For clinical activation patterns, we defined the reference map as the LAT map created by the clinical mapping system using all available LAT points. Reference and re-interpolated maps were compared quantitatively by computing the sum of squared differences between activation times at every mesh node in the region of electrical interest of the chamber model. Results are presented per node (average LAT map error, ms/node) in order to normalize for inter-case differences in the number of nodes per geometry.

The optimal sampling density for each LAT map was defined as the highest sampling density resulting in minimal further error reduction in the re-interpolated maps (defined as an error reduction rate of <0.05 ms per mesh node per additional point sampled). This threshold represents a LAT interpolation error of 1% assuming maximal sampling density of 4 points/cm² and typical conduction velocity of 1 m/s⁻¹. Calculation of the error reduction rate was made as shown in Figure 2. First, each map was re-interpolated with point sampling densities in the range 0.25–10 points/cm² (step size 0.125 points/cm², simulated data repetitions = 10; clinical data repetitions = 10). For each repetition, sample points were distributed randomly across the mapping domain. The mean LAT map accuracy (see above) for each density was plotted against sampling density and a power law of the form \( f(s) = as^b + c \) was fitted to the resulting mean accuracies (where \( s \) is the sampling density and \( a, b, \) and \( c \) are fitted to match the data).

#### Figure 1

Optimal LAT sampling densities. Calculated optimal LAT sampling densities are shown for simulated activation patterns (A) and in vivo LA activation patterns (B). FO, focal origin; MA, macro re-entry; Mi, micro (localised) re-entry.

### Table 1

**Optimal LAT sampling density (points/cm²)**

<table>
<thead>
<tr>
<th>Activation pattern</th>
<th>Linear</th>
<th>FO</th>
<th>Re-entry</th>
<th>Spiral</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Simulated</strong></td>
<td>0.5</td>
<td>1.0</td>
<td>2.0</td>
<td>2.5</td>
</tr>
<tr>
<td><strong>Clinical</strong></td>
<td>0.5</td>
<td>1.0</td>
<td>2.0</td>
<td>2.5</td>
</tr>
</tbody>
</table>

**Note:** P-values for comparison between sampling densities for each activation type were calculated using the Student’s t-test (two-tailed, unequal variances). The optimal sampling density for each LAT map was defined as the highest sampling density resulting in minimal further error reduction in the re-interpolated maps (defined as an error reduction rate of <0.05 ms per mesh node per additional point sampled). This threshold represents a LAT interpolation error of 1% assuming maximal sampling density of 4 points/cm² and typical conduction velocity of 1 m/s⁻¹. Calculation of the error reduction rate was made as shown in Figure 2. First, each map was re-interpolated with point sampling densities in the range 0.25–10 points/cm² (step size 0.125 points/cm², simulated data repetitions = 10; clinical data repetitions = 10). For each repetition, sample points were distributed randomly across the mapping domain. The mean LAT map accuracy (see above) for each density was plotted against sampling density and a power law of the form \( f(s) = as^b + c \) was fitted to the resulting mean accuracies (where \( s \) is the sampling density and \( a, b, \) and \( c \) are fitted to match the data).

**Table 2**

**Optimal LAT sampling density (points/cm²)**

<table>
<thead>
<tr>
<th>Tachycardia type</th>
<th>Linear</th>
<th>FO</th>
<th>Macro</th>
<th>Localised</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Simulated</strong></td>
<td>0.5</td>
<td>1.0</td>
<td>2.0</td>
<td>2.5</td>
</tr>
<tr>
<td><strong>Clinical</strong></td>
<td>0.5</td>
<td>1.0</td>
<td>2.0</td>
<td>2.5</td>
</tr>
</tbody>
</table>

**Note:** P-values for comparison between sampling densities for each activation type were calculated using the Student’s t-test (two-tailed, unequal variances). The optimal sampling density for each LAT map was defined as the highest sampling density resulting in minimal further error reduction in the re-interpolated maps (defined as an error reduction rate of <0.05 ms per mesh node per additional point sampled). This threshold represents a LAT interpolation error of 1% assuming maximal sampling density of 4 points/cm² and typical conduction velocity of 1 m/s⁻¹. Calculation of the error reduction rate was made as shown in Figure 2. First, each map was re-interpolated with point sampling densities in the range 0.25–10 points/cm² (step size 0.125 points/cm², simulated data repetitions = 10; clinical data repetitions = 10). For each repetition, sample points were distributed randomly across the mapping domain. The mean LAT map accuracy (see above) for each density was plotted against sampling density and a power law of the form \( f(s) = as^b + c \) was fitted to the resulting mean accuracies (where \( s \) is the sampling density and \( a, b, \) and \( c \) are fitted to match the data).
(parameters). A power law was chosen owing to the observed distribution of the average LAT map error and to permit an analytical determination of optimal sampling density that was less sensitive to noise. Taking the first derivative of this function (i.e. $f(s) = \frac{abs}{C0^1}$) gives the improvement in map accuracy per node relative to an increase in sampling density. This equation was solved for $f(s) = \frac{C0^0}{D}$, in order to determine the sampling density at which minimal further improvement in map accuracy is achieved by further point sampling.

**Electroanatomic surface area calculation**

Left atrial cardiac chamber models were exported from the Carto3 mapping system and imported into MATLAB 8.2 (The Mathworks Inc., Natick, MA) using custom-written software. The total surface area of the chamber was calculated as the summation of the areas of all triangles within a distance of 10 mm of an original mapping point. The threshold of 10 mm was chosen to delineate electrically inactive regions (e.g. superior/inferior caval veins, pulmonary veins) from the chamber body and is analogous to setting a similar value as the colour fill threshold of the EAM platform.

**Data analysis and statistics**

Custom software was written for determining the optimal LAT sampling point density (MATLAB 8.2, The Mathworks Inc., Natick, MA). Data analysis was performed using Prism version 6.0c (GraphPad Software, San Diego, California, USA). Results are presented as mean ± SD. Student’s unpaired t-test was used to compare group means. Multiple comparisons were performed with the one-way analysis of variance (ANOVA). Proportions in two or more groups were compared with the Chi-square test. A significance level of $P < 0.05$ was considered statistically significant.

**Results**

**Clinical left atrial tachycardias**

Local activation time maps representing 21 left atrial activation patterns were studied including focal activation ($n = 8$), localized re-entry ($n = 5$) and macro-re-entry ($n = 8$) (see Figure 3). Focal activation included four maps created during pacing, from the left atrial appendage ($n = 1$) and coronary sinus ($n = 3$). There were no significant differences in symptomatology, co-morbidities, or use of anti-arrhythmic drugs between the groups (Table 1). The mean LA surface area was $156 ± 44$ cm$^2$ with an original sampled point density of $5.6 ± 4.2$ points/cm$^2$. Optimal LAT sampling density was lower for focal tachycardias than both localised re-entry and macro-re-entry tachycardias (calculated optimal sampling densities $0.67 ± 0.17$ points/cm$^2$ for focal tachycardias, $1.05 ± 0.32$ points/cm$^2$ for macro-re-entrant tachycardias and $1.23 ± 0.26$ points/cm$^2$ for localized re-entry, $P = 0.0031$, Figure 1B). Calculated optimal sampling density was
Insensitive to the selected value of error reduction rate (Table 2). The focal tachycardia maps re-interpolated at optimal LAT sampling density (LAT error rate reduction <0.05 ms/node/point) resulted in correct localization of the foci of earliest activation (Figure 3A). In the re-entrant tachycardias re-interpolation at optimal LAT sampling density resulted in the same overall map appearance, with the same macro-re-entrant circuit remaining identifiable (Figure 3B). However, some variations in map appearance at the extremes of interpolation (e.g. beyond the pulmonary vein ostia) were identifiable. The localized re-entrant maps re-interpolated at optimum sampling density (Figure 3C) were remarkably similar in appearance to the original maps; but some variations were evident in activation timing extrapolated into the pulmonary veins.

Porcine right atrial activation and chamber geometry complexity

Eight pre-ablation LAT maps created under proximal CS pacing were studied. In the absence of ablation, activation proceeded superiorly in a linear fashion around the tubular right atrium in all cases (Figure 4A). After incomplete inter-caval linear ablation, a new activation detour was evident in all maps (Figure 4B, asterisk). Increasing activation pattern complexity, created by ablation, resulted in a significant increase in the optimal LAT sampling density (0.45 ± 0.13 points/cm² pre-ablation vs. 0.78 ± 0.17 points/cm² post-ablation, P = 0.0008, Figure 4C). Overall, the optimal point sampling density for the geometrically-simple porcine right atrium was significantly less than that for the geometrically-complex human left atrium (0.61 ± 0.22 points/cm² vs. 1.0 ± 0.34 points/cm², P = 0.0015).

Simulated activation patterns

Four simulated activation patterns were studied (linear, focal, spiral-wave, and re-entrant activation). Initiation of activation is shown in Figure 5. Reference LAT maps were created from local activation times defined at every mesh node, resulting in an extremely high reference sampling density of ~2500 points/cm². Calculated optimal LAT sampling densities (Figure 1A) for focal, re-entrant and spiral-wave activation patterns were 1.09 ± 0.14 points/cm², 1.44 ± 0.49 points/cm², and 1.50 ± 0.34 points/cm², respectively. Linear activation in a 2-dimensional domain represents a special case where map accuracy is independent of sampling density (Figure 6A).

Examples of re-interpolated isochronal LAT maps at sub-optimal, optimal, and supra-optimal LAT sampling densities for focal, re-entrant, and spiral-wave activation patterns are shown in Figure 6B–D. For focal activation, sub-optimal sampling resulted in incorrect focus localization. For re-entrant activation, sub-optimal sampling density resulted in an activation pattern more consistent with a focal source. Finally, sub-optimal sampling of spiral-wave activation failed to reveal rotational activity in the resulting re-interpolated map. In each case there was limited morphological improvement in the isochronal map appearances at supra-optimal compared to optimal LAT sampling densities (Figure 6B–D, third and final columns).
The main findings of this study are: (1) optimal LAT sampling densities can be identified for a variety of tachycardia mechanisms to maximize the diagnostic yield of interpolated local activation time maps, with a minimal sampling density of 1.0–1.5 points/cm² identified for optimal interpolation of accurate LAT maps; (2) optimal LAT sampling density is dependent on activation pattern with greater sampling densities required to correctly reveal more complicated activation sequences including macro-re-entry and spiral-wave activation; and (3) optimal LAT sampling density is also dependent on chamber geometry, with increasing geometric complexity necessitating an increased optimal LAT sampling density.

The aim of tachycardia activation mapping is 2-fold: to determine the tachycardia mechanism and thereby inform an ablation strategy. The ablation target is determined by the site of tachycardia origin (focal mechanism) or the location of a critical isthmus of conduction (re-entrant mechanism). Local activation time mapping should be supplemented by consideration of the response to entrainment manoeuvres and more importantly the response to ablation when confirming the final tachycardia mechanism. Not infrequently, local activation time maps (which depend on choice of an adequate reference, window of interest and appropriate electrogram annotation) may produce conflicting information. In this context, understanding the mapping density required to achieve optimal sampling of the arrhythmia mechanism is crucial in optimising diagnostic yield of mapping prior to selection of an ablation target. Three-dimensional electro-anatomic mapping systems provide a framework for visualization of electrical activation sequences as an aid to identifying the tachycardia mechanism and corresponding ablation target. Recent technological advances have enabled significant increases in the spatial resolution achievable within reasonable timescales with both established and emerging contact mapping platforms.

Whilst high sampling density is clearly important for substrate-mapping accuracy, for example in identifying areas of low voltage or fractionated signals important to arrhythmogenesis, it is uncertain if high-resolution mapping results in clinical benefit during activation mapping, where the primary aim is to reliably identify the tachycardia mechanism. Indeed high-resolution activation mapping is at odds with earlier strategies aiming to minimize procedure time without impacting success by providing a logical framework for directing the acquisition of LAT points. To this end, we have shown that in systems using interpolation methods to create isochronal LAT maps from a number of sampled LAT times, a theoretical maximal sampling density is reached at which further clinically-relevant improvements in LAT map accuracy are minimal. By using both simulated data in addition to clinical data, we have demonstrated that this maximal sampling density is within the same range (1.0–1.5 points/cm²) irrespective of the resolution at which the ‘gold standard’ activation sequence is defined.

Figure 4 Porcine right atrial LAT maps. (A) Pre-ablation LAT shows uniform activation across the chamber from coronary sinus towards the right atrial appendage. (B) Post-ablation LAT shows a new activation detour (asterisk) via the mid-chamber gap in the ablation line. Ablation points are shown with red tags and catheter positions during ablation with pink dots using the Visitag Carto3 module. (C) Comparison between optimal LAT sampling density for pre- and post-ablation LAT maps.
In performing this analysis, distributions of mapping points at a range of sampling densities were used to identify the global density at which further sampling yielded minimal improvement in activation map accuracy. In doing so, a random distribution of ‘re-sampling’ points was used for the following reasons. First, given the flexible nature of mapping catheters the spatial distribution of clinical mapping points is non-uniform. Secondly, the endocardial surface of the atrium is irregular resulting in non-uniform spatial distribution of mapped points even with uniform catheter electrode spacing. Finally, by repeating the randomization multiple times, and calculating error for each repeated interpolated activation map the overall calculated optimal density should be independent of variations in the spatial distribution of re-sampling points. Nevertheless, non-uniform activation mapping is likely to be important in certain clinical circumstances (e.g. focal tachyarrhythmias—see below).

The identification of atrial spiral-waves has received renewed attention recently as a strategy to identify ablation targets in atrial fibrillation.15 Whilst rotors have been identified in animal16 and computer models17 of atrial activation, there is some debate as to which technologies can reliably detect the occurrence of rotational activity in vivo.18,19 Clinical approaches to identifying rotors typically involve phase mapping in order to average activation over many cardiac cycles. This approach has successfully identified rotors in the left atrium using a 64-pole basket catheter.15 Taking the left atrial

### Table 1  Patient characteristics

<table>
<thead>
<tr>
<th></th>
<th>Focal activation (n = 8)</th>
<th>Localized re-entry (n = 5)</th>
<th>Macro-re-entry (n = 8)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Symptoms</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Palpitations</td>
<td>8 (100%)</td>
<td>3 (60.0%)</td>
<td>7 (87.5%)</td>
<td>0.1317</td>
</tr>
<tr>
<td>Breathlessness</td>
<td>0 (0.0%)</td>
<td>1 (20.0%)</td>
<td>2 (25.0%)</td>
<td>0.3301</td>
</tr>
<tr>
<td>Tachycardia-associated cardiomyopathy</td>
<td>0 (0.0%)</td>
<td>1 (20.0%)</td>
<td>1 (12.5%)</td>
<td>0.4581</td>
</tr>
<tr>
<td><strong>Co-morbidities</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Atrial fibrillation</td>
<td>7 (87.5%)</td>
<td>5 (100%)</td>
<td>7 (87.5%)</td>
<td>0.7079</td>
</tr>
<tr>
<td>Hypertension</td>
<td>2 (25.0%)</td>
<td>2 (40.0%)</td>
<td>2 (25.0%)</td>
<td>0.8106</td>
</tr>
<tr>
<td>Diabetes mellitus</td>
<td>0 (0.0%)</td>
<td>1 (20.0%)</td>
<td>1 (12.5%)</td>
<td>0.4581</td>
</tr>
<tr>
<td>Stroke/TIA</td>
<td>1 (12.5%)</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>0.4261</td>
</tr>
<tr>
<td>Vascular disease</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>n/a</td>
</tr>
<tr>
<td>Heart failure</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>n/a</td>
</tr>
<tr>
<td>Coronary artery disease</td>
<td>1 (12.5%)</td>
<td>0 (0.0%)</td>
<td>1 (12.5%)</td>
<td>0.7079</td>
</tr>
<tr>
<td>Valve disease</td>
<td>1 (12.5%)</td>
<td>1 (20.0%)</td>
<td>0 (0.0%)</td>
<td>0.4581</td>
</tr>
<tr>
<td>COPD</td>
<td>1 (12.5%)</td>
<td>0 (0.0%)</td>
<td>0 (0.0%)</td>
<td>0.4261</td>
</tr>
<tr>
<td>Hypo/hyperthyroidism</td>
<td>1 (12.5%)</td>
<td>1 (20.0%)</td>
<td>0 (0.0%)</td>
<td>0.4581</td>
</tr>
<tr>
<td><strong>Echocardiography</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>LVEF (%)</td>
<td>62.5 ± 2.7%</td>
<td>58 ± 11.0%</td>
<td>53.3 ± 10.8%</td>
<td>0.2361</td>
</tr>
<tr>
<td>LA diameter (cm)</td>
<td>4.3 ± 1.1 cm</td>
<td>5.0 ± 1.1 cm</td>
<td>4.4 ± 0.6 cm</td>
<td>0.1179</td>
</tr>
<tr>
<td>LA area (cm²)</td>
<td>25.6 ± 5.5 cm²</td>
<td>31.6 ± 3.8 cm²</td>
<td>24.8 ± 5.7 cm²</td>
<td>0.1321</td>
</tr>
<tr>
<td>Prior Ablation</td>
<td>4 (50.0%)</td>
<td>5 (100%)</td>
<td>8 (100%)</td>
<td>0.0180 *</td>
</tr>
<tr>
<td><strong>Antiarrhythmic drugs</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Current AAD use</td>
<td>3 (37.5%)</td>
<td>0 (0.0%)</td>
<td>3 (37.5%)</td>
<td>0.2691</td>
</tr>
<tr>
<td>AAD</td>
<td>Flecainide ×3</td>
<td>Flecainide ×1, Sotalol ×2</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>TIA, transient ischaemic attack; COPD, chronic obstructive pulmonary disease; LVEF, left ventricular ejection fraction; AAD, antiarrhythmic drug.</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

### Table 2  Sensitivity of optimal sampling density to chosen error reduction rate threshold

<table>
<thead>
<tr>
<th></th>
<th>ERR (ms/node/Δdensity)</th>
<th>OSD (points/cm²)</th>
<th>Sensitivity (ΔOSD/ΔERR)</th>
<th>1/Sensitivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Focal tachycardia</td>
<td>0.05</td>
<td>0.67±0.17</td>
<td>0.057±0.024</td>
<td>18</td>
</tr>
<tr>
<td>Macro-re-entry</td>
<td>0.05</td>
<td>1.05±0.32</td>
<td>0.052±0.022</td>
<td>19</td>
</tr>
<tr>
<td>Localized re-entry</td>
<td>0.05</td>
<td>1.23±0.26</td>
<td>0.057±0.018</td>
<td>18</td>
</tr>
</tbody>
</table>

Throughout this study an Error Reduction Rate (ERR) of 0.05 ms/node/Δdensity is taken to represent the density at which further sampling offers limited improvement in interpolated LAT map accuracy (termed optimal sampling density, OSD). The sensitivity of OSD to changes in choice of ERR are presented as the derivative of OSD with respect to ERR. The final column (1/Sensitivity) indicates the change in ERR that would result in a change in OSD of 1 point/cm².
endocardial surface area to be 150 cm$^2$ (as in this study) and assuming maximal electrode contact, this equates to a sampling density of 0.43 points/cm$^2$. A recent analysis using the basket catheter found a mean of only 31 electrodes provided interpretable electrograms. Suboptimal contact therefore results in lower sampling densities (e.g. 56 electrodes equates to 0.37 points/cm$^2$, 48 electrodes to 0.32 points/cm$^2$, and 32 electrodes to 0.21 points/cm$^2$). All of these densities are below the optimal LAT sampling density for spiral-wave activity identified in this study. Of note an alternative approach using dynamic voltage mapping applied to identical clinical data failed to identify sustained rotational activity in 100% ($n = 11$) of cases. This result is consistent with our finding that a significantly higher sampling density of around 1.5 points/cm$^2$ should be required to optimally reproduce spiral-wave activity from known local activation time points.

The results presented here should not be interpreted to indicate that sampling densities above 1.5 points/cm$^2$ are never of clinical benefit in activation mapping. One limitation of the ‘optimally’ resampled clinical maps shown in Figure 3 is that interpolation at regions distant to the body of points (e.g. extending into the pulmonary veins or LAA) may be inaccurate. Furthermore a local density of >1.5 points/cm$^2$ may well be required to correctly identify the origin of focal or localised-re-entrant tachycardias. Such variable distributions

Figure 5 Initiation of activation patterns in the atrial monolayer. (A) Linear activation; (B) focal activation; (C) re-entrant activation; and (D) spiral wave activation. Stimulated regions are shown in white in the left-most column of each row.
of mapping points have not been examined in this study. Rather, the results of this study support the hypothesis that once an upper global sampling density of 1.5 points/cm² has been reached (equivalent to around 225 points for a typical LA with endocardial surface area of 150 cm²) then tachycardia mechanisms should be evident or strongly suggested, and, if so, subjected to further testing which may include more detailed activation mapping (for candidate focal or localized re-entry mechanisms) or entrainment mapping (for candidate macro-re-entry mechanisms). Furthermore, where the site of earliest activation through an incomplete linear lesion is sought, higher density mapping at the planned ablation target may be of benefit.⁴

**Limitations**

Whilst this study has assessed mapping density for atrial arrhythmia contact mapping using typical electrode sizes/configurations used in clinical practice, the impact of alternative electrode size on optimal sampling density has not been examined and should be addressed in future studies. The clinical population studied in this manuscript is small and represents a select group of patients. In particular, we highlight (Table 1) that the cases were characterised by mild underlying structural heart disease and were predominantly cases having undergone prior ablation.

**Conclusions**

In conclusion, this work demonstrates that for commonly-encountered clinical tachycardia mechanisms the majority of information discernible from LAT maps can be obtained with a limited sampling density strategy. The exact sampling density required depends on tachycardia mechanism and clinical characteristics but is in the range of 1.0–1.5 points/cm². In the event where candidate mechanisms are not identifiable at this sampling density, rather than collecting further points a better approach may be to begin a new activation map perhaps with a different window of interest, reference electrogram, or LAT assignment technique.

**Conflict of interest**

None declared.

**Funding**

This work was supported by the British Heart Foundation (PG/13/37/30280), an unrestricted educational grant from Biosense Webster and the National Institute for Health Research (NIHR) Biomedical Research Centre at Guy’s and St Thomas’ NHS Foundation Trust and King’s College London. The views expressed here are those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health.

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![Figure 6](https://academic.oup.com/europace/article-abstract/20/2/e11/3098571/3098571)

Figure 6 Simulated isochronal local activation time maps. (A) Linear activation; (B) focal activation; (C) re-entrant activation; and (D) spiral wave activation. Reference LAT maps are shown in the left most column. In A and B, local activation time is taken as the first activation of each node. In C and D, a window of interest (WOI) incorporating 100% of the tachycardia cycle length (TCL) was specified. Grey colour represents tissue that is never activated during the WOI—either non-conducting (C) or functionally refractory (D).
References


