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Ventricular fibrillation (VF), the principle cause of sudden cardiac death,1,2 is largely resistant to ion channel–targeting antiarrhythmic drugs.3–6 The pathophysiology of ischemia (that mediates the adverse electrophysiological milieu) 7 may represent a more promising target.

Animal studies have shown that early (phase I) VF arises during the first 30 minutes of ischemia when arrhythmogenic substances accumulate, 8 but it remains unclear which substances are sufficient and necessary mediators of VF.8 Mediators may derive from ischemic myocytes and from other local tissue (including platelets). Thus, platelets may facilitate VF independently of their ability to participate in occlusive thrombosis. Clinical trials suggest the mortality benefit of antiplatelet agents, such as aspirin and αIIbβ3 integrin antagonists, may result from effects more complex than simple thrombus inhibition,9–11 In animal studies, several platelet-activating factor antagonists12–14 reduced ischemia-induced VF incidence. Moreover, aspirin pretreatment reduced ischemia-induced VF in various animal models.15–17 However, there is a lack of evidence directly linking anti-VF effects to platelet ablation. It has been argued that a systematic examination of the role of platelets in VF initiation during ischemia is long overdue.18

We have tested whether VF is facilitated by the activation of platelets trapped within the ischemic territory and activated to release arrhythmogenic secretome.

**Methods**

**Animals and Core Methods**

Experiments were performed according to the United Kingdom Home Office Guide on the Operation of the Animals (Scientific Procedures) Act 1986. Heart perfusion, induction of ischemia, verification and quantification of IZ size, and data recording and analysis were as described previously.19 Male Wistar rats (220–270 g) were anesthetized (pentobarbitone 60 mg/kg IP) and given sodium heparin 250 IU IP. Blood samples (7–10 mL) for platelet preparation were
drawn by inferior vena caval puncture and collected into acid citrate dextrose buffer (120 mmol/L sodium citrate, 110 mmol/L glucose, 80 mmol/L citric acid). Rats were then killed by cardiac excision, and hearts excised, immersed in cold (4°C) Krebs solution (modified to contain 3 mEq/L K+ and 1.4 mEq/L Ca++), then cannulated for perfusion with similar Krebs solution (gased with 95% O2 plus 5% CO2) at 37°C at a constant pressure (70 mm Hg). A silver electrogram (ECG) lead was positioned to impale a part of the anterior surface of the left ventricle that, from our experience, would be subserved by the artery to be occluded. The ECG was recorded (PowerLab; ADInstruments, United Kingdom) at a sampling rate of 2 kHz and with high- and low-pass frequency filters set at 0.3 Hz and 1 kHz. The standard definitions of rat ECG intervals were used.21 Coronary flow (CF) was measured by timed collection of coronary effluent. A silk suture (3/0 thread), sewn under the left main artery immediately after setup, was tightened to achieve regional ischemia, and the size of the IZ was determined at the end of perfusion using the disulphine blue method.19

Platelet Preparation and Control Groups
Washed platelets were derived by centrifugation, resuspended (2×10^8/mL) in Krebs solution, then rested for ≥10 minutes before use. To validate platelet viability and aggregability when suspended in Krebs solution (the preparation method is validated only with Tyrode solution), platelet aggregometry was performed using fibrillar type I horn collagen (Nycomed, Axis-Shield, United Kingdom), thrombin (Sigma, Poole, United Kingdom), and ADP (Sigma). Three control groups were used in the first of 2 studies: platelet vehicle control solution (Krebs), fluorospheres, and secretome. The fluorosphere control group tested whether any increase in arrhythmias in the platelet group was mediated by the mere presence of platelet-sized particles. The fluorospheres (Invitrogen, Paisley, United Kingdom) mimic platelet size and charge. Platelet secretome controls tested whether secretome facilitates ischemia-induced VF and whether secretome could evoke VF in the absence of ischemia. Secretome contains substances (eg, ATP, serotonin, histamine, and >300 proteins),23 many of which can alter cardiac electrophysiology.24 To obtain secretome, the washed platelet pellet was resuspended in 4 mL Krebs solution and stimulated with 1 U/mL thrombin. The aggregated suspension was then centrifuged (1000 g for 3 minutes) and the secretome collected.

Platelets, fluorospheres, and secretome were prepared in 4 mL of warmed, gased Krebs solution immediately before infusion and contained 2x10^8/mL platelets, 4x10^9/mL fluorospheres, or (as quantified by Bradford assay) 0.21 mg/mL protein, respectively.

Platelet Delivery System
A Langendorff cannula was modified to allow infusion of washed platelets into the perfusate in a nonactivated and functional state (Figure 1). Platelets, platelet vehicle, fluorospheres, or secretome were delivered as infusion solutions to the cannula from polypropylene syringes attached to a syringe driver.

Experimental Design
In the first study, platelets were trapped in the IZ by coronary ligation. To validate the method, hearts were perfused with platelets radiolabelled with 50 MBq 111In oxine sulfate. After coronary ligation, hearts were frozen in liquid nitrogen for autoradiography to be performed on 20-µm sections exposed to CL-XPosure Film (Thermo Fisher Scientific, Loughborough, United Kingdom) for 3 days. Then, 96 rats were randomized into 8 groups: Krebs solution controls, washed platelets, platelet secretome or fluorospheres, each with coronary ligation or sham ligation. All hearts were perfused for 30 minutes with

![Figure 1](image_url)

**Figure 1.** Novel cannula engineered for direct platelet delivery to the isolated heart. A modified Langendorff cannula was made from stainless steel and modified to receive a sidearm to allow passage of an inert 1-mm internal diameter polypropylene tube to the tip of the cannula. The shear rate within the sidearm tubing was subarteriolar (<1000/s).

![Figure 2](image_url)

**Figure 2.** Platelet aggregation studies confirm platelet viability and functionality. Washed rat platelets were derived by standard centrifugation and resuspended at a concentration of 2x10^8/mL. Aggregation traces demonstrated normal dose-response curves to agonist stimulation with collagen, ADP, and thrombin in Tyrode solution (A–C) or Krebs solution (D and E). X refers to the time point at which 500 µL of washed platelets were perfused through the modification to the cannula at 100 µL/s. All traces are representative of n=3.
ischemia and is not reperfusion-induced release, because true reperfusion-induced platelet adhesion and activation takes several minutes to manifest.\textsuperscript{26}

Table 1. Baseline Characteristics of Hearts Subjected to Coronary Ligation

<table>
<thead>
<tr>
<th>Baseline Variables</th>
<th>Krebs Buffer (N=12)</th>
<th>Platelet (N=11)</th>
<th>Secretome (N=12)</th>
<th>Fluorospheres (N=10)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart rate, beats/min</td>
<td>264.4±8.9</td>
<td>278.5±8.9</td>
<td>280.1±12.4</td>
<td>287.0±13.3</td>
<td>0.89</td>
</tr>
<tr>
<td>Coronary flow, mL/min</td>
<td>10.1±0.7</td>
<td>9.7±0.7</td>
<td>10.0±0.7</td>
<td>10.7±0.6</td>
<td>0.73</td>
</tr>
<tr>
<td>PR interval, ms</td>
<td>38.2±1.2</td>
<td>39.7±2.4</td>
<td>37.6±0.8</td>
<td>39.2±1.8</td>
<td>0.82</td>
</tr>
<tr>
<td>QT90 interval, ms</td>
<td>53.3±1.8</td>
<td>48.5±2.3</td>
<td>55.2±2.2</td>
<td>52.3±1.9</td>
<td>0.59</td>
</tr>
<tr>
<td>IZ, % of ventricular mass</td>
<td>34.1±0.8</td>
<td>33.0±0.7</td>
<td>34.2±1.0</td>
<td>34.6±0.8</td>
<td>0.64</td>
</tr>
</tbody>
</table>

IZ indicates ischemic zone.

Figure 3. The delivery of platelets is sufficient to cause platelet trapping. To validate the platelet delivery/trapping method, rat hearts were perfused with platelets radio-labelled with 50 MBq \textsuperscript{111}In oxine sulfate. After coronary ligation, hearts were frozen in liquid nitrogen for autoradiography to be performed on 20-µm sections. Platelets were trapped by coronary ligation in the ischemic zone (left ventricle, LV) and excluded (little adhesion) in the remote region (right ventricle, RV). Representative image of n=3. Krebs solution, then infusion solution was delivered at 0.2 mL/min for 20 minutes, with coronary ligation (or sham ligation) performed at the half way point (10 minutes).

Thromboxane Assay

In separate groups of hearts, at the end of platelet infusion (after 10-minute ischemia), the ligature was released and coronary effluent collected for 60 seconds. Thromboxane (TX) B\textsubscript{2} was measured by ELISA assay (Biotrak thromboxane B\textsubscript{2} enzyme immunoassay system; GE Healthcare, Amersham, United Kingdom). The TX measured is a marker of platelet activation and degranulation during ischemia and is not reperfusion-induced release, because true reperfusion-induced platelet adhesion and activation takes several minutes to manifest.\textsuperscript{26}

Drug Administration and Blood Trapping

In a second study, an equivalent protocol was used to explore the ability of whole blood to mimic the ability of trapped platelets to facilitate ischemia-induced VF and to test the effects of standard anti-platelet drugs. A blood sample was taken before cardiac excision, heparinized (0.1 U/mL), then delivered via the system used earlier (4 mL, >20 minutes, at a rate of 0.2 mL/min), with coronary ligation and blood trapping performed half way through the infusion. Rats received PO indomethacin (n=14; 2.4 mg/kg per day; Sigma), clopidogrel (n=9; 10 mg/kg per day; Wockhardt, Wrexham, United Kingdom), or vehicle (n=20; 1 mL/kg per day of 4% v/v ethanol in distilled water) for a minimum of 5 consecutive days. The vehicle group was subdivided to provide blood-trapped (n=8) and blood-free (Krebs solution throughout; n=12) controls.

Exclusion Criteria

Based on previously published standards, hearts were excluded if preligation sinus heart rate (HR) was <200 beats/minute or CF was <5 mL/min,\textsuperscript{27} or if IZ was outside the range of 25% to 40% of total ventricular weight, or VF occurred before the onset of ischemia. VF susceptibility in individual hearts is not dependent on baseline HR or CF beyond these stated exclusion limits.\textsuperscript{28} Blood-perfused hearts were excluded if they developed an intracoronary thrombus before coronary ligation. The studies required 204 rats, of which 12 were excluded.

Statistical Analyses

Parametric variables (±SEM), including CF, HR, PR interval, QT interval at 90% repolarization (QT90),\textsuperscript{29} and light transmission for platelet aggregation, were evaluated by 1-way ANOVA and Dunnett test to compare treatments with control. Incidences of ventricular tachycardia and VF (dichotomous variables) were analyzed by Fisher exact test because of small sample sizes. This test was used for global multiple comparisons, and if significant, a pairwise comparison was performed versus controls using the same test. Other nonparametric variables were analyzed using the Steel–Dwass test, and the Kruskal–Wallis test was used for the estimation of stochastic probability in intergroup comparisons. Probability values were 2-sided and results were deemed statistically significant when $P<0.05$. Statistical analyses were performed on R, version 2.14.

Results

Delivered Platelets Were Viable and Functional

Aggregation traces demonstrated normal dose–response curves to agonists (Figure 2A–C), and 1 U/mL of thrombin induced a maximal aggregation response (Figure 2D). Washed platelets were nonactivated on exiting the delivery system setup without a heart attached, but could be fully activated by agonist stimulation (Figure 2E), validating the platelet delivery system and Krebs solution as a platelet vehicle.

Platelet Trapping Facilitates Ischemia-Induced VF

Coronary ligation trapped infused platelets within the IZ (Figure 3). Mean IZ values (33%–35%) were suboptimal for VF (Table 1), meaning that ischemia-induced VF occurred in only 42% of platelet-free controls as desired.\textsuperscript{28} Platelet trapping increased VF incidence to 91% ($P<0.05$), whereas trapping of secretome or fluorospheres had no such effect (Figure 4). Delivery of platelets in the absence of ischemia did not elicit VF (either in shams, or preischemia in hearts with...
coronary ligation). However, 2 sham coronary-ligated hearts experienced VF during secretome infusion.

**Hemodynamic and ECG Variables in Platelet Trapping Study**

Baseline HR, CF, PR, and QT90 intervals did not differ between the 4 groups (Table 2) here (and elsewhere in the study). Coronary ligation evoked changes typical for the model, such as a fall in CF. There were no significant differences in CF or PR between groups at any time. The QT90 interval in coronary-ligated controls increased during ischemia from 53±2 to 68±3 ms, then gradually fell toward the end of the experiment to 61±3 ms, as seen previously with this model. QT90 changes were similar with platelet trapping, but the increase was significantly greater versus the other groups 30 minutes after the onset of ischemia (platelet, 71±4 ms; control, 61±3 ms; secretome, 64±4 ms; fluorospheres, 59±4 ms; P<0.05). In sham ligation controls, there were no significant effects of platelet or secretome infusion on CF, PR, or QT90. The only HR variation between groups was a transient increase at the start of secretome infusion in sham (Figure 5A) and coronary-ligated hearts (Figure 5B) with a weaker (nonsignificant) trend toward an increase in hearts with platelets trapped by ligation (Figure 5B).

**Regional Ischemia Activates Platelets**

In Krebs solution-perfused control hearts subjected to coronary ligation, no TX was detected in coronary effluent. However, in hearts with platelets trapped in the IZ, a substantial amount of TX was detected (Figure 6).

**Drug Pretreatment Action on Platelets, VF, and Hemodynamics**

Oral pretreatment with antiplatelet agents inhibited (but did not abolish) ex vivo aggregation responses to ADP (Figure 7A) and thrombin (Figure 7B). CF, HR, PR interval, QT90 interval, and IZ size were similar to values in the platelet trapping study and did not vary significantly between groups (Table 3). Ischemia-induced VF was facilitated by blood trapping compared with blood-free controls (P<0.05; Figure 8). Facilitation of VF was not reduced significantly by pretreatment in vivo with clopidogrel (P>0.05) or indomethacin (P>0.05; Figure 8).

**Discussion**

**Overview**

Platelets trapped in the IZ by coronary ligation became activated by ischemia, resulting in an increase in VF incidence in hearts in which VF risk would ordinarily be low due to suboptimal ischemia (small IZ). Platelets were arrhythmogenic only when trapped in the IZ. Trapping of autologous whole blood mimicked the platelet effect. This was not prevented by in vivo pretreatment with clopidogrel or indomethacin (which reduced platelet aggregability). Platelets, platelet trapping, and secretome did not lower CF, precluding the possibility that platelets induced VF by evoking ischemia or rendering it more severe. This is, therefore, the first direct evidence that platelets facilitate VF during ischemia independently of their role in thrombosis.

**Platelets and the Pathophysiological Reserve for VF**

Previous reports on platelet involvement in VF are contradictory. Pretreatment with aspirin, ticlopidine, meclofenamate, or indomethacin reduced ischemia-induced VF incidence from a high baseline in anesthetized rats. However, aspirin failed to reduce ischemia-induced VF in conscious rats with a large IZ. Likewise, platelet-activating factor antagonists ameliorated ischemia-induced VF in vivo in 2 studies, but not in others. Moreover, platelet-activating factor and platelet-activating factor antagonists possess platelet-independent effects on ischemia-induced VF. In most published studies, the IZ was not determined. It is well established that when the IZ is large in vivo and even in Krebs solution-perfused rat hearts, all controls develop VF due to pathophysiological reserve. A pathophysiological reserve precludes identification of individual mediators of VF. In the present study, the pathophysiological reserve was reduced by making IZ suboptimal, thereby revealing the pathophysiological ability of trapped platelets to facilitate ischemia-induced VF. The question remaining is to

**Table 2. Baseline Characteristics of Sham Hearts in the Platelet-Trapping Study**

<table>
<thead>
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<th>Platelet (N=12)</th>
<th>Secretome (N=11)</th>
<th>Fluorospheres (N=11)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart rate, beats/min</td>
<td>285.8±14.5</td>
<td>296.3±12.6</td>
<td>295.5±12.5</td>
<td>277.0±14.7</td>
<td>0.81</td>
</tr>
<tr>
<td>Coronary flow, mL/min</td>
<td>9.5±0.8</td>
<td>9.0±0.9</td>
<td>9.0±0.8</td>
<td>9.6±0.5</td>
<td>0.69</td>
</tr>
<tr>
<td>PR interval, ms</td>
<td>40.4±1.4</td>
<td>35.6±1.3</td>
<td>39.6±1.4</td>
<td>39.8±1.3</td>
<td>0.78</td>
</tr>
<tr>
<td>QT90 interval, ms</td>
<td>56.5±3.2</td>
<td>56.5±1.7</td>
<td>55.1±1.9</td>
<td>52.1±2.9</td>
<td>0.63</td>
</tr>
</tbody>
</table>

![Figure 4. Platelet trapping facilitates ischemia-induced ventricular fibrillation (VF). Incidence of VF evoked by 30-min regional ischemia in the Langendorff-perfused rat heart. Platelets, secretome, or fluorospheres were delivered for 20 min beginning 10 min before the start of ischemia. *P<0.05 vs controls.](image-url)
what extent does this knowledge inform better drug targeting of VF?

Mechanistic Insights

The increase in QT90 interval in hearts with platelets trapped in the IZ resonates with data from De Jong and Dekker who exposed human platelet secretome to rabbit cardiomyocytes and observed action potential duration prolongation and afterdepolarizations. However, other findings provide better clues to mechanisms.

Platelets were inert in nonischemic hearts, so the mechanism of VF facilitation required an interaction with components of the ischemic milieu. Additionally, platelet-sized fluorospheres were not arrhythmogenic (in shams or when trapped by ligation), confirming that VF facilitation by platelets does not operate by simple physical presence.

Trapped platelets released TX in the present study. Moreover, in 2 sham hearts (no ischemia), exogenously administered secretome caused VF. In our historical database of >200 controls, otherwise normal hearts do not exhibit spontaneous VF. The mechanism was primary, because secretome did not induce ischemia (no reduction in CF) in shams. The secretome effect in shams contrasts with the lack of platelet and blood effects. Conversely, secretome had little effect when trapped in the IZ, in sharp contrast to the facilitation of VF by trapped platelets and blood.

These observations provide coherent mechanistic insight: ischemia activates trapped platelets to release secretome containing a labile and short-lived arrhythmogenic mediator, sufficient to facilitate VF when IZ is suboptimal. It is important to emphasize that these mechanistic insights are tentative.

Clinical Relevance

Due to the complex interaction between platelets and other blood components, we modified our platelet-trapping
approach to examine whether standard antiplatelet drugs administered in vivo could reduce the ability of trapped whole blood to facilitate VF in vitro. Clopidogrel and indomethacin were not effective in this regard. Clopidogrel reduces platelet P-selectin expression, but does not prevent platelets from secreting α-granular contents; indomethacin inhibits cyclooxygenase, but is not selective for TX synthesis.\(^3\)\(^5\)\(^,\)\(^3\)\(^6\) Moreover, despite high dosage, the effect of each drug on responses to platelet activators in vitro, although significant, was not substantial. Thus, standard antiplatelet drugs are unable to affect ischemia-trapped platelet function sufficiently to block facilitation of VF. This agrees with their lack of benefit against sudden cardiac death in humans.

**Limitations**

The isolated perfused unloaded rat heart exhibits ischemic VF, which is not subject to normal mechanical or autonomic influences. Confidence that present findings have clinical relevance is limited by the extent to which the lack of loading and intact innervation may change the ability of platelets to facilitate ischemia-induced VF in suboptimal ischemia. To test this, in vivo experimentation will be required.

**Future Work**

There are a large number of constituents of the platelet secretome, and identification of the one(s) necessary for facilitation of VF is beyond the scope of the present study. TXA\(_2\) is a noteworthy labile and short-lived constituent. In addition to our evidence of ischemia-specific TX release from platelets trapped in the IZ, studies with the TX synthase blockers, UK38485, R-68070, and U-63557A,\(^2\)\(^5\)\(^,\)\(^3\)\(^7\) suggest that TXA\(_2\) may facilitate ischemia-induced VF in vivo. Together, this justifies future work to clarify links between the platelet, TX, and ischemia-induced VF.

**Acknowledgments**

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**Sources of Funding**

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**Disclosures**

None.

**References**


Acute myocardial ischemia (MI) may result in ventricular fibrillation (VF) and sudden death, often with a brief interlude between MI onset and death (hence the out-of-hospital prevalence), confirmed as ≥5 to 20 minutes by numerous animal model studies. This brief interlude necessitates pretreatment of those at risk. Unfortunately, available drugs are limited in utility due to safety or efficacy issues. New approaches/targets are needed. Animal studies show that the MI region produces or accumulates numerous pathological mediators that may facilitate ischemia-induced VF. Using blood-free perfused rat hearts, we made ischemic territory sufficiently small (34±1% of left ventricle) to evoke VF in only 42% of controls. Importantly, VF incidence was increased to 91% (P<0.05) by coronary ligation–induced trapping of freshly prepared autologous platelets (trapping confirmed by 111In-labeled platelet autoradiographic imaging) without any change in ischemic territory. The trapped platelets released thromboxane, a component of platelet secretome. Freshly prepared secretome evoked VF in dogs in the absence of hemodynamic changes. J Exp Pharmacol Ther. 1984;24:898–904.

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