An FGFR1-SPRY2 Signaling Axis Limits Basal Cell Proliferation in the Steady-State Airway Epithelium

Graphical Abstract

Highlights

- FGFR1 signaling is important for inhibiting airway stem cell proliferation
- FGFR1 post-translationally activates SPRY2, which inhibits activity of other RTKs
- Balancing the activity of different RTKs tunes airway stem cell proliferation rates
- Ciliated cell fate specification also requires FGFR1 but does not require SPRY2

Authors

Gayan I. Balasooriya,
Jo-Anne Johnson, M. Albert Basson,
Emma L. Rawlins

Correspondence
e.rawlins@gurdon.cam.ac.uk

In Brief

Airway basal cells are highly quiescent at homeostasis. Rawlins et al. show that this quiescence is actively maintained, in part by an FGFR1-SPRY2 signaling mechanism. This FGFR1 signaling limits steady-state basal cell proliferation by repressing an RTK-mediated pro-proliferative signal.
An FGFR1-SPRY2 Signaling Axis Limits Basal Cell Proliferation in the Steady-State Airway Epithelium

Gayan I. Balasooriya,1 Jo-Anne Johnson,1 M. Albert Basson,2 and Emma L. Rawlins1,*

1Wellcome Trust/CRUK Gurdon Institute, Wellcome Trust/MRC Stem Cell Institute, Department of Pathology, University of Cambridge, Cambridge CB2 1QN, UK
2Department of Craniofacial Development and Stem Cell Biology, King’s College London, London SE1 9RT, UK
*Correspondence: e.rawlins@gurdon.cam.ac.uk
http://dx.doi.org/10.1016/j.devcel.2016.03.001

SUMMARY

The steady-state airway epithelium has a low rate of stem cell turnover but can nevertheless mount a rapid proliferative response following injury. This suggests a mechanism to restrain proliferation at steady state. One such mechanism has been identified in skeletal muscle in which pro-proliferative FGFR1 signaling is antagonized by SPRY1 to maintain satellite cell quiescence. Surprisingly, we found that deletion of Fgr1 or Spry2 in basal cells of the adult mouse trachea caused an increase in steady-state proliferation. We show that in airway basal cells, SPRY2 is post-translationally modified in response to FGFR1 signaling. This allows SPRY2 to inhibit intracellular signaling downstream of other receptor tyrosine kinases and restrain basal cell proliferation. An FGFR1-SPRY2 signaling axis has previously been characterized in cell lines in vitro. We now demonstrate an in vivo biological function of this interaction and thus identify an active signaling mechanism that maintains quiescence in the airway epithelium.

INTRODUCTION

The airway epithelium of both mouse and humans is essentially quiescent at steady state, with an extremely low rate of stem cell proliferation (Cole et al., 2010; Kauffman, 1980; Teixeira et al., 2013). Nevertheless, airway basal cells (BCs) can rapidly enter the cell cycle in response to luminal cell loss (Hong et al., 2004; Pardo-Saganta et al., 2015; Rawlins et al., 2007). Several paracrine signaling pathways that promote airway stem cell proliferation following injury have been characterized (reviewed in Hogan et al., 2014). In addition, autocrine signaling mechanisms can initiate airway proliferation in response to local damage (Vermeer et al., 2003). A critical question remains: are there also mechanisms which actively inhibit airway proliferation at homeostasis and therefore function to maintain quiescence?

In overall organization the mouse trachea is very similar to human smaller airways (Hackett et al., 2011; Rock et al., 2010; Teixeira et al., 2013). The adult mouse tracheal epithelium comprises three main cell types. BCs include both slowly dividing stem cells and committed luminal precursors (Mori et al., 2015; Rock et al., 2009; Watson et al., 2015). Luminal secretory cells can self-renew and produce luminal ciliated cells, while ciliated cells are terminally differentiated (Rawlins and Hogan, 2008; Rawlins et al., 2007, 2009). In vitro and in vivo evidence suggests that airway BC proliferation requires epidermal growth factor receptor (EGFR) activity (Brechbuhl et al., 2014; You et al., 2002). Moreover, inhibition of EGFR signaling via contact inhibition is necessary to restrain BC proliferation following injury (Lu et al., 2013). WNT and Notch signaling can also promote BC proliferation in some contexts (Giangreco et al., 2012; Paul et al., 2014; Rock et al., 2011). By contrast, YAP prevents differentiation of BCs (Mahoney et al., 2014; Zhao et al., 2014). However, no specific signaling pathways that actively inhibit BC proliferation at steady state have been identified.

In other organs, stem cell quiescence is actively maintained by feedback inhibition. For example, in the satellite cells of skeletal muscle steady-state quiescence requires the function of specific receptor tyrosine kinase (RTK) inhibitors, SPRY proteins, to antagonize pro-proliferative fibroblast growth factor receptor 1 (FGFR1) signaling (Chakkalakal et al., 2012; Shea et al., 2010). We speculated that similar mechanisms would operate in the steady-state airway epithelium.

FGFR signaling has been extensively studied in lung development and the smaller conducting airways (e.g., Abier et al., 2009; Volckaert et al., 2011, 2013; Yin et al., 2011) where, similar to its role in muscle, it has been found to have a pro-proliferative function. However, the role of FGFR signaling in airway BCs remains undetermined. We therefore tested whether antagonism of FGFR1 activity by SPRY proteins is required for BC quiescence. Surprisingly, we found that deletion of either Fgr1 or Spry2 resulted in increased levels of BC proliferation. We demonstrate that in airway BCs, SPRY2 is post-translationally modified downstream of FGFR1, allowing SPRY2 to antagonize signaling from other RTKs, most likely EGFR, and maintain quiescence. There is a well-documented in vitro relationship between FGFR1-mediated modification of SPRY2 and RAS-ERK inhibition (Lao et al., 2006, 2007). However, a role for this interaction has never previously been identified in vivo.
RESULTS

FGFR1 Signaling Is Required for Normal Tracheal Cellular Homeostasis

FGFR signaling pathway components are readily detected in the steady-state adult mouse trachea by RT-PCR (Figure S1A). Fgfr1 and Spry2 mRNA were also detected in purified BC, secretory, and ciliated cell populations by qRT-PCR (Figures 1A, S1B, and S1C) and by single-cell qRT-PCR (Watson et al., 2015). Moreover, FGFR1 protein and mRNA were detected in BCs and luminal cells in the intact mouse trachea (Figures S1D and S1F). We conditionally deleted Fgfr1 and activated a GFP reporter in tracheal BCs using Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1+/fx (Fgfr1 conditional knockout, cKO) and control Tg(KRT5-CreER); Rosa26R-fGFP mice. Four doses of tamoxifen (tmx) were administered to adult (>8 weeks old) mice (Figure 1B). To confirm deletion of Fgfr1fx in the GFP+ BCs, we performed qRT-PCR on pools of isolated GFP+ BCs and showed that the level of Fgfr1 mRNA in the Fgfr1 cKOs was reduced to 20% of the control level (Figure 1C). This result showed that widespread co-recombination of the Fgfr1fx and Rosa26RfGFP loci was occurring, and we used GFP+ cells as a surrogate marker for Fgfr1+/fx cells in further experiments. However, it is important to note that Rosa and Fgfr1 co-recombination is ~80%, and we therefore underestimate the extent of the Fgfr1 phenotype using this scoring method.

Tracheae were harvested at intervals and processed for histology to assess the contribution of GFP+, Fgfr1+/fx BCs to the epithelium during normal homeostatic turnover. At 1.5 weeks after tmx administration, ~30% of total BCs were GFP+ in both Fgfr1 cKOs and controls. In control animals this percentage increased modestly to 40%–50% at 5–13 weeks before dropping to initial levels by 24 weeks. This change was much more marked in the Fgfr1 cKO mice, in which the percentage of labeled BCs peaked at 60% at 13 weeks before returning to initial levels (Figures 1D and 1E). In both genotypes, labeled BCs produced labeled luminal cells as expected. However, luminal cell production was greater in the Fgfr1 cKO mice, with levels of GFP+ luminal cells reaching 38% ± 2% of the total luminal cell population by 24 weeks, compared with 21% ± 4% in the controls (Figure 1F). Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1+/fx (Fgfr1 conditional heterozygous) mice displayed a phenotype similar to that
of the cKOs (Figures S2A and S2B). Thus, greater numbers of both basal and luminal cells are produced by the Fgfr1 Δ/Δ BCs than controls, showing that FGFR1 signaling influences the rate of epithelial cell turnover in the steady-state trachea.

Increased Levels of Basal Cell Proliferation in the Fgfr1 cKO Tracheae

We assessed proliferation rates using Ki67 antibody staining. This showed that in vivo rates of proliferation were >3-fold higher in the Fgfr1 Δ/Δ BCs than in controls (Figures 2A and 2B). Moreover, the level of Ki67 mRNA in GFP⁺ BCs isolated from Fgfr1 cKO tracheae was increased 11-fold compared with controls (Figure 2C). Proliferation levels were intermediate in the Fgfr1 conditional heterozygotes (Figure S2C). We confirmed the increase in BC proliferation in vitro using a limiting dose of Cre recombinase adenovirus (Ad-Cre) to induce recombination in small numbers of cells from Rosa26R-fGFP; Fgfr1 Δ/Δ and control Rosa26R-fGFP tracheae (Figure 2D). The Fgfr1 Δ/Δ cells produced larger GFP⁺ colonies than the controls, and these cultures also contained higher numbers of Ki67⁺ cells (Figures 2E–2G).

We predicted that if deletion of Fgfr1 results in BC proliferation, activating FGFR1 signaling in vitro would inhibit growth of BC colonies. FGF2 is expressed in the homeostatic trachea (Figure S1A), is known to activate FGFR1 preferentially in vitro, and is strongly linked to FGFR1 activation in multiple in vivo situations (Arai et al., 2015; Chakkalakal et al., 2012; Garmy-Susini et al., 2004; Hackett et al., 2011; Hadjab et al., 2013; Kranenburg et al., 2005; Ornitz et al., 1996). We therefore tested the effects of the addition of exogenous FGF2 protein on the proliferation of wild-type primary BCs in culture. We plated wild-type BCs at low density and added FGF2 on culture day 2 after plating. As predicted, addition of FGF2 had the
Figure 3. Loss of FGFR1 Signaling in BCs Leads to an Increase in Phosphorylation of Downstream Effector Proteins and a Decrease in Levels of a SPRY2 Isoform

(A) Schematic of in vitro experiment.

(B) Representative western blots from control and Fgfr1 cKO day-5 BCs showing pERK1/2, total ERK1/2, pAKT, total AKT, SPRY2, and histone H3.

(C) Quantification of protein in (B).

(D) Model. A major function of FGFR1 in BCs is to post-translationally modify SPRY2, resulting in a SPRY2 isoform which can negatively regulate the ERK/MAPK and AKT/PI3K pathways downstream of other RTKs.

(E) Confocal images of control Tg(KRT5-CreER); Rosa26RfGFP/+ and cKO Tg(KRT5-CreER); Rosa26RfGFP/+; Fgfr1<sup>D</sup>x/Fgfr1<sup>D</sup>x tracheal sections at 1.5 weeks after tmx administration showing an increase in pERK1/2 levels in the Fgfr1 cKO BCs. Green, GFP (reporter); red, pERK1/2 (active ERK1/2); blue, DAPI (nuclei). Scale bar, 25 μm.

(legend continued on next page)
opposite effect to removal of Fgfr1 and significantly reduced colony size (Figures 2I and 2J).

**Fgfr1^{−/−} BCs Have Increased Levels of MAPK and AKT Pathway Activity**

To identify the mechanism by which FGFR1 influences BC proliferation, we assessed the levels of activity of well-characterized downstream intracellular signaling pathways. Given that we had successfully replicated the proliferation phenotype in primary cell culture (Figures 2D–2F), we used primary cultures of Fgfr1^{−/−} cells with a high dose of Ad-Cre and harvested cells 3 days after infection for western analysis of the whole cell lysate (Figure 3A). Genotyping of the cultures suggested that many cells were Fgfr1^{−/−} in these conditions (Figure S3A). These experiments showed that there was a 2.2-fold increase in levels of phosphorylated ERK1/2 in the Fgfr1^{−/−} cells. In addition, there was a 3.8-fold increase in total AKT protein and a 1.3-fold increase in the fraction of phosphorylated AKT (Figures 3B and 3C). The change in MAPK pathway activity was confirmed in vivo by immunostaining for pERK1/2, which showed higher levels in Fgfr1^{−/−} BCs than controls at 1.5 weeks after tmx administration (Figure 3E). Thus, the loss of an RTK surprisingly resulted in increased levels of signaling through the RAS-ERK and PI3K intracellular pathways, which are typically associated with active RTK signaling. We therefore analyzed negative regulators of these signaling cascades. SPRY2 is a well-known transcriptional target of FGFR signaling and has previously been shown to be post-translationally modified downstream of FGFR1 activity in vitro (Lao et al., 2007; Rubin et al., 2005). Two isoforms of SPRY2 can be separated on polyacrylamide gels. Levels of the faster-migrating (lower) isoform have been shown in vitro to increase following FGFR1 activation. In addition, this isoform has a greater ability to bind to GRB2 in vitro and has been proposed to be the active isoform for inhibition of RAS-ERK signaling (Lao et al., 2007). Interestingly, in the Fgfr1^{−/−} cells there was a 2.2-fold decrease in levels of the faster-migrating (lower, active for ERK/AKT inhibition) SPRY2 isoform and a 1.3-fold increase in levels of the slower-migrating (upper, unable to inhibit ERK/AKT) isoform (Figures 3B and 3C). This suggests that a wild-type function of FGFR1 signaling in the tracheal BCs is to modulate SPRY2 activity post-translationally, thus altering the ability of SPRY2 to inhibit signaling downstream of other RTKs (Figure 3D).

There is a strong connection between EGFR signaling and airway BC proliferation (Brechbuhl et al., 2014). We tested the hypothesis that because the Fgfr1 cKO BCs have more constitutive activity of the ERK/AKT pathways, they would be more resistant to EGFR inhibition than controls. We grew Fgfr1 cKO and control BCs in the presence of 0.25 μM of the EGFR inhibitor erlotinib (Figure S4A). This resulted in a >5-fold decrease in proliferation rates in the control cells, versus a 2-fold decrease in the Fgfr1 cKO cells (Figures S4B–S4D). Moreover, levels of EGFR protein were unchanged in the Fgfr1 cKO cells (Figure S4E). The Fgfr1 cKO BCs are thus more resistant to EGFR inhibition, supporting the hypothesis that the downstream signaling pathways (ERK/PI3K) are more constitutively active in these cells.

**Stimulating BCs with FGF2 Results in Increased Levels of the Faster-Migrating SPRY2 Isoform**

We tested whether activating FGF signaling is sufficient to modulate levels of the SPRY2 isoforms in wild-type cells (Figure 3F). A 10-min or 24-hr FGF2 stimulation resulted in a reproducible small decrease in pERK1/2 compared with control cells, particularly at the 10-min harvest time (Figures 3G, 3H, and S5A–S5C). Following FGF2 stimulation we observed no change in total SPRY2 levels but a 1.6-fold increase in levels of the faster-migrating SPRY2 isoform (Figures 3G and 3H). These changes never reached statistical significance. However, the experiments were performed in primary cells that have multiple inputs into the ERK pathway in the presence of serum and the data were reproducible, supporting our hypothesis that manipulating FGFR1 signaling results in post-translational changes in SPRY2.

**Fgfr1^{−/−} BCs Cannot Produce Ciliated Cells**

It has previously been reported that FGFR1 signaling in zebrafish and Xenopus embryos can regulate cilia assembly and length (Hong and Dawid, 2009; Neugebauer et al., 2009). We therefore examined ciliogenesis in the Fgfr1 cKO tracheae. At 24 weeks after tmx administration, Fgfr1 cKO tracheae had an excess of GFP+ secretory cells and fewer GFP+ ciliated cells than the controls (Figures 4A and 4B), suggesting that ciliated cell production was impaired in the mutant. Indeed, the only GFP+ ciliated cells that could be detected in the Fgfr1 cKO trachea still retained FGFR1 protein (Figures 4A and 4C), indicating that they were derived from Fgfr1^{−/−} cells, which had undergone partial recombination to activate the Rosa allele. Based on scoring the percentage of luminal GFP+ cells that retained detectable FGFR1 protein in these samples, we were able to estimate the rate of co-recombination of the two floxed alleles as 84% ± 2% (Figure 4C), in good agreement with our qRT-PCR-based estimate (Figure 1C).

To assess the stage at which ciliated cell differentiation was blocked in the Fgfr1 cKO cells, we stained tracheal sections for the transcription factor FOXJ1, which is expressed during ciliated cell differentiation and has a role in basal body anchoring (Blatt et al., 1999; Huang et al., 2003). In the Fgfr1 cKO tracheae the number of GFP+, FOXJ1+ cells was significantly decreased, confirming an early block in ciliated cell differentiation (Figures 4B and 4D). These data suggest that in the Fgfr1 cKO trachea, luminal cell fate choice is disrupted and secretory cells are produced at the expense of ciliated cells. This phenotype is somewhat different to the published role for FGFR1 in controlling cilia length in multi-ciliated cells in the...
zebrafish and *Xenopus* systems (Hong and Dawid, 2009; Neugebauer et al., 2009). This may be a true cross-species difference or may reflect the limitations of the different experimental systems used.

Homeostatic production of new ciliated cells occurs at an extremely slow rate in vivo (Rawlins and Hogan, 2008). To confirm the ciliated cell differentiation phenotype in the *Fgfr1<sup>D/D</sup>* cells, we grew air-liquid interface (ALI) epithelial cultures from *Rosa26R-fGFP; Fgfr1<sup>D/D</sup>* and control *Rosa26R-fGFP; Fgfr1<sup>+/fx</sup>*

![Figure 4](image)

**Figure 4. Loss of FGFR1 Signaling Results in a Block in Ciliated Cell Differentiation**

(A) Confocal image of *Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1<sup>D/D</sup>* tracheal section. Green, GFP (reporter); blue, FGFR1; red, SCGB1A1 (secretory cells); white, acetylated tubulin (ACT, cilia). In the patch of GFP<sup>+</sup> cells marked by the dashed bracket, FGFR1 is absent and there are no ciliated cells. In the GFP<sup>+</sup> cells within the solid bracket, FGFR1 has not been deleted, there are fewer GFP<sup>+</sup> cells, and ciliated cell differentiation has occurred.

(B) Percentage of GFP<sup>+</sup> cells that co-express SCGB1A1, ACT, or FOXJ1 in control *Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1<sup>+/fx</sup>* and cKO *Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1<sup>D/D</sup>* animals 24 weeks after tmx administration.

(C) Percentage of GFP<sup>+</sup> luminal cells in cKO *Tg(KRT5-CreER); Rosa26R-fGFP; Fgfr1<sup>D/D</sup>* trachea that retain FGFR1 protein 24 weeks after tmx administration.

(D) Sections from control and *Fgfr1 cKO* tracheae at 24 weeks after tmx administration. Green, GFP (reporter); red, γ-tubulin (basal bodies); white, FOXJ1 (ciliated cells).

(E) Schematic of in vitro experiment.

(F) Day-11 ALI cultures grown from control and *Fgfr1 cKO* animals. Green, GFP (reporter); red, ACT (cilia); blue, DAPI (nuclei).

Error bars denote SEM. Scale bars represent 100 μm in (A) and (F), and 50 μm in (D).

tracheae and deleted the floxed alleles using *Ad-Cre* (Figure 4E). All control cultures had robust differentiation of ciliated cells, based on acetylated tubulin (ACT) localization, but ciliated cells were not observed in the *Fgfr1 cKO* cultures (Figure 4F).

In summary, the *Fgfr1 cKO* BCs have an increased proliferation rate from 1.5 weeks after injection. They are also unable to produce mature ciliated cells, likely due to a fate-choice defect. We focused on BC proliferation as the primary phenotype and, based on our observation that levels of the fast-migrating SPRY2 isoform were decreased in the *Fgfr1 cKO* cells (Figures 3B and 3C), we hypothesized that a major role of FGFR1 signaling in the tracheal BCs is to modulate SPRY2 activity and, moreover, that the function of SPRY2 in these cells is to inhibit signaling activity downstream of RTKs other than FGFR1, for example EGFR, which is necessary for tracheal BC proliferation (Brechbuhl et al., 2014; Lu et al., 2013). To test this hypothesis, we deleted *Spry2* in BCs of the adult mouse trachea.

**Loss of Spry2 Results in Increased Levels of Tracheal Basal Cell Proliferation**

*Spry2* is widely expressed in basal and luminal cells of the homeostatic mouse trachea (Figures 1A and S1). We generated
Tg(KRT5-CreER); Rosa26R-fGFP, Spry2<sup>Dfx</sup> (Spry2 cKO) and control Tg(KRT5-CreER); Rosa26R-fGFP adult mice and administered tmx to activate GFP and delete Spry2 in a subset of BCs (Figure 5A). During normal turnover there was an increase in the proportion of GFP<sup>+</sup>, Spry2<sup>D/D</sup> BCs compared with GFP<sup>+</sup>, Spry2<sup>+/+</sup> controls (Figures 5B and 5C). At the same time the proportion of GFP<sup>+</sup>, Spry2<sup>D/D</sup> luminal cells also increased (Figures 5B and 5D). These changes most likely resulted from the ~4-fold increase in rates of BC proliferation observed in the Spry2 cKOs compared with controls (Figures 5E and 5F), of epithelial stratification, and levels of E-cadherin protein (Figure 5G) and apical-basal polarity (Figure S6C) appeared normal. These data suggest that the observed change in cell density may result from an altered threshold for contact inhibition in the Spry2 cKO cells.

We tested whether we could recapitulate the Spry2 cKO phenotypes using Ad-Cre in primary tracheal cell culture (Figure 6A). Deletion with a high titer of Ad-Cre resulted in loss of all detectable SPRY2 protein (Figure 6E) and failure to amplify the Spry2<sup>fx</sup> allele from the genomic DNA (Figure S3B), suggesting that we

---

<table>
<thead>
<tr>
<th>Tmx administration (weeks)</th>
<th>Harvest (weeks)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>1.5</td>
<td>5</td>
</tr>
<tr>
<td>13</td>
<td></td>
</tr>
</tbody>
</table>

---

Figure 5. Deletion of Spry2 in BCs Results in Increased Rates of Proliferation

(A) Schematic of Spry2 conditional knockout experiment.

(B) Representative confocal sections from control Tg(KRT5-CreER); Rosa26R-fGFP and cKO Tg(KRT5-CreER); Rosa26R-fGFP; Spry2<sup>Dfx</sup> tracheae. Green, GFP (reporter); red, KRT5 (BCs); blue, DAPI (nuclei). Brackets indicate patches of labeled cells, more prominent in the Spry2 cKO.

(C) Percentage of the total T1<sup>α</sup> BCs that are also GFP<sup>+</sup>.

(D) Percentage of the total T1<sup>α</sup> luminal cells that are also GFP<sup>+</sup>.

(E) Representative confocal sections from control and cKO tracheae at 1.5 and 13 weeks after tmx administration. Green, GFP (reporter); red, Ki67 (proliferating cells); white, KRT5 (BCs); blue, DAPI (nuclei). Arrowheads indicate Ki67<sup>+</sup> cells.

(F) Percentage of GFP<sup>+</sup> cells that co-express Ki67.

(G) Confocal z projections from sections of control and Spry2 cKO trachea. Green, GFP (reporter); red, KI67 (proliferating cells); white, E-cadherin (lateral cell membranes); blue, DAPI (nuclei).

Error bars denote SEM. Scale bars represent 50 μm in (B) and (E), and 10 μm in (G). See also Figure S6.

The proliferation phenotypes were thus very similar between the Fgfr1 and Spry2 cKO tracheae, supporting a strong connection between the two proteins. However, unlike the Fgfr1 cKO, no effects on ciliated cell differentiation were observed in the Spry2 cKO tracheae (Figures S6A and S6B), suggesting that the function of FGFR1 in luminal fate choice is SPRY2 independent. In addition, in contrast to Fgfr1, no phenotypes were observed in Spry2 conditional heterozygous tracheae (data not shown).

The Spry2 cKO cells were packed more tightly than control cells in the epithelium (at 13 weeks after tmx administration there were 16 ± 1.3 cells per 40 μm in the control tracheal epithelia versus 23.7 ± 1.6 cells per 40 μm in the Spry2 cKOs; p = 0.01; Figure 5B), consistent with the significantly increased proliferation rate. However, there was no evidence
Figure 6. Rapidly Proliferating Spry2 Conditional Knockout BCs Have Elevated Levels of RAS-ERK and PI3K Signaling

(A) Schematic of in vitro experiment.

(B) Spry2 cKO (Rosa26R<sup>Cre<sup>ER</sup></sup><sup>T2</sup>; Spry2<sup>Dfx</sup>) cells proliferate more rapidly than controls (Rosa26R<sup>GFP</sup>/GFP) in vitro and are more densely packed at culture day 5. Note that not all cKO cells are GFP<sup>+</sup>, even though SPRY2 protein is undetectable. Green, GFP (reporter); red, KRT5 (BCs); blue, DAPI (nuclei). Arrowheads indicate similar regions of the cultures.

(C) Spry2 cKO cells still undergo some contact inhibition in vitro at day 7 after ALI induction. Their proliferation rate slows following induction of ALI and there are no regions of multi-layering, although KI67<sup>+</sup> cells are still visible (arrowhead). Green, GFP (reporter); red, KI67 (proliferating cells); blue, DAPI (nuclei).

(D) Spry2 cKO primary cells continue to proliferate faster in vitro following passage at day 2 after replating. Green, GFP (reporter); red, KI67 (proliferating cells, e.g. arrowhead); blue, DAPI (nuclei).

(E) Representative western blots from control and Spry2 cKO day-5 BCs showing SPRY2, pAKT, total AKT, pERK1/2, total ERK1/2, SOX2, and histone H3.

(F) Quantification of protein levels. Error bars denote SEM.

(G) Confocal images of control Tg(KRT5-CreER); Rosa26R<sup>Cre<sup>ER</sup></sup><sup>T2</sup>; Spry2<sup>Dfx</sup> and cKO Tg(KRT5-CreER); Rosa26R<sup>Cre<sup>ER</sup></sup><sup>T2</sup>; Spry2<sup>Dfx</sup> tracheal sections at 1.5 weeks after tmx administration showing an increase in pERK1/2 levels in the Spry2 cKO BCs. Green, GFP (reporter); red, pERK1/2 (active ERK1/2); blue, DAPI (nuclei).

Scale bars, 25 μm. See also Figure S3.
achieved complete recombination of the Spry2Δ/Δ allele. In spite of this there were large patches of GFP− cells in our cultures that had not recombined the Rosa reporter (Figure 6B). This indicates that the Spry2Δ/Δ allele recombines more efficiently than the Rosa26R-fGFP reporter and supports the conclusion that most of the GFP+ cells observed in vivo were co-recombinants with Spry2Δ/Δ. Therefore, it is likely that scoring GFP+ cells in the Spry2 cKO tracheae results in a good approximation to the Spry2Δ/Δ phenotype (compare with the Fgr1 cKO tracheae, where scoring GFP+ cells results in an underestimate of the Fgr1Δ/Δ phenotype).

The Spry2Δ/Δ cultures had a greater rate of proliferation than the controls and also reached a higher cell density (Figures 6B and 6C). Proliferation of the Spry2Δ/Δ cells slowed after the cells reached confluence, but did continue (Figure 6D). Moreover, confluent cultures could easily be passaged and continued to show a faster proliferation rate (Figure 6D). Importantly, although the Spry2Δ/Δ cells reached a greater density than controls, we never observed phenotypes consistent with a complete loss of contact inhibition, such as multi-layering or cell detachment. This is again consistent with an altered threshold for, but not complete loss of, contact inhibition.

Similar to the Fgr1Δ/Δ cells, the Spry2Δ/Δ cells had a 3.3-fold increase in pERK1/2 and a 5.2-fold increase in pAKT levels compared with the controls (Figures 6E and 6F). These changes were highly reproducible, although not always statistically significant. The increase in MAPK pathway activity was confirmed in vivo by immunostaining for pERK1/2, which showed higher levels in Spry2Δ/Δ BCs than controls at 1.5 weeks after tmx administration (Figure 6G). We also observed that the Spry2Δ/Δ cells had a 3.3-fold increase in levels of EGFR protein (Figures 6E and 6F). This is consistent with the published function of the slower-migrating SPRY2 isoform (which is absent in the Spry2Δ/Δ cells, but is unaffected in the Fgr1Δ/Δ cells) in EGFR trafficking/degradation (Gao et al., 2012; Kim et al., 2007; Walsh and Lazzara, 2013).

**DISCUSSION**

The tracheal epithelium is normally quiescent, with a minimal amount of cell turnover. By combining in vivo genetic analysis and primary cell culture, we have shown that BC proliferation is actively repressed under steady-state conditions by an FGFR1-SPRY2 signaling axis that functions to inhibit signaling through the ERK and AKT intracellular pathways. This result argues that airway quiescence does not result simply from the absence of a proliferative signal or from feedback inhibitory loops. Rather, we propose a model in which quiescence is maintained by a balance between signaling mechanisms that either promote proliferation or repress it. This is an emerging theme in the stem cell biology of quiescent organs. For example, a recent publication has found a similar tonic Hedgehog signal that inhibits steady-state proliferation in the smaller mouse airways (Peng et al., 2015), but it is not known whether this Hedgehog-based mechanism applies in the BC-containing trachea.

SPRY2 was previously characterized in vitro as a protein with two isoforms, which can either inhibit ERK/AKT signaling (faster-migrating SPRY2 isoform) or modulate the subcellular trafficking/degradation of EGFR protein (slower-migrating isoform) (Guy et al., 2009). However, the relative roles of these SPRY2 isoforms in vivo and how they function downstream of RTK receptors have not previously been explored. Several lines of evidence presented here support a model in which the faster-migrating SPRY2 functions downstream of FGFR1 to control cell proliferation and tracheal epithelial homeostasis (Figure 7A). First, following the loss of FGFR1 protein in airway BCs we observe reduced levels of the faster-migrating (ERK/AKT-inhibiting) SPRY2 isoform, as well as increased levels of pERK and pAKT, increased rates of proliferation, and increased resistance to EGFR inhibition. By contrast, the slower-migrating SPRY2 isoform is unaffected (Figure 7B). Second, exposing wild-type BCs to FGF2 ligand results in increased levels of...
faster-migrating SPRY2, a decrease in dpERK, and inhibition of BC proliferation. Third, loss of all SPRY2 results in increased ERK/AKT signaling and BC proliferation (Figure 7C). Taken together, our data suggest that active FGFR1 signaling in the steady-state trachea maintains a pool of SPRY2 protein, which functions to antagonize ERK/AKT signaling and limit cell proliferation downstream of other RTKs, most likely EGFR (Figure 7A).

A relationship between FGFR1 and post-translational modification of SPRY2 protein has previously been documented in cell lines in vitro (Lao et al., 2007; Lao et al., 2009). We now demonstrate its importance in vivo, clearly illustrating that the context in which a cell interprets EGFR/RTK signaling can be highly dependent on the activity of other RTKs present. This has clear implications for cancer development and the evolution of drug-resistant tumors.

Another important finding of our work is that FGFR1 and SPRY2 can also function independently in airway epithelial homeostasis. FGFR1 is required for multi-ciliated cell differentiation (Figure 4). Moreover, our data suggest that secretory cells are produced at the expense of ciliated cells by the Fgfr1cko BCs. A role for FGFR1 in ciliated cell differentiation is also consistent with previous reports from other model organisms (Hong and Dawid, 2009; Neugebauer et al., 2009), but the details of the cellular requirements remain to be worked out. SPRY2 regulates EGFR protein levels in BCs (Figure 6). Similar observations have previously been made in vitro or in cancer models, but we demonstrate this SPRY2-EGFR relationship for the first time in a homeostatic tissue (Gao et al., 2012; Kim et al., 2007; Walsh and Lazzara, 2013). SPRY2 thus emerges as a highly flexible switch for cellular homeostasis by SPRY2.

A key question raised by our model concerns what activates FGFR1 in vivo. We observed that FG2 is transcribed in the tracheal mesenchyme in vivo at steady state, and that recombinant FG2 inhibits tracheal BC proliferation and modulates SPRY2 in vitro. These data are consistent with a role for FG2 in activating FGFR1 signaling in vivo, although we note that other FG ligands are expressed in the steady-state airways. Interestingly, in a recent screen of secreted proteins for their effect on BCs in vitro (DaSilva et al., 2006; Edwin et al., 2009; Lao et al., 2007), these data suggest that SPRY2 is extensively and flexibly modified in many different cellular settings in vivo and that our observations may be the tip of the iceberg for the control of cellular homeostasis by SPRY2.

In conclusion, airway epithelial quiescence requires active inhibition of proliferation by FGFR1 signaling, which results in the post-translational modification of SPRY2 and inhibition of signaling downstream of other RTKs, partly EGFR. We have thus demonstrated a specific in vivo function for an FGFR1-SPRY2 signaling axis that was previously documented in vitro. This illustrates the complexity of RTK signaling networks and the importance of further investigation in both homeostatic tissues and developing malignancies.

**EXPERIMENTAL PROCEDURES**

See Supplemental Experimental Procedures for a full description of all materials and methods used.

**Mice**

Experiments were performed under Home Office licenses PPL80/2326 and 70/812. Fgf10loxP (Xu et al., 2002), Tg(KRT5-CreER) (Rock et al., 2009), Rosa26R-GFP (Rawlins et al., 2009), Spry2loxP (Shim et al., 2003) were described previously. Fgf10loxP and Spry2loxP were generated by crossing floxed alleles to Zp3-Cre (de Vries et al., 2009). All strains were maintained on a C57Bl/6J background. Males and females >8 weeks old were used. Wild-type mice were C57Bl/6J.

**Tamoxifen**

Animals were injected intraperitoneally with tmx, 0.2 mg/g body weight, four times every other day.

**Tracheal Epithelial Cell Culture**

Tracheae were cut into small pieces and incubated in 50% Dispase II (Gibco, 16 U/ml). Epithelial sheets were isolated and dissociated to single cells. Unless otherwise stated, 5 × 10^5 cells in 0.5 ml of MTEC/+ medium (You et al., 2003) were plated on collagen-coated 12-well tissue culture inserts (BD Falcon, 353180). Differentiation was induced in confluent cultures by removal of insert medium and addition of MTEC serum-free medium to the outer chamber. Ad-Cre (University of Iowa, Gene Transfer Vector Core) M01 was used for scattered recombination and M01 2,500 for complete recombination; 1 × 10^5 vector plaque-forming units, 100 ng/ml recombinant mouse FG2, and 0.25 μM erlotinib (Sigma).

**Immunostaining**

Cryosections (6 μm) or whole-mount primary cultures were stained with ACT (mouse, 1:3,000, Sigma, T7451), E-cadherin (rat, 1:3,000, Invitrogen, 13-1900), pERK1/2 (rabbit, 1:200, Cell Signaling Technology, 4695), FGFR1 (rabbit, 1:200, Novus Biologicals, NBP1-20067), FOXJ1 (mouse, 1:200, eBioscience, clone 2A5, 14-9965-80), GFP (chick, 1:1,000, Abbac, Ab13970), KRT5 (rabbit, 1:500, Covance, PRB-160P), Ki67 (mouse, 1:200, Becton Dickinson, 550609), SCGB1A1 (goat, 1:400, Santa Cruz Biotechnology, sc9772), SPRY2 (rabbit, 1:200, Abbac, Ab50317), and T14 (1:1,000, DSHB, 8.1.1). Antigen retrieval was in 10 mM boiling sodium citrate for Ki67 (pH 8), pERK1/2 (pH 6.5), and FOXJ1 (pH 6). Conjugated secondary antibodies were Phalloidin-568 and Alexa Fluor (Life Technologies, 1:2,000).
mRNA In Situ Hybridization
Cryosections (10 μm) were hybridized with DIG-labeled anti-sense mRNA probes to Fgfr1 (EST: AW495528) or Spry2 (Minowada et al., 1999) (DIG RNA labeling kit, Roche, 11175025910), and visualized using alkaline phosphatase-coupled anti-DIG (1:2,000, Roche, 129300223) and BCIP/NBT (Roche, 1382313).

Microscopy and Image Scoring
Slides were imaged on a Zeiss Axioslager compound microscope or a Leica SP8/SP5 confocal microscope as stated. Cells were scored manually in Fiji (Imagej), or using the Gurdon Institute Imaging Facility’s plugin ObjectScan. For cryosections, every epithelial cell along the entire proximal to distal length of a longitudinal section from the center of the trachea was scored. BCs were T1+ or KRT5+ with the apical surface not reaching the lumen. For cultured cells, at least three random fields of view per insert were scored.

RT-PCR
Samples were enriched for epithelium or mesenchyme using Dispose digestion and manual peeling. Total RNA was extracted using TRIzol reagent.

qRT-PCR
Primary tracheal epithelial cells were isolated and sorted using a fluorescence-activated cell sorting MoFlo flow cytometer. Wild-type cells were gated as basal EpCAM, GSI4 lectin; secretory EpCAM, SSEA1; and ciliated EpCAM, CD24. GFP+ BCs from control and Fgfr1 cKO tracheae were sorted as GFP+, GSI4 lectin. Total RNA was extracted using a Qiagen RNeasy Mini Kit, and Taqman gene expression assays (Life Technologies) were used.

Immunoblots
Cells were collected in Cell Extraction Buffer (Invitrogen, FNN0011) with protease inhibitor (Roche, 04693116010) and PMSF (Sigma, P7626). Proteins were separated on 10% or 12% SDS-PAGE gels before transfer to Millipore membranes (Millipore, 04-290, 1:300). Detection was performed with horseradish peroxidase-coupled anti-DIG (1:2,000, Roche, 129300223) and BCIP/NBT (Roche, 1382313). Total RNA was extracted using a Qiagen RNeasy Mini Kit, and Taqman gene expression assays (Life Technologies) were used.

Statistics
Statistics were calculated using Student’s two-tailed t test with unequal variance.

SUPPLEMENTAL INFORMATION
Supplemental Information includes Supplemental Experimental Procedures and six figures and can be found with this article online at http://dx.doi.org/10.1016/j.devcel.2016.03.001.

AUTHOR CONTRIBUTIONS
G.B. designed and performed experiments, analyzed data, and edited the manuscript. J.J. performed experiments. M.A.B. provided mice and trachea samples and assisted with experimental design. E.R. conceived and led the project, performed experiments, analyzed data, and wrote and edited the manuscript.

ACKNOWLEDGMENTS
This study was supported by the Medical Research Council (G0900424 to E.R.) and a Wellcome Trust clinical PhD fellowship (J.J.). Core grants: Gurdon Institute: Wellcome Trust (092096), Cancer Research UK (C6946/A14492); Stem Cell Initiative: Wellcome Trust/MRC.

REFERENCES


Developmental Cell 37, 85–97, April 4, 2016 95


